**Reviewer’s Comments**

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**Prevalence of *Pseudomonas aeruginosa (P. aeruginosa)* and Antimicrobial Susceptibility Patterns at a Private Hospital in Sana'a, Yemen**

**Abstract**

**Background:***Pseudomonasaeruginosa* is clinically significant and opportunistic pathogenthat causes infections in hospitalized patients. Antibiotic resistance is a major concern in clinical practice. The ongoing emergence of resistant strains that cause nosocomial infections contributes substantially to the morbidity and mortality of hospitalized patients.

**Objective:**To estimate the prevalence of *Pseudomonasaeruginosa*and antimicrobial-resistant *P. aeruginosa*and the antimicrobial resistance patterns of *P. aeruginosa*clinical isolates from hospitalized patients.

**Methods:** The study was performed at microbiology department of local hospital in Sana’a, Yemen. All the patients' samples of hospital departments from January, 2017 to December, 2017 were included. A Total of 2079 samples were collected during the study period. Among them, 193 strains of *Pseudomonas spp.* were isolated.

**Results:** One hundred ninty three isolates of *P. aeruginosa*were isolated from different clinical specimens and fully characterized by standard bacteriological procedures. Antimicrobial susceptibility pattern of each isolates was carried out by the Kirby-Bauer disk diffusion method as per CLSI guidelines. Majority of *P. aeruginosa*were isolated from Sputum, followed by urine specimens. The isolate pathogen shows the highestsensitive to Meropenem (85.5%), followed by Amikacin (80.5%), Imipenem (80.0%), and Piperacillin/tazobactam (77.2). The highest frequency of resistance (96.2%) was observed with amoxicillin /clavulinic Acidfollowed by cefuroxime 94.6%, ampicillin/ sulbactam94.5%, Co-Trimoxzole 80.5%, andnorfloxacin 54%.

**Conclusion**: The result confirmed the occurrence of drug resistance strains of *P.aeruginosa*. Meropenem, imipenem, and amikacin, were found to be the most effective antimicrobial drugs. It therefore calls for a very judicious, appropriatetreatmentregimens selection by the physicians to limit the further spread of antimicrobial resistance *P. aeruginosa*.

***Key words****: Pseudomonas aeruginosa, Antimicrobial susceptibility, Multidrug-resistant, Imipenem*

1. **Introduction:**

*Pseudomonasaeruginosa*is clinically significant and opportunistic pathogenthat causes infections in hospitalized patients. In addition, most *Pseudomonas species*have intrinsic resistance to many antibiotics and ongoing emergence of new resistance can be developedafter commonly prescribed antimicrobial agents**(1)**.*Pseudomonas aeruginosa*has naturally resistant to many antibiotics due to thepermeability barrier afforded by its outer membrane lipopolysaccharide (LPS). Only few antibiotics are effective against *Pseudomonas*and even these antibiotics are not effective against all strains **(2)**.Antibiotic resistance is a major concern in clinical practice. The resistant strains of *Pseudomonas aeruginosa*that cause nosocomial infections contributes substantially to the morbidity and mortality of hospitalized patients**(3)**. Despite the availability of a variety of effectiveantimicrobial agents, treatment of *pseudomonalaeruginosa* is often challenging**(4)**. Antimicrobial resistance is a growing problem worldwide, especially in hospitals, where resistant organisms are often first detected in ICUs**(5)**. The organism had been isolated from various infections like respiratory tract infections, cystic fibrosis, ear infections, orthopaedic infections, urinary tract infections, surgical infections, severe burns, etc. It was also reported frequently from patients undergoing chemotherapy for neoplastic diseases**(6)**.The variations of antibiotic protocols in clinics or in regions result in the different resistance profiles**(4)**. It is, therefore, the goal of this study to determine the prevalence of *P.aeruginosa*isolates in a private hospital in Sana'a, Yemenalso to evaluate its susceptibility against certain antibiotics, as limited work has been previously conducted on this subject.

**2. Method:**

The study was performed at university of science and technology hospitalin Sana’a, Yemen. It is one of the major private hospitals in Yemen. All the patients' samples from January,2017 to December, 2017 were included. A Total of 2079 samples were gatheredduring the study period. Among them, 193 strains of *Pseudomonas aeruginosa*were isolated.The medicalrecords of these patients were retrieved and reviewed. All information regarding patients' gender andage as well as origin of clinical samples were collected.

Antimicrobial susceptibility testing of all the *Pseudomonas aeruginosa*isolates was performed by Kirby-Bauer disk diffusion method and the result were interpreted by the Clinical Laboratory Standard Institute (CLSI) guidelines**(7)**.

The antimicrobial susceptibility patterns of all the Pseudomonas *aeruginosa*strains were determined against the following antibiotics of standard strength:ceftazidime, amikacin, gentamicin, imipenem, meropenem, ciprofloxacin, cefoperazone, piperacillin/tazobactam,amoxicillin / clavulinic acid,moxifloxacin,cefepime,ceftizoxime,ampicillin/ sulbactam,cefuroxime, ceftriaxone,Co-Trimoxzole, and levofloxacin.Full ethical clearance was obtained from the qualified authorities who approved the study design.All data were analyzed using SPSS Statistics 21. Data was presented in tables and graphs.

**3. Result:**

According to result findings, there were more than half of Pseudomonas*aeruginosa* isolates in age group of 60 years and greater with 55(28%), followed by the age between 46 to 60 years in second rank about 38(20%), and finally the age between 31 to 45 years only about 20(10%). In this study, Overall MRSA prevalence was 9.3 % (n=193/2079).

**Figure 1. Distribution of *Pseudomonas aeruginosa i*solates according to age.**

The figure 2 showed that there were about 154(80%) of *Pseudomonas isolates* form male, whereas the female had only about 39(20%).

**Figure 2.Distribution of *Pseudomonasaeruginosa* according to gender.**

According to the study results, the medical department had the highest prevalence of *Pseudomonasaeruginosa* isolates about 48(25%), followed by the intensive care unit in second rank about 41 (21%), the surgical department in third rank about 37(19%) and finally the pediatric and gynecology departments had only about 16(8%).

**Figure 3. Distribution of *Pseudomonasaeruginosa*isolates according to hospital departments.**

The figure 4 showed that the most of sample tests from sputum culture about 82(42.5%), followed by the sample from urine culture in second rank about 34(17.6%), and finally the sample test from other rout only about 6(3.1%).

**Figure 4. Distribution of *P.aeruginosa*isolates according to sample types.**

According to the current study findings (table 1),more than half of medication was sensitive to *P.aeruginosa*test about 12 drugs (54.5%), whereas the medication that resistance to *pseudomonas* tests about 10 drugs (45.5%).

*Pseudomonasaeruginosa* strains showed resistance to ciprofloxacin 50.89%, ceftazidime 31.5%, ceftriaxone 78%, amoxicillin /clavulinic Acid 96.2%,ampicillin/ sulbactam 94.5%, cefuroxime 94.6%, nalidixic acid 83%, nitrofurantoin 88%, doxycycline 82.6%,norfloxacin 54%,and Co-Trimoxzole 80.5%. The highest frequency of sensitivity (85.5%) was observed with meropenem followed by amikacin 80.5%, imipenem 80%, piperacilline/tazobactam 77.2%, ceftizoxime 75%, ciprofloxacin 71.5%, levofloxacin 66%, cefoperazone 64%, gentamicin 56%, ceftazidime 54.5%, moxifloxacin 49%, and cefepime 44.5%.

**Table 1. Antimicrobial susceptibility patterns for *P.aeruginosa*isolates**

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Antibiotics** | **Expected options** | **Response** | **Antibiotics** | **Expected options** | **Response** |
| **F** | **%** | **F** | **%** |
| **Ceftriaxone** | **S** | **11** | **18.5%** | **Ceftazidime** | **S** | **103** | **54.5%** |
| **R** | **46** | **78%** | **R** | **60** | **31.5%** |
| **I** | **2** | **3.5%** | **I** | **26** | **14%** |
| **Cefoperazone/ sulbactam** | **S** | **58** | **64%** | **Ciprofloxacin** | **S** | **118** | **71.5%** |
| **R** | **27** | **29.5%** | **R** | **40** | **24%** |
| **I** | **6** | **6.5%** | **I** | **7** | **4.5%** |
| **Levofloxacin** | **S** | **108** | **66%** | **Co-Trimoxzole** | **S** | **37** | **19.5%** |
| **R** | **44** | **26.8%** | **R** | **152** | **80.5%** |
| **I** | **12** | **7.2%** | **I** | **0** | **0.0%** |
| **ampicillin/ sulbactam** | **S** | **2** | **3.7%** | **Imipenem** | **S** | **150** | **80%** |
| **R** | **51** | **94.5%** | **R** | **29** | **15.4%** |
| **I** | **1** | **1.8%** | **I** | **9** | **4.6%** |
| **Amoxicillin / Clavulinic Acid** | **S** | **4** | **2.1%** | **Norfloxacin** | **S** | **10** | **38.5%** |
| **R** | **179** | **96.2%** | **R** | **14** | **54%** |
| **I** | **3** | **1.7%** | **I** | **2** | **8%** |
| **Amikacin** | **S** | **152** | **80.5%** | **Cefepime** | **S** | **83** | **44.5%** |
| **R** | **28** | **14.8%** | **R** | **81** | **43.5%** |
| **I** | **9** | **4.7%** | **I** | **22** | **12%** |
| **Gentamicin** | **S** | **105** | **56%** | **Meropenem** | **S** | **89** | **85.5%** |
| **R** | **65** | **35%** | **R** | **10** | **9.5%** |
| **I** | **17** | **9%** | **I** | **5** | **5%** |
| **Moxifloxacin** | **S** | **77** | **49%** | **Piperacillin/****tazobactam** | **S** | **146** | **77.2%** |
| **R** | **69** | **44%** | **R** | **31** | **16.5%** |
| **I** | **11** | **7%** | **I** | **12** | **6.3%** |
| **Cefuroxime** | **S** | **8** | **4.2%** | **ceftizoxime** | **S** | **1** | **4%** |
| **R** | **178** | **94.6%** | **R** | **18** | **75%** |
| **I** | **2** | **1.2%** | **I** | **5** | **21%** |

According to figure 5 below, the highest resistance rate of anti-pseudomonal agent was with cefepime about 43.5% and the lowest resistance rate with imipenem.Resistance to antipseudomonal drugs in our study was found to be cefepime (43.5%), ceftazidime (31.5%), ciprofloxacin (24%), piperacillin /tazobactam (16.5%), imipenim (15.4%).In the present study, multidrug resistance (MDR) rate (resistance to three or more of anti-*Pseudomonal* antimicrobials (i.e. piperacillin + tazobactam, imipenem, ceftazidime and amikacin) was determined to be 4.2% (8/193). Also MDR rate for only three anti Pseudomonal antimicrobials without imipenem was 4.2% (n= 8/193) (i.e. piperacillin + tazobactam, ceftazidime and amikacin)

**Figure 5. Resistance rates of anti-*pseudomonal* agents.**

**4. Discussion:**

*Pseudomonasaeruginosa* has defined as one of the most common nosocomial pathogens. Hence we have undertaken this study to analyse the prevalence and antimicrobial susceptibility pattern of *pseudomonas aeruginosa* from various clinical samples of a private hospital.Periodic antimicrobial resistance monitoring in *P. aeruginosa*is fundamental to updating the current activity level of commonly used antipseudomonal drugs.The present study measures the rate of isolation of *pseudomonas aeruginosa*(n=193/2079; 9.3%) as which is lower than previous studies as by Tadvi et al. **(8)** (22.67%) and Viren et al **(9)**.

The occurrence of *P.aeruginosa*was found to be higher in males, inpatients in age group >60 years and in surgery department, which is same as reported by Marzoqi et al. **(10)**. This might be due to prolonged hospitalization and other associated co-morbidities in these age groups.The distribution of *pseudomonasaeruginosa* isolates specimens may vary with each hospital as each hospital and each health facility has a different environment associated with it. According to the study results, more than 42.5% of the pseudomonas *aeruginosa*isolates were obtained from sputum samples.

The distribution rank of the isolates according to the types of specimens was (respiratory sputum > urine > blood > pus >wound swap > others). Respiratory isolates (42.5%) were the most frequently encountered. *P. aeruginosa*isolates from respiratory tract as observed in a similar study of inpatient isolates done in a Saudi Arabian hospital**(11)**. In the present study, the maximum clinical isolates of *P. aeruginosa*were isolated from medical department (25%), followed by ICU (21%) & surgical department (19%). This wassimilar to study of Pathmanathan SG**(12)**. The distribution of specimens of *Pseudomonas aeruginosa*might vary with each hospital as each hospital facility has a different environment associated with it.The correlation between specimen type and multidrug resistance would have been more noteworthy if supported by data on patients’ clinical conditions.Prevalence of infection was higher in medical ward followed by ICU as maximum isolates were isolated from sputum samples. According to vancomycin and linezolide usage there was no significant correlation between drug resistance and the wards from which isolates originated.

However, there was statistical significant relationship between the piperacilline/tazobactam susceptibility and sample types (P value= 0.04). On other hand, there was no statistical significant relationship between the other antibiotics susceptibility (ceftazidime, imipenem, cefipeme) and sample types.As with this study, *P. aeruginosa* infection was primarily noted among older adults (n = 55, 28%) particularly respiratory infection (n = 82, 42.5%). There are a number of reasons why older adults are burdened by this type of infection. These include age-associated impairments in immunity that lead to reduced response to vaccination, a constellation of chronic and comorbid diseases, and functional limitations associated with advanced age. Additionally, older adults are at risk for aspiration pneumonia, outbreaks of gastroenteritis, recurrent urinary tract infection, and prosthetic device infections **(13)**.In the European Prevalence of Infection in Intensive Care (EPIC), *P.aeruginosa* was predominant gram-negative bacteria isolated from bronchopulmonary infections and accounts for 17% of health care-associted pneumonia and late–onset ventilate associated pneumonia **(14)**and accounts for significant cases of cystic fibriosis**(15)**.The distribution of isolates differs with studies and clinical specimens**(16)**.Intensive care patients especially create an environment for infection because of the debilitating effect of a prolonged hospitalisation and the application of medical equipment (airways, catheters etc) **(17)**.ICUs are generally considered epicenters of antibiotic resistance and the principal sources of outbreaks of multi-resistant bacteria. The most important risk factors are excessive consumption of antibiotics exerting selective pressure on bacteria, the frequent use of invasive devices and relative density of a susceptible patient population with severe compelling diseases**(18)**. Thus, in ICUs, empirical antibiotic treatments should be avoided and treatment should be carried out using antibiotic susceptibility tests. ICUs should be regularly monitored resistance pattern against the various antibiotics. *P. aeruginosa*was responsible for pneumonia and septicaemia with deaths rate about 30% in immunocompromised patients **(19).**

In the current study results, *Pseudomonasaeruginosa* showed resistance to amoxicillin /clavulinic Acid 96.2%,ampicillin/ sulbactam 94.5%,cefuroxime 94.6%, nalidixic acid 83%, nitrofurantoin 88%, doxycycline 82.6%,ciprofloxacin 50.89%, ceftazidime 31.5%, ceftriaxone 78%, norfloxacin 54%,and Co-Trimoxzole 80.5%. However, the highest frequency of sensitivity (85.5%) was observed with meropenem followed byamikacin 80.5%, imipenem 80%, piperacilline/tazobactam 77.2%,ceftizoxime 75%, ciprofloxacin 71.5%, Levofloxacin 66%,Cefoperazone 64%, Gentamicin 56%, ceftazidime 54.5%,moxifloxacin 49%, and cefepime 44.5%.This may be explained by the fact that routine use of these antibiotics can lead to clinically significant resistance. One remarkable finding in the present study was the highest frequency of sensitivity (85.5%) was observed with meropenem, 85.5%, amikacin (80.5%), and piperacillin/tazobactem (77.2%). These drugs were the most effective drugs against *P*.*aeruginosa* infections. This similar to study finding byTaranasarwat et al.**(20)**, who reported highest sensitivity to imipenam. Also it was quite similar to the findings of Shaikh et al (100%)**(21)**and Mohan et al., (94.3%)**(22)**. One striking feature in this study was that all the *P. aeruginosa*isolates were found to be sensitive to imipenem. This may be due to the restricted use of imipenem in this hospital. This is consistent with a report published in 2002 in Mangalore, India**(23)**. The emergence of carbapenem resistance is a serious concern **(24)**. In various studies across the world, varying rates of resistance from 4-60% have been reported for imipenem and meropenem**(25)**. Another survey found that resistance to imipenem was 19%, while other studies have reported low rates (5.8% and 9%) and high rates (38.6%) of resistance to imipenem**(26)**.Piperacillin+ tazobactam showed a sensitive rate of 77.2 % in this study and cefoperazone-sulbactum showed a lower resistance of 29.5% only, indicating beta-lactamase inhibitor markedly expands the spectrum of activity of beta-lactams, which makes the combination drug the preferred choice against *Pseudomonasaeruginosa* infections. Thus, emphasis should be given towards use of combined antibiotics in the treatment of *pseudomonal* infections **(27)**. Bayani et al. found that the resistance rate of *P. aeruginosa*to amikacin, ceftazidime*,* cefepime, imipenem,and ciprofloxacin was 53.3%, 43.3%, 40%,40%, and 33.3%, respectively, and the prevalence of *P. aeruginosa*resistant isolates has increased**(28)**.According to previous evidence, the rate of susceptibility was most productive for antimicrobial agent of class carbapenem against *Pseudomonasaeruginosa***(29)**Supported current results as 85.5% of strains were susceptible to Meronem and 80% to imipenem of class carbapenems. Although the resistance to carbapenems that include imipenem (16%) and meropenem (17.1) was low in this study, quite alarming should take into account that carbapenems are the last line of antibiotics for treating Gram-negative bacilli infections. Resistance to carbapenems may be due to a result of complex interactions of several mechanisms including production of carbapenemase, overproduction of efflux system and loss of outer membrane porins*. P. aeruginosa* isolates that are carbapenem resistant, specifically carbapenemase producing, are the worst, for the reason that they are associated with a higher mortality rate **(24)**.Amikacin in this study was noted to be the most effective drug(80.5% sensitive). However, it is not commonly prescribed drug, because of its numerous side effects including renal toxicity, blurred vision, hearing loss, Bartter-like syndromes**(30)**, neuromuscular blockade, arthralgia, and apnoea. In addition, ciprofloxacin (71.5% sensitive) proved to be within the most effective drugs for routine use among the *P. aeruginosa*strains investigated in this study. The result finding in this study was similar in a previous study finding that reported that amikacinhad the highest sensitivity against *P. aeruginosa***(9).**Also **i**n France, a higher susceptibility rate of 86% of amikacin was reported by Cavallo et al. **(31)**.An earlier study reported from Kathmandu, Nepal **(32)**shown amikacin (81.4% sensitive) and ciprofloxacin (70.3% sensitive) among *P. aeruginosa*strains examined. Amikacin seems to be a promising therapy for *pseudomonal* infection. Hence, its use should be restricted to severe nosocomial infections, in order to avoid rapid emergence of resistant strains**(33)**.However, high resistance to aminoglycosides had been reported in studies done in Bangladesh **(34)**, Turkey **(4)** and Malaysia **(35)**.Similarly, higher rates of resistance to fluoroquinolones such as ciprofloxacin resistance (92%) were shown in a study from Malaysia **(36)**.Also study findings by Zhanel et al. **(37)**reported moxifloxacin 58% and ciprofloxacin 46.7%.Because of the increasing resistance to fluroquinolone in many hospitals, its empirical usage is either banned or restricted, to bring the developing resistance rates under control.Recently, ceftazidime and cefepime are the most frequently prescribed third and fourth generation cephalosporins respectively. Ceftadizime is known antipseudomonal drug that has demonstrated high susceptibility pattern with *P. aeruginosa* isolates. The increased prevalence of ceftazidime resistant *P.*aeruginosa is related to the increased use of beta lactam antibiotics such as amoxicillin and ceftazidime.However, the resistance to cefadizime was reported as 31.5% in this study. This value of resistance waslessthan reported from Gujarat, with a resistance value of 75% **(9)**. *P. aeruginosa*strains in this study exhibited a high rate of resistance to the third generation cephalosporin drug such as ceftriaxone (78%). A much higher resistance to ceftriaxone of 75%, 86% and 93.9% had been reported in studies done in India **(38)**Bangladesh **(34)**and Nepal **(27).**Several studies have confirmed that *Pseudomonas aeruginosa*was mostly resistant against ceftriaxone. However, this high level of resistance is not quite surprising as some suggest that ceftriaxone has considerably low activity against *P. aeruginosa***(39,40)**.Another study reported the following rates of resistance to cefepime 64.8%, piperacilline/tazobatctam45%, ciprofloxacin 38.9%, levofloxacin 36.1%, gentamicin 37.3% and amikacin 30% **(41)**.Relatively low piperacilline/tazobactam resistance (11.5%) had been reported in a hospital isolates of *P. aeruginosa*in a study from Saudi Arabia **(11)**. In a study done in Kathmandu, Nepal **(27)**, *P. aeruginosa*isolates obtained from intensive care unit of a national heart centre showed a high cefoperazone-sulbactum sensitivity rateof 84.8%.A previous study discovered an increased mortality rate associated with empiric piperacillin-tazobactam therapy given to patients with *P. aeruginosa*bacteraemia; the isolates had reduced piperacillin-tazobactam susceptibility **(42)**.In this study, amoxicillin /clavulinicacid had established 96.2 % resistance. Similarly, in a study conducted in Pakistan reported by Khan et al. **(43)**had a high resistance rate of penicillin that is 98%; our findingsare also in agreement with other studies as reported bySasirekha et al. **(44)**and Ullah et al. **(45)**with respect to penicillin’s. Also the same findings were obtained with amoxicillin /clavulinic acid (1.88%) and showed increasing resistance. Multi drug efflux pumps in the inner and outer membrane of *Ps. aeruginosa*may protect the bacterium from β-lactam agents**(46)**.Similar pattern had been reported in study in Nigeria **(47)**. In addition, susceptibility to fourth-generation such as cefepime reported in India 32% **(48)** and in Bulgaria 42% **(49)**against *Pseudomonas aeruginosa* isolates. The high resistance to cephalosporins may be due to production of extended spectrum β-lactamases by the bacteria involved**(50)**.Cefuroxime was one of the cephalosporin drugs tested in this study, with resistance value of 94.6%. These high resistance value observed were comparable with the report from Gujarat, India with resistance value of 73.2% **(9)**, but higher than reports from Malaysia of 40% **(51)**.Selective pressure from the use of antimicrobial agents is a major determinant for the emergence of resistant strains.The rate of resistance for the anti-folate drug co-trimoxazole in the present study was 80.5%. In similar to previous study done in Bangladesh **(34)**showed rate of resistance for co-trimoxazole to be 93.5% in wound swab and pus isolates of *P. aeruginosa*while a Nigerian study **(52)**showed *P. aeruginosa*isolates 100% resistant to co-trimoxazole.According to resistance rates of antipseudomonal agents, imipenem and piperacillin /tazobactam were found to be effective when compared to Ceftazidime, cefepime, and Ciprofloxacin. So, imipenem which is both an anti-pseudomonal drug and carbapenem was the best drug.According tothe study findings, MDR rate (resistance to three or more of anti Pseudomonal antimicrobials (i.e. piperacillin + tazobactam, imipenem, ceftazidime and amikacin) was determined to be 4.2% (8/193). Also MDR rate for only three anti Pseudomonal antimicrobials without imipenem was 4.2% (8/193) (i.e. piperacillin + tazobactam, ceftazidime and amikacin)A study done by Unan et al.**(53)**.in Turkey reported rates of MDR, which were as high as 60%, whereas study done by Sabir et al., in Pakistan detected lower rates of MDR (22.08%)**(54)**. However, the rates of our study are comparable to a study done in Egypt, where Gad et al.**(55)**observed 36% MDR P. aeruginosa.On comparing the sensitivity patterns of these antimicrobials, it was found that there was a considerable difference in the sensitivity pattern among these studies. This indicates that the sensitivity pattern changes from hospital to hospital and population to population. Also nowadays the common antimicrobial agents are losing their efficacy against pathogens like *Ps.aeruginosa*. This has been possibly resulted from indiscriminate use of antibiotic, lack of awareness, patient non-compliance and unhygienic conditions **(56)**.According to Berglund **(57)**one of the reasons for resistance among bacteria is a result of either overuse and misuse of antibiotics. By misuse, this refers to the prescription of antibiotics without establishing bacterial infection, and the non-compliance of the patient to the full prescription. Moreover, antibiotic resistance can also be transferred horizontally between bacteria.

The currentstudy resultsindicated that *P. aeruginosa*was becoming resistant to commonly used antibiotics due to excessive consumption of antibiotics exerting selected pressure on bacteria, frequently used invasive devices and severs under laying diseases. The empirical antibiotic treatment should be limited and treatment should be carried out using antibiotic susceptibility test and efforts should be made to prevent spread of resistant bacteria**(56)**.

**5. Conclusion:**

In conclusion, results of the present study clearly demonstrated the occurrence of resistance to various antipseudomonal agents among the *pseudomonas aeruginosa*isolates. The statistics in this study showed low rates of antibiotic resistance to meropenem, amikacin, and meropenem, and piperacillin/tazobactam and maximum sensitivity against pseudomonas *aeruginosa* strains. We suggest a more restricted and a more rational use of these drugs in hospital setting in order to avoid rapid emergence of resistant strains. Regular anti-microbial susceptibility monitoring is essential of local, regional and national level isolates. This would held and guide the physicians in prescribing the right. Every effort should be made to prevent spread of resistant organisms. The solution can be planned by continuous efforts of microbiologist, clinician, pharmacist and community to promote greater understanding of this problem. Frequent hand washing to prevent spread of organism should be encouraged. Better surgical and medical care should be provided to patients during hospital stay.

**Conflict of interest**

The authors declare that they have no competing interests

**References:**

1. Kenneth T. Textbook of bacteriology: Pseudomonas aeruginosa. Wisconsin university, France. 2004; 7-15.
2. ALshaiki JMM, Toweir AA. Prevalance Pseudomonas aeruginosa Among Libyan Patients and its Association with Hospital’s Environment in Benghazi. J Med MicrobDiagn.2017; 6: 257.
3. Acar JF. Consequences of bacterial resistance to antibiotics in medical practice. Clin Infect Dis.1997; 24 (1):17 8.
4. Savas L, Duran N, Savas N, Onlen Y, Ocak S. The prevalence ans resistance pattterns of Pseudomonas aeruginosa in intensive care units in a university hospital. Turk J Med Sci. 2005; 35:317-22.
5. Carmeli Y, Troillet N, Eliopoulos GM, Samore MH. Emergence of antibiotic-resistant Pseudomonas aeruginosa: comparison of risks associated with different anti pseudomonal agents. Antimicrob Agents Chemother. 1999; 43: 1379-82.
6. RenugaS , Lakshmi K, Chitralekha S, Illamani V. Prevalence of Pseudomonas aeruginosa and its antibiotic susceptibility pattern in a Tertiary Care Hospital. Int. J. Res. Pharm. Sci. 2015; 6(1), 27-30.
7. F.R. Cockerill (Ed.), Performance Standards for Antimicrobial Susceptibility Testing: Twenty-first Informational Supplement, Clinical and Laboratory Standards Institute (CLSI), 2011.
8. Tadvi J, Javadekar TB, Bhavsar R, Garala N. Prevalence and antibiogram of Pseudomonas aeruginosa at S.S.G. Hospital, Baroda, Gujarat, India. J Res Med Den Sci. 2015; 3:204-207.
9. Javiya VA, Ghatak SB, Patel KR, Patel JA. Antibiotic susceptibility patterns of Pseudomonas aeruginosa at a tertiary care hospital in Gujarat, India. Indian J Pharmacol. 2008; 40:230-4.
10. Al-Marzoqi A.H, Al Taee Z.M. Pseudomonas aeruginosa: Antibiotic resistance pattern to different isolates in Al-Hillah city, Iraq. Journal of Natural Sciences Research. 2013; 3:23-30.
11. Al-Tawfiq JA. Occurrence and antimicrobial resistance pattern of inpatient and outpatient isolates of Pseudomonas aeruginosa in a Saudi Arabian hospital: 19982003. Int J Infect Dis. 2007; 11:109-114.
12. Pathmanathan SG, Samat NA, Mohamed R. Antimicrobial susceptibility of clinical isolates of *Pseudomonas aeruginosa* from a Malaysian Hospital. *The Malaysian Journal of Medical Sciences : MJMS*. 2009;16(2):27-32..
13. TortoraG.,FunkeB. Case, C. Microbiology: An Introduction, 6th ed.; Benjamin Cummings: California, CA, USA, 1998.
14. Vincent JL, Bihari DL, Suter PM, Bruining HA, White J, Nicolas-chanion M. The prevalence of nosocomial infection in intensive care units in Europe: results of the European Prevalence of Infection in Intensive Care (EPIC) study. JAMA 1995; 74:639-644
15. Pier GB. Role of cystic fibrosis transmembrane conductance regulator in innate immunity to Pseudomonas aeruginosa infections. Proceedings of National Academy of Science, USA. 2000; 97:8822-8828.
16. Okon KO, Agukwe PC, Oladosu W, Balogun ST, Uba A. Antibiotic resistance pattern of *Pseudomonas aeruginosa* isolated from clinical specimens in a tertiary care hospital in Northeastren Nigeria. Internet J Microbiol. 2010;8:1-6
17. Jarlier V, Fosse T, Philippon A. Antibiotic susceptibility in aerobic gram-negative bacilli isolated in intensive care units in 39 French teaching hospitals (ICU study). Intensive Care Med.1996; 22: 1057-65.
18. Weber DJ, Raasch R, Rutala WA. Nosocomial infections in the ICU: the growing importance of antibiotic-resistant pathogens. Chest. 1999; 115: 34S-41S.
19. OlayinkaA.T.,OnileB.A., Olayinka B.O.Prevalence of multidrug resistant (MDR) Pseudomonas Aeruginosa isolates in Surgical Units of Ahmadu Bello University Teaching Hospital, Zaria, Nigeria: an Indication for Effective Control Measures Annals of African Medicine.2004; 3 (1):13 - 16
20. Sarwat T, Rashid M, RastogiV, Chander Y. A comparative study of Antibiogram of Pseudomonas aeruginosa in Hospital and community acquired infections.Int.J.Curr.Microbiol.App.Sci. 2015; Special Issue-1: 286-291
21. Shaikh S, Fatima J, Shakil S, Mohd S., Rizvi D., Kamal MA. Prevalence of multidrug resistant and extended spectrum beta-lactamase producing Pseudomonas aeruginosa in a tertiary care hospital. Saudi Journal of Biological Sciences. 2015; 22, 62–64
22. Mohan, B.S., Lava, R., Prashanth, H.V., VinodNambiar, MetriBasavaraj, NayakVenkatesh, R., Baragundi Mahesh, Sri Krishna, R. Prevalence and antibiotic sensitivity pattern of Pseudomonas aeruginosa; an emerging nosocomial pathogen. Int. J. Biol. Med. Res., 2013; 4(1): 2729-2731.
23. Shenoy S, Baliga S, Saldanha DR, Prashanth HV. Antibiotic sensitivity patterns of Pseudomonas aeruginosa strains isolated from various clinical specimens. Indian J Med Sci 2002; 56(9):427-30.
24. Liu Q., Li X., Li W. Influence of carbapenem resistance on mortality of patients with Pseudomonas aeruginosa infection: a meta-analysis. Sci. Rep. 2015; 5, 11715.
25. Gonlugur U., Bakici MZ., Akkurt I., Efeoglu T. Antibiotic susceptibility patterns among respiratory isolates of Gram-negative bacilli in a Turkish university hospital. BMC Microbiol. 2004; 4, 32.
26. Khan M.A., Faiz A. Antimicrobial resistance patterns of Pseudomonas aeruginosa in tertiary care hospitals of Makkah and Jeddah. Ann. Saudi Med. 2016; 36, 23-28.
27. Bhandari S, Banjara MR, Lekhak B, Bhatta DR, Regmi SR. Multi-drug and pan-drug resistant Pseudomonas aeruginosa: a challenge in post-antibiotic era. Nepal J Sci Tech. 2012; 13(2):197-202.
28. Bayani M., Siadati S., Rajabnia R., Taher AA. Drug Resistance of *Pseudomonas aeruginosa*and *Enterobacter cloacae* Isolated from ICU, Babol, Northern Iran*. Int. J. Mol. Cell Med*. 2012; 2, 204-209.
29. Turner P. J. “Meropenem and imipenem activity against Pseudomonas aeruginosa isolates from the MYSTIC Program,” DiagnosticMicrobiology and InfectiousDisease. 2006; 56 (3): 341–344.
30. Juayang AC, Maestral DG Jr, de Los Reyes GB, Acosido MA, Gallega CT. Review on the antimicrobial resistanceof pathogens from tracheal and endotracheal aspirates of patients with clinical manifestations of pneumonia in Bacolod City in 2013. Int J Bacteriol. 2015; 2015:942509.
31. Cavallo J. D., Hocquet D., Plesiat P., Fabre R., Roussel-Delvallez M.Susceptibility of Pseudomonas aeruginosa to antimicrobials: a 2004 French multicentre hospital study. Journal of Antimicrobial Chemotherapy. 2007;59, (5): 1021–1024.
32. Koirala P, Bhatta DR, Ghimire P, Pokhrel BM, Devkota U. Bacteriological profile of tracheal aspirates of the patients attending a neuro-hospital of Nepal. Int J Life Sci.2010;4:60-65
33. Poole K. Aminoglycosides resistance in Pseudomonas aeruginosa. Antimicrob Agents Chem. 2005; 49:479-87.
34. Rashid A, Chowdhury A, Rahman SHZ, Begum SA, Muazzam N. Infections by Pseudomonas aeruginosa and antibiotic resistance pattern of the isolates from Dhaka Medical College Hospital. Bangladesh J Med Microbiol. 2007; 1(2):48-51.
35. Fazlul MKK, Zaini MZ, Rashid MA, Nazmul MHM. Antibiotic susceptibility profile s of clinical isolates of Pseudomonas aeruginosa from Selayang Hospital, Malaysia. Biomed Res. 2011; 22(3):263-66.
36. Al-KabsiAM, Yusof MYBM, Sekaran SD. Antimicrobial resistance pattern of clinical isolates of Pseudomonas aeruginosa in the University of Malaya Medical Center, Malaysia. Afr J Microbiol Res. 2011; 5(29):5266-72.
37. ZhanelG.G., LaingN.M., Nichol K. A.Antibiotic activity against urinary tract infection (UTI) isolates of vancomycinresistant enterococci (VRE): results from the 2002 North American vancomycin resistant enterococci susceptibility study (NAVRESS),” Journal of Antimicrobial Chemotherapy. 2003; 52 (3): 382–388.
38. Arora D, Jindal N, Kumar R, Romit. Emerging antibiotic resistance in Pseudomonas aeruginosa. Int J Pharm PharmSci 2011;3(2):82-4.
39. Bassetti D, Cruciani M, Solbiati M, Rubini F, Gandola L, Valenti G, et al. Comparative efficacy of ceftriaxone versus ceftazidime in the treatment of nosocomial lower respiratory tract infections. Chemotherapy. 1991; 37:371-5. 20.
40. Mody L, Bradley SF, Strausbaugh LJ, Muder RR. Prevalence of ceftriaxone - And ceftazidime-resistant gram-negative bacteria in longterm-care facilities. Infect Control HospEpidemiol. 2001; 22(4):193-4.
41. Dash M., Padhi S., Narasimham M.V., Pattnaik S. Antimicrobial resistance pattern of Pseudomonas aeruginosa isolated from various clinical samples in a tertiary care hospital, South Odisha, India. Saudi J. Health Sci.2014; 3, 15-19.
42. Tam VH, Gamez EA, Weston JS, Gerard LN, LaRocco MT, Caeiro JP, et al. Outcomes of Bacteremia due to Pseudomonas aeruginosa with Reduced Susceptibility to Piperacillin-Tazobactam: Implications on the Appropriateness of the Resistance Breakpoint. Clin Infect Dis. 2008; 46:862-867.
43. Khan J. A., IqbalZ., Ur RahmanS., FarzanaK., Khan A. “Prevalence and resistance pattern of Pseudomonas aeruginosa against various antibiotics,” Pakistan Journal of Pharmaceutical Sciences.2008; 21(3):311–315.
44. Sasirekha B., ManasaR., RamyaP., SnehaR.Frequency and antimicrobial sensitivity pattern of extended spectrum 𝛽-lactamases producing E. coli and Klebsiellapneumoniae isolated in a tertiary care hospital. Journal ofMedical Sciences. ; 2010; 3(4): 265–271.
45. Ullah F., MalikS. A., Ahmed J.Antimicrobial susceptibility and ESBL prevalence in Pseudomonas aeruginosa isolated from burn patients in the North West of Pakistan,” Burns. 2009; 35 (7): 1020–1025.
46. Srikumar R, Li XZ, Poole K. Inner membrane efflux components are responsible for Î²-lactam specificity of multi drug efflux pumps in Pseudomonas aeruginosa. J. Bacteriol. 1997; 179(2): 7875-7881.
47. GTA, Jonah P, Ayeni JA. Multidrug Resistant Pseudomonas aeruginosa in Contemporary Medical Practice: Findings From Urinary Isolates At A Nigerian University Teaching Hospital. Nigerian Journal of Physiological Sciences. 2008; 23(1-2):105-109.
48. ChaudhuryA. In vitro activity of Cefpirome: a new fourth generation cephalosporin. Indian Journal of Medical Microbiology. 2003; 21 (1): 52–55,.
49. StratevaT., Ouzounova-RaykovaV.,MarkovaB.,TodorovaA.,Marteva-ProevskaY., MitovI.Widespread detection of VEB-1-type extended-spectrum beta-lactamases among nosocomial ceftazidime-resistant Pseudomonas aeruginosa isolates in Sofia, Bulgaria. Journal of Chemotherapy. 2007; 19 (2): 140–145.
50. Mathur P, Kapil A, Das B, DhawanB.. Prevalence of extended spectrum Î²-lactamase producing gram negative bacteria in a tertiary care hospital. Indian J Med Res 2002; 115(2): 153-157.
51. Nwankwo EOK, Shuaibo SA. Antibiotic susceptibility patttern of clinical isolates of Pseudomonas aeruginosa in a tertairy health institution in Kano, Nigeria. J Med Biomed Sci 2010; 37-40.
52. Lim KT, Yasin RY, Yeo CC. Genetic fingerprinting and antimicrobial susceptibility profiles of Pseudomonas aeruginosa hospital isolates in Malaysia. Journal of Microbiology and Infectious Diseases. 2009; 42:197-209.
53. Unan D, Gnseren F. The resistance of P. aeruginosa strains isolated from nosocomial infections against various antibiotics. MikrobiyolBult. 2000; 34: 255-60
54. Sabir R, Alvi SFD, Fawwad A. Antimicrobial susceptibility pattern of aerobic microbial isolates in a clinical laboratory in Karachi- Pakistan. Pak J Med Sci. 2013; 29(3): 851–5
55. Gad GF, El-Domany RA, Zaki S, Ashour HM. Characterization of Pseudomonas aeruginosa Isolated from Clinical and Environmental Samples in Minia, Egypt: Prevalence, Antibiogram and Resistance Mechanisms. J AntimicrChemother. 2007; 60: 1010–7.
56. THE CURRENT STATUS OF ANTIBIOTIC SENSITIVITY OF PSEUDOMONAS AERUGINOSA ISOLATED FROM VARIOUS CLINICAL SAMPLES HemaliParmar \*1, AditiDholakia 2, Dharmesh Vasavada3, Hitesh Singhala4 Int J Res Med. 2013; 2(1);1-6.
57. Berglund, B. Environmental dissemination of antibiotic resistance genes and correlation to anthropogenic contamination with antibiotics. J. Infect. Ecol. Epidemiol. 2015, 5, 28564.