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### **REVIEW ARTICLE**

# **PHARMACOKINETIC DRUG-DRUG INTERACTIONS MEDIATED BY ORGANIC CATION TRANSPORTERS: ARE ANTIMALARIAL DRUGS SIGNIFICANTLY AFFECTED?**

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interactions mediated by organic cation transporters: Are antimalarial drugs significantly affected? Universal Journal of Pharmaceutical

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Pharmacokinetic drug-drug

### **Abstract**

A clear understanding of all the processes that influence drug disposition is important to enable a prediction of pharmacokinetic drug interactions. Xenobiotics are transported across bio-membranes and the process is mediated by various membrane transporters which include "Organic Cation Transporters" (OCTs). OCTs are specifically involved in the transport of several molecules that have positive charges *in vivo* and these include a wide variety of weakly basic drugs.As several antimalarial drugs are weakly basic and can be cationic in biological fluids, the contribution of OCTs to the drug disposition appears to be underestimated because most studies on the pharmacokinetic drug interaction with antimalarial drugs are focused on the interactions at the sites of drug metabolism. This review provides an update on the significance of OCTs on the pharmacokinetic disposition of antimalarial agents with a view to identifying potentials for drug interactions that could involve concurrently administered drugs which are either inhibitors or substrates of OCTs. A very significant likelihood exists for concurrent use of antimalarial drugs with other medicines because of the occurrence of comorbidities with malaria. There are limited studies on antimalarial pharmacokinetic drug-drug interaction studies in which role of OCTs are investigated. From the literature, and using *in vitro* studies, the following antimalarial drugs, chloroquine, piperaquine, proguanil, and cycloguanil have been reported to be substrates of different OCTs while tafenoquine, pyrimethamine, trimethoprim, quinine, and mefloquine were shown to be inhibitors. Atovaquone and artesunate were shown not to be substrates and did not demonstrate any inhibitory potency. This information provide basis for prediction of any potential interaction between antimalarial drugs and other coadministered medicines which are inhibitors/substrates of the transporter proteins. **Keywords:** Antimalarial drugs, drug-drug interactions, organic cation transporters, pharmacokinetics.

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**INTRODUCTION**

The OCTs along with "Organic Anion Transporters" (OATs) belong to the family of membrane transporters which itself is a part of the "Solute Carrier" (SLC) super family which consists of 65 SLC families. The function of the various SLC families is to control the transport of most endogenous compounds and xenobiotics across bio-membranes**<sup>1</sup>** [.](#page-5-0) OCTs are specifically involved in the transport of several molecules that are positive in charge at the pH of biological fluids, examples of which include the endogenous amines and a variety of weakly basic drugs**<sup>2</sup>** [.](#page-5-1) These Transporters include OCT3, OCT2, OCT1, OCTN1, OCTN2, Plasma membrane Monoamine Transporter (PMAT), "Multidrug And Toxin

Extrusion-1" (MATE-1), and "Multidrug And Toxin Extrusion-2-k"(MATE2–K) **[2,](#page-5-1)[3](#page-5-2)** They all have differences in their organ or tissue localizations. In humans, for example, OCT1 is most abundant on the membrane of intestines and hepatocytes where it is involved in drug uptake, while OCT2 is most abundant on the cell membrane of kidney tubules where it facilitates drug secretion**<sup>4</sup>** [.](#page-5-3) The other OCTs are distributed in different specific tissues in the body but the MATEs are more found at the hepatocytes and kidney where they mediate the efflux of diverse organic cation substrates**[1](#page-5-0)** . OCT2 operates in concert with MATE in facilitating renal tubular secretion of cationic species, as OCT2 drives cationic uptake into the proximal tubule epithelial cells while MATE mediates efflux into the renal tubules for elimination

into urine. From the tissue localization of these OCTs, it is not surprising that OCT2 and OCT1 play more prominent roles in intestinal drug absorption, as well as hepatic plus renal elimination of several drug[s](#page-5-4)**<sup>5</sup>** .

All the processes that influence disposition of drugs in the body ought to be fully understood as this will enhance the prediction of potential pharmacokinetic drug interactions which have attracted increased attention due to popularity of co-administration of several drugs, especially in the elderly. It is reported that approximately 40% of orally applied drugs are cationic at the pH of body fluids. Since these cationic moieties are hydrophilic, they undergo minor or no passive transfer across biological membranes, necessitating the involvement of OCTs**<sup>6</sup>** [.](#page-5-5) Indeed, different Drug regulatory Agencies including the "Food and Drug Administration" (FDA) and "European Medicines Agency" (EMA) have recommended preclinical testing of new drug molecules for their inhibition potentials for OCT1, OCT2, MATE1, and MATE2-K**<sup>6</sup>** [.](#page-5-5) Most studies on the pharmacokinetic drug interactions with antimalarial drugs are focused on interactions at the sites of drug metabolism. As most antimalarial drugs are weakly basic and can be cationic at pH of biological fluids, it is likely that OCTs contribute to disposition of the drugs. The World Malaria Report of 2023 indicates that higher levels of treatment failure have been shown in some studies in the WHO African Region, and these results could be an indication of emergence of resistance to ACT partner drugs**[7](#page-5-6)** . Although several investigators are working on development of new antimalarial medicines, optimization of the therapeutic utility of the existing ones is still an important approach. A full understanding of processes of DDIs involving antimalarials will, undoubtedly, contribute to enhancing the treatment outcomes with these drugs. This review provides an update on the significance of OCTs on the disposition kinetics of antimalarial drugs with a view to identifying potentials for drug interactions that could involve concurrently administered medicines which are either inhibitors or substrates of OCTs.

Relevant keywords were employed to perform a thorough search of selected databases including PubMed, Elsevier, Scopus and Science Direct, to obtain the articles published in the English language that were used for this review.

# **Drug interactions at OCTs**

Drug with drug interactions (DDIs) through drug transporter proteins have been receiving increased attention following several observations that inhibition of these transporters affect drug pharmacokinetics especially drug absorption and elimination**[8](#page-5-7)** . There is a high potential for occurrence of OCT-mediated DDI since OCTs have high affinity to bind to a wide range of drug molecules that have positive charges *in vivo***<sup>9</sup>** [.](#page-5-8) These DDIs can result in increased plasma drug levels leading to increase in drug efficacy or toxicity, if renal secretion of the drug occurring at the OCT2 expressed at the tubular cells of the kidney, is inhibited by a concurrently administered drug. On the other hand, the

DDI may result in decreased efficacy or therapeutic failure following decreased absorption as a result of inhibition of OCT1 at the enterocytes which mediate intestinal drug absorption of cationic drugs. The relevance of OCT-mediated DDI is re-enforced by the fact that the pharmacokinetics of at least 120 drugs is influenced by the activities of OCTs**[9,](#page-5-8)[10](#page-5-9)**. There are several drugs and compounds demonstrating *in vitro* inhibitory activities on OCTs but translation to clinically significant effects are limited.

# **Clinically relevant DDIs at OCTs**

The cellular uptake of several organic cationic drugs through membranes of hepatocytes and enterocytes is facilitated by OCT1 and this contributes to regulating the gastrointestinal uptake and metabolism of these drugs**[11](#page-5-10)**. When two or more drugs that are substrates of OCT are co-administered, they can compete for binding at the OCT site resulting in inhibition of transport of the drug with lower binding affinity or lower concentration. Also, similar to other transporter proteins, there are several compounds that are nonsubstrates of OCT but can inhibit substrates of OCT. These inhibitors can undergo competitive or noncompetitive inhibition. In general, the molar mass of these inhibitors of OCT are larger than those of the substrates<sup>[12](#page-5-11)</sup>. Although there is abundance of preclinical studies supporting a role of OCT in DDI, only a few of these OCT-mediated DDIs have been reported in humans**<sup>9</sup>** [.](#page-5-8) Also, the clinically relevant effects of individual OCTs on the modulation of the pharmacokinetics of specific drugs have only been elucidated in a few cases. This is attributable to the fact that more than one OCT with different effects are often available at different biological membranes that are involved in the drug uptake or excretion**[6](#page-5-5)** . Thus, the effect of inhibition of an OCT at a location on plasma drug levels can counteract or mask the effect of inhibition of the same OCT at another tissue location. For example, inhibition of OCT1 at intestinal membrane can result in a decrease in drug absorption and subsequent decreased plasma drug levels. On the other hand, if OCT1 is inhibited at the hepatocytes, there would be decreased hepatic uptake (and decreased metabolism) which can cause blood levels of OCT1 substrates to increase. Drug-drug interactions in human subjects involving OCTs have been severally documented with metformin and a few other drugs shown in Table 1**[13](#page-5-12)[-25](#page-5-13)** .

It is evident in these studies that inhibition of OCT2 and MATE at the proximal renal tubules results in increase in plasma drug concentrations due to reduction in drug secretion. Hence, OCT2 and MATEs at the kidneys do contribute to DDIs with clinical relevance as they accept several substrates of OCT1**<sup>2</sup>** [.](#page-5-1) This informs the decision for drug interactions through these transporter proteins to now be recognized in the drug development and approval process**<sup>6</sup>** [.](#page-5-5) It is pertinent to note that when no inhibitors in clinical use are available for an OCT, additional evidence of involvement of the OCT in drug disposition can be provided by pharmacogenetic studies in carriers of the OCT null alleles**[26](#page-5-14)** .



### **Table 1: Some clinically observed drug-drug interactions resulting from modulation of organic cation transport proteins.**

For example, individuals carrying OCT1 alleles of reduced function were observed to have higher "area under plasma drug concentration" (AUC) and "maximum plasma drug concentration"  $(C_{\text{max}})$ attributed to reduced hepatic clearance of the drug**[27](#page-5-26)** .

### **Pharmacokinetic interactions of antimalarials through Organic Cation Transporters**

Concurrent administration of antimalarial drugs with other medicines is very common because of the occurrence of comorbidities with malaria. A number of studies have shown that a significant percentage of patients with malaria had a reported comorbidity**[28,](#page-6-0)[29](#page-6-1)**. In addition, the common environmental and socioeconomic factors in some resource-limited countries promote the coexistence of different infectious and/or noninfectious diseases in the same individual resulting in comorbidity**[30,](#page-6-2)[31](#page-6-3)**. Therefore, there is a high prevalence of concurrent administration of antimalarial drugs with other drugs for treatment of comorbidities, and these can potentially result in significant drug-drug interactions**[31](#page-6-3)**. For example, in a study in our laboratories, plasma quinine concentrations increased 4-fold following concurrent ritonavir administration and this was accompanied by a marked reduction in plasma drug clearance (CL/F) of quinine<sup>[32](#page-6-4)</sup>. There are limited studies on antimalarial pharmacokinetic DDI studies in which role of OCTs are investigated. With availability of copious literature

on compounds and medicines that are OCT substrates orinhibitors**[33](#page-6-5)**, it may be possible to predict the possibility of a pharmacokinetic DDI through the OCT pathway. Also, previous DDI studies can be reevaluated to ascertain the involvement of OCTs in the overall outcome of the interaction. A summary of the antimalarial drugs with reported DDIs mediated through OCTs are presented in Table 2. Also, suggestions are proffered on potential DDIs involving antimalarial drugs reported as substrates of OCTs and other drugs that are potent OCT inhibitors which may likely be co-administered with the antimalarial. A compilation of drugs which are MATE1 inhibitors is available at drug bank**[34](#page-6-6)**. In this list are commonly used drugs such as cimetidine, levofloxacin, and trimethoprim. While a list of drugs which include codeine, cimetidine, desipramine, probenecid, clopidogrel, efavirenze, nevirapine etc, that are inhibitors of OCT1 is also available at drug bank**[35](#page-6-7)** and complied by Zhou *et al***[9](#page-5-8)** .

# **4-Aminoquinoline**

**Amodiaquine**: *In vitro* inhibition of OCT2 by amodiaquine demonstrated that the calculated DDI index of the drug is below 0.1, indicating that the occurrence of DDIs with clinical significance is unlikely for amodiaquine through the involvement of OCTs**[36](#page-6-8)** .





**Chloroquine:** In a study to elucidate the renal molecular mechanism of chloroquine tubular secretion, it was demonstrated that OCT2 appears not to be involved in chloroquine transport into the kidney, however, the drug is a MATE substrate. This finding suggests that inhibitors of MATE1 can potentially alter chloroquine elimination at renal pathway, if concurrently administered**[37](#page-6-9)** .

### **8-Aminoquinolines**

**Primaquine:** In an *in vitro* study to verify the involvement of OCT1 in uptake of primaquine into the hepatocytes, it was observed that primaquine transport was inhibited by quinidine, an OCT1 inhibitor. The study concluded that OCT1 effectively transports primaquine and that increased uptake of primaquine into hepatocyte may be mediated by OCT1**[38](#page-6-10)**. The clinical implication of inhibition of OCT1 on the efficacy of primaquine is not clear as the drug is effective at the hypnozoites phase of *P. vivax* in the liver.

**Tafenoquine:** Studies have shown that tafenoquine inhibited metformin transport by the OCTs and MATEs**[39](#page-6-11)**. This shows that, following a concurrent administration of OCT2 and MATE substrates with tafenoquine, there is a potential for increased plasma concentrations of these substrates with an attendant increased efficacy or risk of toxicity of the drugs. Therefore, concurrent administration of tafenoquine with OCT2 and MATE substrates should be done with caution.

### **Artemisinin derivatives**

Artemisinin derivatives are not weakly basic compounds and they are not cationic *in vivo*, hence, are not expected to be transported by OCTs or MATE. However, an *in vitro* study that used transporteroverexpressing MDCKII cells and probe substrate for OCT1 showed that artemisinin but not artesunate

inhibited OCT1-mediated transport**[40](#page-6-12)** . Elucidation of the clinical implications of co-administration of artemisinin with OCT1 substrates requires more studies.

# **Biguanides**

Proguanil and its major active metabolite, cycloguanil, have been reported to be substrates of OCTs and MATEs, and these suggest that these setransporters can play significant roles in pharmacokinetics of both drugs**[38](#page-6-10)**. This entails that there is possibility of DDI when OCT substrates are concurrently administered with proguanil or cycloguanil. Thus, for typical substrates of these transporters such as metformin, DDIs may likely occur following simultaneous drug administration.

# **Diaminopyrimidines**

**Pyrimethamine:** *In vitro* studies have shown Pyrimethamine to be a specific MATE inhibitor<sup>[10,](#page-5-9)[41](#page-6-13)</sup>. The achieved plasma concentrations of the drug when given at therapeutic doses are sufficient to inhibit MATE-mediated renal drug excretion as evident in a marked decrease in metformin (MATE substrate) renal elimination**[14](#page-5-15)** . Thus, DDI between pyrimethamine and metformin results in significantly increased metformin AUC and has been ascribed to MATEs inhibition and not OCT2**[14](#page-5-15)**. It is recommended that pyrimethamine can serve as a probe in inhibition studies of MATE transport proteins**[41](#page-6-13)** .

**Trimethoprim:** Trimethoprim has been identified to possess an inhibitory potency for MATE1-mediated transport, and this was further investigated by using metformin (a model substrate). It was observed that a concentration-related MATE1 inhibitory effect was produced by trimethoprim**[37](#page-6-9)** .

# **Phenanthrene methanol**

Phenanthrene methanol antimalarials include halofantrine and lumefantrine. An exhaustive search of the literature did not reveal any study indicating that these drugs are substrates or inhibitors of OCTs or MATE. Until this is verified, DDIs with these drugs mediated through cationic transporters pathway cannot be ascertained.

# **Quinoline-methanol**

**Quinine:** Studies have demonstrated *in vitro* that quinine is not only a substrate but also an inhibitor of MATEs. At therapeutic doses, quinine achieves plasma concentrations high enough to inhibit MATEs**[10](#page-5-9)** . Further studies have also revealed that quinine is effective as an inhibitor of OCTs**[40](#page-6-12)** . In a pharmacokinetic study that investigated the interaction of quinine with ritonavir, the reported four-fold increase in plasma levels of quinine was attributed to interaction at the level of metabolizing enzyme, CYP3A4**[32](#page-6-4)**. With the increasing knowledge of involvement of OCTs in drug disposition, the remarkable increase in quinine blood concentrations along with modest increase in ritonavir blood levels, may, in part, be as a result of inhibition of MATEs since ritonavir**[10](#page-5-9)** just like quinine, is an inhibitor of MATEs. Elucidation of impact of inhibition of renal secretion of quinine by ritonavir through inhibition of efflux by MATEs, requires further studies.

**Mefloquine:** The interactions of different antimalarial drugs (artesunate, artemisinin, chloroquine, mefloquine, pyrimethamine, and quinine) with OCTs were evaluated in an *in vitro* study that used MDCKII cells and relevant OCTs probe compounds**[40](#page-6-12)**. The results indicated that all the compounds except artesunate demonstrated OCT1 and OCT2 inhibition. The inhibitory potencies for OCT1 were: quinine> artemisinin>mefloquine>pyrimethamine> chloroquine; while the degrees of inhibition of OCT2 by the antimalarials were: pyrimethamine>quinine> chloroquine>mefloquine**[40](#page-6-12)**. The implication of this finding for mefloquine disposition *in vivo* is not certain.

# **Quinone**

**Atovaquone:** Atovaquone is known to be predominantly cleared from the body through biliary excretion as the drug concentration in bile is ≥100-fold higher compared to that of the blood, and this is indicative of active biliary excretion. An *in vitro* study failed to demonstrate the involvement of OCTs and OATs in atovaquone liver uptake and excretion into bile**[42](#page-6-14)** . Hence, co-administration of atovaquone with OCT substrates is not expected to result in any DDI.

# **Sulfonamides/sulfones**

Antimalarial Sulfonamides include sulphadoxine and Dapsone. Sulfonamides dissociate in aqueous media into anionic moieties while OCTs substrates are cationic or weakly basic compounds that have positive charges *in vivo***<sup>6</sup>** [.](#page-5-5) There is no report in the literature on inhibitory potency of sulfonamides on OCTs. Thus, DDIs with sulphadoxine or dapsone with OCT substrates or inhibitors are theoretically not expected to occur.

# **CONCLUSIONS**

Malaria continues to constitute a source of major concern to health authorities and this merits embarking on more research including pharmacokinetic studies on interactions of antimalarial agents with other drugs that may be co-administered in situations of malaria comorbidity. There is a high prevalence of concurrent administration of antimalarials together with other medicines used for malaria comorbidities. This can potentially result in significant DDIs through drug metabolism and/or transporter proteins. The relevance of OCT-mediated DDI is re-enforced by the fact that there is a high number of drugs demonstrating inhibitory potencies toward OCTs and MATEs. However, translation to clinically significant outcomes has not been very apparent. There are limited studies on antimalarial pharmacokinetic drug-drug interaction studies in which role of OCTs are investigated. From the literature, and using *in vitro* studies, some antimalarial drugs (Chloroquine, Piperaquine, proguanil, and cycloguanil) have been reported to be substrates of different OCTs while Tafenoquine, pyrimethamine, trimethoprim, quinine, and mefloquine were shown to be inhibitors. Atovaquone and artesunate were shown not to be substrates and did not demonstrate any inhibitory potency. This information provide basis for prediction of any potential interaction between antimalarial drugs and other co-administered

medicines which are substrates/inhibitors of OCTs and MATEs. Generally, investigations on drug-drug interactions coupled with studies of impact of pharmacogenetics have demonstrated that OCTs and MATEs contribute to the pharmacokinetics of some drugs, including the antimalarials.

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### **AUTHOR'S CONTRIBUTION**

**ONYEJI CO:** Writing original draft, review, methodology, data curation, literature survey, editing.

# **DATA AVAILABILITY**

Data will be made available on request**.**

#### **CONFLICT OF INTEREST**

There is no conflict of interest.

#### **REFERENCES**

- <span id="page-5-0"></span>1. Samodelov SL, Kullak-Ublick GA, Zhibo Gai Z, Visentin M. Organic cation transporters in human physiology pharmacology, and toxicology. Int J Mol Sci 2020; 21: 7890. *<https://doi:10.3390/ijms21217890>*
- <span id="page-5-1"></span>2. Koepsell H. Organic cation transporters in health and disease. Pharm Rev 2020; 72: 253-319. *<https://doi.org/10.1124/pr.118.015578>*
- <span id="page-5-2"></span>3. Wang J. The plasma membrane monoamine transporter (PMAT): structure, function, and role in organic cation disposition. Clin Pharmacol Ther 2016;100(5):489–499. *<https://doi.org/10.1002/cpt.442>*
- <span id="page-5-3"></span>4. Motohashi H, Sakurai Y, Saito H, *et al*. Gene expression levels and immunolocalization of organic ion transporters in the human kidney. J Am Soc Nephrol 2002; 13:866 - 874. *<https://doi.10.1681/ASN.V134866>*
- <span id="page-5-4"></span>5. Neuho S, Ungell AL, Zamora I, Artursson P. pHdependent bidirectional transport of weakly basic drugs across Caco-2 monolayers: Implications for drug-drug interactions. Pharm Res. 2003; 20: 1141–1148. *<https://doi.org/10.1023/A:1025032511040>*
- <span id="page-5-5"></span>6. Koepsell H. Update on drug-drug interaction at organic cation transporters: mechanisms, clinical impact, and proposal for advancing *in vitro* testing. Expert Opin Drug Metabolism Toxicol 2021; 17(6): 635-653. *<https://doi.10.1080/17425255.2021.1915284>*
- <span id="page-5-6"></span>7. World malaria report 2023. Geneva: World Health Organization; 2023.
- <span id="page-5-7"></span>8. Liu CX, Yi XL, Fan HR, *et al*. Effects of drug transporters on pharmacological responses and safety. Curr Drug Metabol 2015; 16: 732–752.
	- *<https://doi:10.2174/138920021609151201112629>*
- <span id="page-5-8"></span>9. Zhou S, Zeng S, Shu Y. Drug-drug interactions at organic cation transporter 1. Front Pharmacol 2021; 12:628705. *<https://doi.10.3389/fphar.2021.628705>*
- <span id="page-5-9"></span>10. Nies AT, Schwab M. Organic cation transporter pharmacogenomics and drug-drug interaction. Expert Rev Clin Pharmacol 2010; 3: 707–711. *<https://doi.10.1586/ecp.10.60>*
- <span id="page-5-10"></span>11. Shu Y. Research progress in the organic cation transporters. Zhong Nan Da Xue Xue Bao Yi Xue Ban 2011; 36: 913–926. *<https://doi:10.3969/j.issn.1672-7347.2011.10.001>*
- <span id="page-5-11"></span>12. Brosseau N, Ramotar, D. The human organic cation transporter OCT1 and its role as a target for drug responses. Drug Metab Rev 2019; 51: 389–407. *<https://doi.org/10.1080/03602532.2019.1670204>*
- <span id="page-5-12"></span>13. Tsuda M, Terada T, Ueba M, *et al*. Involvement of human multidrug and toxin extrusion 1 in the drug interaction between cimetidine and metformin in renal epithelial cells. J Pharmacol Exp Ther 2009; 329(1):185–191. *<https://doi.org/10.1124/jpet.108.147918>*
- <span id="page-5-15"></span>14. Kusuhara H, Ito S, Kumagai Y, *et al*. Effects of a MATE protein inhibitor, pyrimethamine, on the renal elimination of metformin at oral microdose and at therapeutic dose in healthy subjects. Clin Pharmacol Ther 2011;89(6): 837– 844. *<https://doi.org/10.1038/clpt.2011.36>*
- <span id="page-5-16"></span>15. Johansson S, Read J, Oliver S, *et al*. Pharmacokinetic evaluations of the co-administrations of vandetanib and metformin, digoxin, midazolam, omeprazole or ranitidine. Clin Pharmacokinet 2014; 53(9): 837–847. *<https://doi.org/10.1007/s40262-014-0161-2>*
- <span id="page-5-17"></span>16. Song IH, Zong J, Borland F, *et al*. The effect of dolutegravir on the pharmacokinetics of metformin in healthy subjects J Acquir Immune Defic Syndr, 2016; 42:400-407. *[https://10.1097/QAI.0000000000000983](https://10.0.4.73/QAI.0000000000000983)*
- <span id="page-5-18"></span>17. Shibata M, Toyoshima J, Kaneko Y, Oda K, Nishimura T. A drug–drug interaction study to evaluate the impact of peficitinib on OCT1- and MATE1-mediated transport of metformin in healthy volunteers. Eur J Clin Pharmacol 2020; 76:1135–1141. *[https://doi.org/10.1007/s00228-020-02876-2](https://doi.org/10.1007/s00228-020-02876-)*
- <span id="page-5-19"></span>18. Cho SK, Yoon JS, Lee MG, *et al*. Rifampin enhances the glucose-lowering effect of metformin and increases OCT1 mRNA levels in healthy participants. Clin Pharmacol Ther 2011; 89: 416–421. *<https://doi:10.1038/clpt.2010.266>*
- <span id="page-5-20"></span>19. Topletz-Erickson AR, Lee AJ, Mayor JG, *et al*. Tucatinib inhibits renal transporters OCT2 and MATE without impacting renal function in healthy subjects. J Clin Pharmacol. 2020; 61(4):461–471. *<https://doi.org/10.1002/jcph.1750>*
- <span id="page-5-21"></span>20. Moore KH, Yuen GJ, Raasch RH, *et al*. Pharmacokinetics of lamivudine administered alone and with trimethoprimsulfamethoxazole. Clin Pharmacol Ther 1996;59(5):550– 558. *[https://doi.org/10.1016/S0009-9236\(96\)90183-6](https://doi.org/10.1016/S0009-9236(96)90183-6)*
- <span id="page-5-22"></span>21. van Crugten J, Bochner F, Keal J, Somogyi A. Selectivity of the cimetidine-induced alterations in the renal handling of organic substrates in humans. Studies with anionic, cationic and zwitterionic drugs. J Pharmacol Exp Ther 1986; 236(2): 481–487.
- <span id="page-5-23"></span>22. Somogyi AA, Bochner F, Sallustio BC. Stereoselective inhibition of pindolol renal clearance by cimetidine in humans. Clin Pharmacol Ther 1992; 51(4), 379– 387. *<https://doi.org/10.1038/clpt.1992.37>*
- <span id="page-5-24"></span>23. Abel S, Nichols DJ, Brearley CJ, Eve MD. Effect of cimetidine and ranitidine on pharmacokinetics and pharmacodynamics of a single dose of dofetilide. Br J Clin Pharmacol. 2000; 49(1): 64–71. *<https://doi.org/10.1046/j.1365-2125.2000.00114.x>*
- <span id="page-5-25"></span>24. Kosoglou T, Rocci ML Jr, Vlasses PH. Trimethoprim alters the disposition of procainamide and Nacetylprocainamide. Clin Pharmacol Ther 1988; 44(4), 467–477. *<https://doi.10.1038/clpt.1988.181>*
- <span id="page-5-13"></span>25. Blackhall FH, O'Brien M, Schmid P, *et al*. A phase I study of vandetanib in combination with vinorelbine/cisplatin or gemcitabine/cisplatin as first-line treatment for advanced non-small cell lung cancer. J Thorac Oncol 2010; 5(8): 1285–1288. *<https://doi.10.1080/03602532.2019.1670204>*
- <span id="page-5-14"></span>26. Zamek-Gliszczynski MJ, Taub ME, Chothe PP, *et al*. Transporters in drug development: 2018 ITC recommendations for transporters of emerging clinical importance. Clin Pharmacol Ther 2018; 104 (5), 890–899. *<https://doi:10.1002/cpt.1112>*
- <span id="page-5-26"></span>27. Shu Y, Brown C, Castro RA, *et al*. Effect of genetic variation in the organic cation transporter 1, OCT1, on metformin pharmacokinetics. Clin Pharmacol Ther 2008; 83(2): 273- 279. *<https://doi.10.1038/sj.clpt.6100275>*
- <span id="page-6-0"></span>28. Ghanghoriya P, Borkar R , Lazarus M, Ajmariya M. Study of malaria and associated co-morbidity in children admitted with fever manifestation in a tertiary care centre. Int J Contemp Pediatr 2020; 7(8):1705-1710. *<https://doi.org/10.18203/2349-3291.ijcp20203161>*
- <span id="page-6-1"></span>29. Watts C, Atieli H, Alacapa J, *et al*. Rethinking the economic costs of hospitalization for malaria: accounting for the comorbidities of malaria patients in western Kenya. Malar J 2021; 20:429: 1 – 9. *<https://doi.org/10.1186/s12936-021-03958-x>*
- <span id="page-6-2"></span>30. Osakunor DNM, Sengeh DM, Mutapi F. Coinfections and comorbidities in African health systems: At the interface of infectious and noninfectious diseases. PLoS Negl Trop Dis 2018; 12(9):e0006711. *<https://doi.org/10.1371/journal.pntd.0006711>*
- <span id="page-6-3"></span>31. Noor S, Ismail M, Khadim F. Potential drug–drug interactions associated with adverse clinical outcomes and abnormal laboratory findings in patients with malaria. Malar J 2020; 19(316): 1–13. *[https://doi.org/10.1186/s12936-020-03392-5](https://doi.org/10.1186/s12936-020-03392-)*
- <span id="page-6-4"></span>32. Soyinka JO, Onyeji CO, Omoruyi SI, Owolabi AR, Sarma PV, Cook JM. Pharmacokinetic interactions between ritonavir and quinine in healthy volunteers following concurrent administration. Br J Clin Pharmacol 2010; 69:3: 262–270.
	- *<https://doi.org/10.1111/j.1365-2125.2009.03566.x>*
- <span id="page-6-5"></span>33. Wenzel C, Drozdzik M, Oswald S. Organic Cation Transporter 1 an intestinal uptake transporter: Fact or Fiction? Front Pharmacol 2021; 12:648388. *<https://doi.10.3389/fphar.2021.648388>*
- <span id="page-6-6"></span>34. https://go.drugbank.com/categories/DBCAT003950. (Accessed April 2024)
- <span id="page-6-7"></span>35. https://go.drugbank.com/categories/DBCAT004549 (Accessed April, 2024)
- <span id="page-6-8"></span>36. van der Velden M, Bilos A, van den Heuvel JJM, *et al*. Proguanil and cycloguanil are organic cation transporter and multidrug and toxin extrusion substrates. Malar J 2017; 16:422, 1–12. *<https://doi.10.1186/s12936-017-2062-y>*
- <span id="page-6-9"></span>37. Muller F, Konig J, Glaeser H, *et al*. Molecular mechanism of renal tubular secretion of the antimalarial drug Chloroquine. Antimicrob Agents Chemother, 2011; 55(7): 3091–3098. *<https://doi:10.1128/AAC.01835-10>*
- <span id="page-6-10"></span>38. Louisa M, Suyatna FD, Wanandi SI, Asih PBS, Syafruddin D. Intrahepatic transport of primaquine with OCT1: An *in vitro* study in HepG2 cells. AIP Conf Proc 22019; 193: 040011. *<https://doi.org/10.1063/1.5139373>*
- <span id="page-6-11"></span>39. FDA. FDA briefing document. Tafenoquine tablet, 100 mg. Meeting of the Antimicrobial Drugs Advisory Committee (AMDAC). July 26, 2018. [November 11, 2019].
- <span id="page-6-12"></span>40. Hubeny A, Keiser M, Oswald S, *et al*. Expression of organic anion transporting polypeptide 1A2 in red blood cells and its potential impact on antimalarial therapy. Drug Metab Dispos 2016; 44:1562–1568. *<http://dx.doi.org/10.1124/dmd.116.069807>*
- <span id="page-6-13"></span>41. Ito S, Kusuhara H, Kuroiwa Y, *et al*. Potent and specific inhibition of mMate1-mediated efflux of type I organic cations in the liver and kidney by pyrimethamine. J Pharmacol Exp Ther 2010, 333 (1) : 341-350. *<https://doi.org/10.1124/jpet.109.163642>*
- <span id="page-6-14"></span>42. Patel M, Johnson M, Sychterz CJ, *et al*. Hepatobiliary disposition of atovaquone: A case of mechanistically unusual biliary clearance. J Pharmacol Exp Ther 2018; 366 (1): 37-45. *<https://doi.org/10.1124/jpet.117.247254>*