



RESEARCH ARTICLE

PREVALENCE OF *PSEUDOMONAS AERUGINOSA* (*P. AERUGINOSA*) AND ANTIMICROBIAL SUSCEPTIBILITY PATTERNS AT A PRIVATE HOSPITAL IN SANA'A, YEMEN

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Abstract



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Objective: *Pseudomonas aeruginosa* is clinically significant and opportunistic pathogen that causes infections in hospitalized patients. Antibiotic resistance is a major concern in clinical practice. The ongoing emergence of resistant strains that cause nosocomial infections contributes substantially to the morbidity and mortality of hospitalized patients. Objective of present study was to estimate the prevalence of *Pseudomonas aeruginosa* and the antimicrobial resistance patterns of *P. aeruginosa* isolates from hospitalized patients.

Methods: The study was performed at microbiology department of a local hospital in Sana'a, Yemen. All the patients' samples of hospital departments from January, 2017 to December, 2017 were included. A Total of 2079 samples were collected during the study period. Among them, 193 strains of *Pseudomonas spp.* were isolated. Antimicrobial susceptibility pattern of each isolates was carried out by the Kirby-Bauer disk diffusion method as per CLSI guidelines. Majority of *P. aeruginosa* were isolated from Sputum, followed by urine specimens.

Results: The isolate pathogen showed the highest sensitive to Meropenem (85.5%), followed by Amikacin (80.5%), Imipenem (80.0%), and Piperacillin/tazobactam (77.2). The highest frequency of resistance (96.2%) was observed with amoxicillin/clavulanic Acid followed by cefuroxime 94.6%, ampicillin/sulbactam 94.5%, Co-Trimoxzole 80.5%, and norfloxacin 54%.

Conclusion: The result confirmed the occurrence of drug resistance strains of *P. aeruginosa*. Meropenem, imipenem, and amikacin, were found to be the most effective antimicrobial drugs. It therefore calls for a very judicious, appropriate treatment regimens selection by the physicians to limit the further spread of antimicrobial resistance *P. aeruginosa*.

Keywords: Antimicrobial susceptibility, Imipenem, multi drug-resistant, *Pseudomonas aeruginosa*.

INTRODUCTION

Pseudomonas aeruginosa is clinically significant and opportunistic pathogen that causes infections in hospitalized patients. In addition, most *Pseudomonas* species have intrinsic resistance to many antibiotics and ongoing emergence of new resistance can be developed after commonly prescribed antimicrobial agents¹. *P. aeruginosa* has naturally resistant to many antibiotics due to the permeability barrier afforded by its outer membrane lipopolysaccharide (LPS). Only few antibiotics are effective against *Pseudomonas* and even these antibiotics are not effective against all strains². Antibiotic resistance is a major concern in clinical practice. The resistant strains of *P. aeruginosa*

that cause nosocomial infections contributes substantially to the morbidity and mortality of hospitalized patients³. Despite the availability of a variety of effective antimicrobial agents, treatment of *P. aeruginosa* is often challenging⁴ antimicrobial resistance is a growing problem worldwide, especially in hospitals, where resistant organisms are often first detected in ICUs⁵. The organism had been isolated from various infections like respiratory tract infections, cystic fibrosis, ear infections, orthopaedic infections, urinary tract infections, surgical infections, severe burns, etc. It was also reported frequently from patients undergoing chemotherapy for neoplastic diseases⁶. The variations of antibiotic protocols in clinics or in regions result in the different resistance profiles⁴. It is,

therefore, the goal of this study to determine the prevalence of *P. aeruginosa* isolates in a private hospital in Sana'a, Yemen also to evaluate its susceptibility against certain antibiotics, as limited work has been previously conducted on this subject.

METHODS

The study was performed at university of science and technology hospital in Sana'a, Yemen. It is one of the major private hospitals in Yemen. All the patients' samples from January, 2017 to December, 2017 were included. A Total of 2079 samples were gathered during the study period. Among them, 193 strains of *P. aeruginosa* were isolated. The medical records of these patients were retrieved and reviewed. All information regarding patients' gender and age as well as origin of clinical samples were collected.

Antimicrobial susceptibility testing of all the *P. aeruginosa* isolates was performed by Kirby-Bauer disk diffusion method and the result were interpreted by the Clinical Laboratory Standard Institute (CLSI) guidelines⁷. The antimicrobial susceptibility patterns of all the *P. aeruginosa* strains were determined against the following antibiotics of standard strength: ceftazidime, amikacin, gentamicin, imipenem, meropenem, ciprofloxacin, cefoperazone, piperacillin /tazobactam, amoxicillin/clavulanic acid, moxifloxacin, cefepime, ceftizoxime, ampicillin/ sulbactam, cefuroxime, ceftriaxone, co-trimoxzole, and levofloxacin. Full ethical clearance was obtained from the qualified authorities who approved the study design. All data were analyzed using SPSS Statistics 21. Data was presented in tables and graphs.

RESULTS

According to result findings, there were more than half of *P. aeruginosa* isolates in age group of 60 years and greater with 55(28%), followed by the age between 46 to 60 years in second rank about 38(20%), and finally the age between 31 to 45 years only about 20(10%). In this study, overall *P. aeruginosa* prevalence was 9.3 % (n=193/2079). The Figure 2 showed that there were about 154(80%) of *Pseudomonas* isolates form male, whereas the female had only about 39(20%). According to the study results, the medical department had the highest prevalence of *P. aeruginosa* isolates about 48(25%), followed by the intensive care unit in second rank about 41 (21%), the surgical department in third rank about 37(19%) and finally the pediatric and gynecology departments had only about 16(8%).

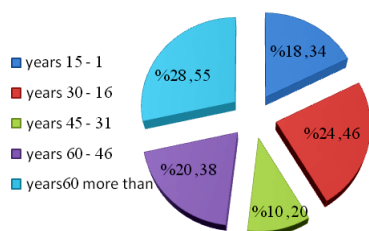


Figure 1: Distribution of *P. aeruginosa* isolates according to age.

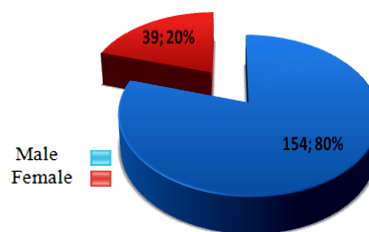


Figure 2: Distribution of *P. aeruginosa* according to gender.

The Figure 4 showed that the most of sample tests from sputum culture about 82(42.5%), followed by the sample from urine culture in second rank about 34(17.6%), and finally the sample test from other rout only about 6(3.1%).

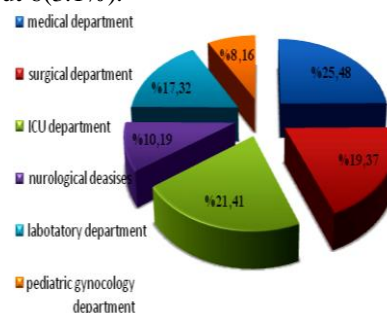


Figure 3: Distribution of *P. aeruginosa* isolates according to hospital departments.

According to the current study findings (Table 1), more than half of medication was sensitive to *P. aeruginosa* test about 12 drugs (54.5%), whereas the medication that resistance to *pseudomonas* tests about 10 drugs (45.5%).

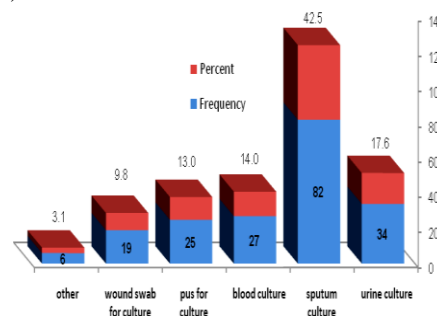


Figure 4: Distribution of *P. aeruginosa* isolates according to sample types.

P. aeruginosa strains showed resistance to ciprofloxacin 50.89%, ceftazidime 31.5%, ceftriaxone 78%, amoxicillin /clavulanic Acid 96.2%, ampicillin/ sulbactam 94.5%, cefuroxime 94.6%, nalidixic acid 83%, nitrofurantoin 88%, doxycycline 82.6%, norfloxacin 54%, and Co-Trimoxzole 80.5%. The highest frequency of sensitivity (85.5%) was observed with meropenem followed by amikacin 80.5%, imipenem 80%, piperacilline/tazobactam 77.2%, ceftizoxime 75%, ciprofloxacin 71.5%, levofloxacin 66%, cefoperazone 64%, gentamicin 56%, ceftazidime 54.5%, moxifloxacin 49%, and cefepime 44.5%. According to Figure 5 below, the highest resistance rate of anti-pseudomonal agent was with cefepime about 43.5% and the lowest resistance rate with

imipenem. Resistance to antipseudomonal drugs in current study was found to be cefepime (43.5%), ceftazidime (31.5%), ciprofloxacin (24%), piperacillin /tazobactam (16.5%), imipenim (15.4%). In the present study, multi drug resistance (MDR) rate (resistance to three or more of anti-*Pseudomonal* antimicrobials (i.e. piperacillin+tazobactam, imipenem, ceftazidime and amikacin) was determined to be 4.2% (8/193). Also MDR rate for only three anti *Pseudomonal* antimicrobials without imipenem was 4.2% (n=8/193) (i.e. piperacillin+tazobactam, ceftazidime and amikacin).

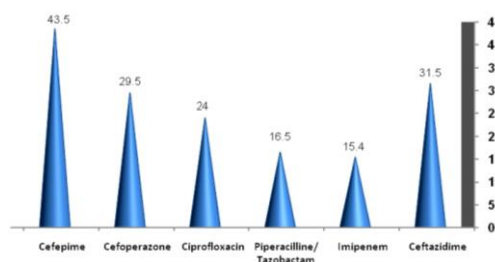


Figure 5: Resistance rates of anti-pseudomonal agent.

Table 1: Antimicrobial susceptibility patterns for *P. aeruginosa* isolates

Antibiotics	Expected options	Response		Antibiotics	Expected options	Response	
		F	%			F	%
Ceftriaxone	S	11	18.5	Ceftazidime	S	103	54.5
	R	46	78		R	60	31.5
	I	2	3.5		I	26	14
Cefoperazone sulbactam	S	58	64	Ciprofloxacin	S	118	71.5
	R	27	29.5		R	40	24
	I	6	6.5		I	7	4.5
Levofloxacin	S	108	66	Co-Trimoxzole	S	37	19.5
	R	44	26.8		R	152	80.5
	I	12	7.2		I	0	0.0
Ampicillin/sulbactam	S	2	3.7	Imipenem	S	150	80
	R	51	94.5		R	29	15.4
	I	1	1.8		I	9	4.6
Amoxicillin/Clavulanic Acid	S	4	2.1	Norfloxacin	S	10	38.5
	R	179	96.2		R	14	54
	I	3	1.7		I	2	8
Amikacin	S	152	80.5	Cefepime	S	83	44.5
	R	28	14.8		R	81	43.5
	I	9	4.7		I	22	12
Gentamicin	S	105	56	Meropenem	S	89	85.5
	R	65	35		R	10	9.5
	I	17	9		I	5	5
Moxifloxacin	S	77	49	Piperacillin/tazobactam	S	146	77.2
	R	69	44		R	31	16.5
	I	11	7		I	12	6.3
Cefuroxime	S	8	4.2	Ceftizoxime	S	1	4
	R	178	94.6		R	18	75
	I	2	1.2		I	5	21

DISCUSSION

P. aeruginosa has defined as one of the most common nosocomial pathogens. Hence we have undertaken this study to analyze the prevalence and antimicrobial susceptibility pattern of *P. aeruginosa* from various clinical samples of a private hospital. Periodic antimicrobial resistance monitoring in *P. aeruginosa* is fundamental to updating the current activity level of commonly used antipseudomonal drugs. The present study measures the rate of isolation of *P. aeruginosa* (n=193/2079; 9.3%) as which is lower than previous studies as by Tadv *et al.*,⁸ (22.67%) and Viren *et al.*⁹. The occurrence of *P. aeruginosa* was found to be higher in males, inpatients in age group >60 years and in surgery department, which is same as reported by Marzoqi *et al.*,¹⁰. This might be due to prolonged hospitalization and other associated co-morbidities in these age groups. The distribution of *P. aeruginosa* isolates specimens may vary with each hospital as each hospital and each health facility has a different

environment associated with it. According to the study results, more than 42.5% of the *P. aeruginosa* isolates were obtained from sputum samples. The distribution rank of the isolates according to the types of specimens was (respiratory sputum > urine > blood > pus > wound swap > others). Respiratory isolates (42.5%) were the most frequently encountered. *P. aeruginosa* isolates from respiratory tract as observed in a similar study of inpatient isolates done in a Saudi Arabian hospital¹¹. In the present study, the maximum clinical isolates of *P. aeruginosa* were isolated from medical department (25%), followed by ICU (21%) and surgical department (19%). This was similar to study of Pathmanathan SG¹². The distribution of specimens of *P. aeruginosa* might vary with each hospital as each hospital facility has a different environment associated with it. The correlation between specimen type and multidrug resistance would have been more noteworthy if supported by data on patients' clinical conditions. Prevalence of infection was higher in medical ward

followed by ICU as maximum isolates were isolated from sputum samples.

There was statistical significant relationship between the piperacilline/tazobactam susceptibility and sample types (p value=0.04). On other hand, there was no statistical significant relationship between the other antibiotics susceptibility (ceftazidime, imipenem, cefipeme) and sample types. As with this study, *P. aeruginosa* infection was primarily noted among older adults ($n=55$, 28%) particularly respiratory infection ($n=82$, 42.5%). There are a number of reasons why older adults are burdened by this type of infection. These include age-associated impairments in immunity that lead to reduced response to vaccination, a constellation of chronic and comorbid diseases, and functional limitations associated with advanced age. Additionally, older adults are at risk for aspiration pneumonia, outbreaks of gastroenteritis, recurrent urinary tract infection, and prosthetic device infections¹³. In the European Prevalence of Infection in Intensive Care (EPIC), *P. aeruginosa* was predominant gram-negative bacteria isolated from broncho pulmonary infections and accounts for 17% of health care-associated pneumonia and late-onset ventilate associated pneumonia¹⁴ and accounts for significant cases of cystic fibrosis¹⁵. The distribution of isolates differs with studies and clinical specimens¹⁶. Intensive care patients especially create an environment for infection because of the debilitating effect of a prolonged hospitalisation and the application of medical equipment (airways, catheters etc)¹⁷. ICUs are generally considered epicenters of antibiotic resistance and the principal sources of outbreaks of multi-resistant bacteria. The most important risk factors are excessive consumption of antibiotics exerting selective pressure on bacteria, the frequent use of invasive devices and relative density of a susceptible patient population with severe compelling diseases¹⁸. Thus, in ICUs, empirical antibiotic treatments should be avoided and treatment should be carried out using antibiotic susceptibility tests. ICUs should be regularly monitored resistance pattern against the various antibiotics. *P. aeruginosa* was responsible for pneumonia and septicaemia with deaths rate about 30% in immune compromised patients¹⁹. In the current study results, *P. aeruginosa* showed resistance to amoxicillin/ clavulanic Acid 96.2%, ampicillin/sulbactam 94.5%, cefuroxime 94.6%, nalidixic acid 83%, nitrofurantoin 88%, doxycycline 82.6%, ciprofloxacin 50.89%, ceftazidime 31.5%, ceftriaxone 78%, norfloxacin 54%, and cotrimoxzole 80.5%. However, the highest frequency of sensitivity (85.5%) was observed with meropenem followed by amikacin 80.5%, imipenem 80%, piperacilline/tazobactam 77.2%, ceftizoxime 75%, ciprofloxacin 71.5%, Levofloxacin 66%, Cefoperazone 64%, Gentamicin 56%, ceftazidime 54.5%, moxifloxacin 49%, and cefepime 44.5%. This may be explained by the fact that routine use of these antibiotics can lead to clinically significant resistance. One remarkable finding in the present study was the highest frequency of sensitivity (85.5%) was observed with meropenem, 85.5%, amikacin (80.5%), and piperacilline/ tazobactem (77.2%). These drugs were the

most effective drugs against *P. aeruginosa* infections. This similar to study finding by Taranasarwat *et al.*,²⁰ who reported highest sensitivity to imipenem. Also it was quite similar to the findings of Shaikh *et al.*, (100%)²¹ and Mohan *et al.*, (94.3%)²². One striking feature in this study was that all the *P. aeruginosa* isolates were found to be sensitive to imipenem. This may be due to the restricted use of imipenem in this hospital. This is consistent with a report published in 2002 in Mangalore, India²³. The emergence of carbapenem resistance is a serious concern²⁴. In various studies across the world, varying rates of resistance from 4-60% have been reported for imipenem and meropenem²⁵. Another survey found that resistance to imipenem was 19%, while other studies have reported low rates (5.8% and 9%) and high rates (38.6%) of resistance to imipenem²⁶. Piperacillin+ tazobactam showed a sensitive rate of 77.2 % in this study and cefoperazone-sulbactam showed a lower resistance of 29.5% only, indicating beta-lactamase inhibitor markedly expands the spectrum of activity of beta-lactams, which makes the combination drug the preferred choice against *P. aeruginosa* infections. Thus, emphasis should be given towards use of combined antibiotics in the treatment of *pseudomonal* infections²⁷. Bayani *et al.*, found that the resistance rate of *P. aeruginosa* to amikacin, ceftazidime, cefepime, imipenem, and ciprofloxacin was 53.3%, 43.3%, 40%, 40%, and 33.3%, respectively, and the prevalence of *P. aeruginosa* resistant isolates has increased²⁸. According to previous evidence, the rate of susceptibility was most productive for antimicrobial agent of class carbapenem against *P. aeruginosa*²⁹. Supported current results as 85.5% of strains were susceptible to Meronem and 80% to imipenem of class carbapenems. Although the resistance to carbapenems that include imipenem (16%) and meropenem (17.1) was low in this study, quite alarming should take into account that carbapenems are the last line of antibiotics for treating Gram-negative bacilli infections. Resistance to carbapenems may be due to a result of complex interactions of several mechanisms including production of carbapenemase, overproduction of efflux system and loss of outer membrane porins. *P. aeruginosa* isolates that are carbapenem resistant, specifically carbapenemase producing, are the worst, for the reason that they are associated with a higher mortality rate²⁴. Amikacin in this study was noted to be the most effective drug (80.5% sensitive). However, it is not commonly prescribed drug, because of its numerous side effects including renal toxicity, blurred vision, hearing loss, Bartter-like syndromes³⁰, neuromuscular blockade, arthralgia, and apnoea. In addition, ciprofloxacin (71.5% sensitive) proved to be within the most effective drugs for routine use among the *P. aeruginosa* strains investigated in this study. The result finding in this study was similar in a previous study finding that reported that amikacin had the highest sensitivity against *P. aeruginosa*⁹. Also in France, a higher susceptibility rate of 86% of amikacin was reported by Cavallo *et al.*,³¹. An earlier study reported from Kathmandu, Nepal³² shown amikacin (81.4% sensitive) and ciprofloxacin (70.3% sensitive)

among *P. aeruginosa* strains examined. Amikacin seems to be a promising therapy for *pseudomonal* infection. Hence, its use should be restricted to severe nosocomial infections, in order to avoid rapid emergence of resistant strains³³. However, high resistance to aminoglycosides had been reported in studies done in Bangladesh³⁴, Turkey⁴ and Malaysia³⁵. Similarly, higher rates of resistance to fluoroquinolones such as ciprofloxacin resistance (92%) were shown in a study from Malaysia³⁶. Also study findings by Zhanel *et al.*, reported moxifloxacin 58% and ciprofloxacin 46.7%³⁷. Recently, ceftazidime and cefepime are the most frequently prescribed third and fourth generation cephalosporins respectively. Ceftazidime is known antipseudomonal drug that has demonstrated high susceptibility pattern with *P. aeruginosa* isolates. The increased prevalence of ceftazidime resistant *P. aeruginosa* is related to the increased use of beta lactam antibiotics such as amoxicillin and ceftazidime. However, the resistance to cefadizime was reported as 31.5% in this study. This value of resistance was less than reported from Gujarat, with a resistance value of 75%⁹. *P. aeruginosa* strains in this study exhibited a high rate of resistance to the third generation cephalosporin drug such as ceftriaxone (78%). A much higher resistance to ceftriaxone of 75%, 86% and 93.9% had been reported in studies done in India³⁸ Bangladesh³⁴ and Nepal²⁷. Several studies have confirmed that *P. aeruginosa* was mostly resistant against ceftriaxone. However, this high level of resistance is not quite surprising as some suggest that ceftriaxone has considerably low activity against *P. aeruginosa*^{39,40}. Another study reported the following rates of resistance to cefepime 64.8%, piperacilline/tazobactam 45%, ciprofloxacin 38.9%, levofloxacin 36.1%, gentamicin 37.3% and amikacin 30%⁴¹. Relatively low piperacilline/tazobactam resistance (11.5%) had been reported in a hospital isolates of *P. aeruginosa* in a study from Saudi Arabia¹¹. In a study done in Kathmandu, Nepal²⁷, *P. aeruginosa* isolates obtained from intensive care unit of a national heart centre showed a high cefoperazone-sulbactam sensitivity rate of 84.8%. A previous study discovered an increased mortality rate associated with empiric piperacillin-tazobactam therapy given to patients with *P. aeruginosa* bacteraemia; the isolates had reduced piperacillin-tazobactam susceptibility⁴². In this study, amoxicillin /clavulanic acid had established 96.2 % resistance. Similarly, in a study conducted in Pakistan reported by Khan *et al.*,⁴³ had a high resistance rate of penicillin that is 98%; current findings are also in agreement with other studies as reported by Sasirekha *et al.*,⁴⁴ and Ullah *et al.*,⁴⁵ with respect to penicillin's. Also the same findings were obtained with amoxicillin/clavulanic acid (1.88%) and showed increasing resistance. Multi drug efflux pumps in the inner and outer membrane of *Ps. Aeruginosa* may protect the bacterium from β -lactam agents⁴⁶. Similar pattern had been reported in study in Nigeria⁴⁷. In addition, susceptibility to fourth-generation such as cefepime reported in India 32%⁴⁸ and in Bulgaria 42%⁴⁹ against *P. aeruginosa* isolates. The high resistance to cephalosporins may be due to production

of extended spectrum β -lactamases by the bacteria involved⁵⁰. Cefuroxime was one of the cephalosporin drugs tested in this study, with resistance value of 94.6%. This high resistance value observed were comparable with the report from Gujarat, India with resistance value of 73.2%⁹, but higher than reports from Malaysia of 40%⁵¹. In similar to previous study done in Bangladesh³⁴ showed rate of resistance for cotrimoxazole to be 93.5% in wound swab and pus isolates of *P. aeruginosa* while a Nigerian study⁵² showed *P. aeruginosa* isolates 100% resistant to cotrimoxazole. So, imipenem which is both an anti-pseudomonal drug and carbapenem was the best drug. According to the study findings, MDR rate (resistance to three or more of anti Pseudomonal antimicrobials (*i.e.* piperacillin+ tazobactam, imipenem, ceftazidime and amikacin) was determined to be 4.2% (8/193). Also MDR rate for only three anti Pseudomonal antimicrobials without imipenem was 4.2% (8/193) (*i.e.* piperacillin+ tazobactam, ceftazidime and amikacin). A study done by Unan *et al.*,⁵³ in Turkey reported rates of MDR, which were as high as 60%, whereas study done by Sabir *et al.*, in Pakistan detected lower rates of MDR (22.08%)⁵⁴. Moreover, the rate of current study was lower than a study done in Egypt, where Gad *et al.*,⁵⁵ observed 36% MDR *P. aeruginosa*. On comparing the sensitivity patterns of these antimicrobials, it was found that there was a considerable difference in the sensitivity pattern among these studies. This indicates that the sensitivity pattern changes from hospital to hospital and population to population. This has been possibly resulted from indiscriminate use of antibiotic, lack of awareness, patient non-compliance and unhygienic conditions⁵⁶. According to Berglund⁵⁷ one of the reasons for resistance among bacteria is a result of either overuse and misuse of antibiotics. The current study results indicated that *P. aeruginosa* was becoming resistant to commonly used antibiotics due to excessive consumption. The empirical antibiotic treatment should be limited and treatment should be carried out using antibiotic susceptibility test and efforts should be made to prevent spread of resistant bacteria⁵⁶.

CONCLUSIONS

In conclusion, results of the present study clearly demonstrated the occurrence of resistance to various antipseudomonal agents among the *P. aeruginosa* isolates. The statistics in this study showed low rates of antibiotic resistance to meropenem, amikacin, and meropenem, and piperacillin/ tazobactam and maximum sensitivity against *P. aeruginosa* strains. We suggest a more restricted and a more rational use of these drugs in hospital setting in order to avoid rapid emergence of resistant strains. Regular anti-microbial susceptibility monitoring is essential of local, regional and national level isolates. Every effort should be made to prevent spread of resistant organisms. The solution can be planned by continuous efforts of microbiologist, clinician, pharmacist and community to promote greater understanding of this problem. Frequent hand

washing to prevent spread of organism should be encouraged.

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AUTHOR'S CONTRIBUTION

Ali A: writing original draft, conceptualization. **Alhomidi AM:** methodology, investigation. **Al-Henhena N:** writing, review, and editing, supervision, resources. All authors read and approved the final manuscript for publication.

DATA AVAILABILITY

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

CONFLICT OF INTEREST

None to declare.

REFERENCES

- Kenneth T. Textbook of bacteriology: *Pseudomonas aeruginosa*. Wisconsin University, France 2004; 7-15.
- ALshaiki JMM, Toweir AA. Prevalance *Pseudomonas aeruginosa* Among Libyan Patients and its Association with Hospital's Environment in Benghazi. J Med Microb Diagn 2017; 6: 257. <https://doi.org/10.4172/2161-0703.1000257>
- Acar JF. Consequences of bacterial resistance to antibiotics in medical practice. Clin Infect Dis 1997; 24 (1):17 8. https://doi.org/10.1093/clinids/24.Supplement_1.S17
- Savas L, Duran N, Savas N, Onlen Y, Ocak S. The prevalence and resistance patterns of *Pseudomonas aeruginosa* in intensive care units in a university hospital. Turk J Med Sci 2005; 35:317-22.
- Carmeli Y, Troillet N, Eliopoulos GM, Samore MH. Emergence of antibiotic-resistant *Pseudomonas aeruginosa*: comparison of risks associated with different anti pseudomonal agents. Antimicrob Agents Chemother 1999; 43: 1379-82. PMID: 10348756
- Renuga S, Lakshmi K, Chitralkha S, Illamani V. Prevalence of *Pseudomonas aeruginosa* and its antibiotic susceptibility pattern in a Tertiary Care Hospital. Int J Res Pharm Sci 2015; 6(1), 27-30.
- Cockerill FR (Ed.), Performance Standards for Antimicrobial Susceptibility Testing: Twenty-first Informational Supplement, Clinical and Laboratory Standards Institute (CLSI), 2011.
- Tadvi J, Javadekar TB, Bhavsar R, Garala N. Prevalence and antibiogram of *Pseudomonas aeruginosa* at S.S.G. Hospital, Baroda, Gujarat, India. J Res Med Den Sci 2015; 3:204-207. <https://doi.org/10.5455/jrmds.20153310>
- Javiya VA, Ghatak SB, Patel KR, Patel JA. Antibiotic susceptibility patterns of *Pseudomonas aeruginosa* at a tertiary care hospital in Gujarat, India. Indian J Pharmacol. 2008; 40:230-4. <https://doi.org/10.4103/0253-7613.44156>
- Al-Marzoqi AH, Al Tae ZM. *Pseudomonas aeruginosa*: Antibiotic resistance pattern to different isolates in Al-Hillah city, Iraq. J Nat Sci Res 2013; 3:23-30. <https://doi.org/10.3389/fmicb.2016.00586>
- Al-Tawfiq JA. Occurrence and antimicrobial resistance pattern of inpatient and outpatient isolates of *Pseudomonas aeruginosa* in a Saudi Arabian hospital: 1998-2003. Int J Infect Dis 2007; 11:109-114. <https://doi.org/10.1016/j.ijid.2005.11.004>
- Pathmanathan SG, Samat NA, Mohamed R. Antimicrobial susceptibility of clinical isolates of *Pseudomonas aeruginosa* from a Malaysian Hospital. The Malaysian J Med Sci 2009; 16(2):27-32. PMID: 22589655
- Tortora G, Funke B, Case C. Microbiology: An Introduction, 6th ed.; Benjamin Cummings: California, CA, USA, 1998.
- Vincent JL, Bihari DL, Suter PM, Bruining HA, White J, Nicolas-ghanion M. The prevalence of nosocomial infection in intensive care units in Europe: results of the European Prevalence of Infection in Intensive Care (EPIC) study. J American Med Assoc 1995; 74:639-644. PMID: 7637145
- Pier GB. Role of cystic fibrosis transmembrane conductance regulator in innate immunity to *Pseudomonas aeruginosa* infections. Proceedings of National Academy of Science, USA 2000; 97:8822-8828. <https://doi.org/10.1073/pnas.97.16.8822>
- Okon KO, Agukwe PC, Oladosu W, Balogun ST, Uba A. Antibiotic resistance pattern of *Pseudomonas aeruginosa* isolated from clinical specimens in a tertiary care hospital in Northeastern Nigeria. Internet J Microbiol 2010;8:1-6 https://doi.org/10.4103/ijmr.IJMR_14_18
- Jarlier V, Fosse T, Philippon A. Antibiotic susceptibility in aerobic gram-negative bacilli isolated in intensive care units in 39 French teaching hospitals (ICU study). Intensive Care Med 1996; 22: 1057-65. <https://doi.org/10.1086/647284>
- Weber DJ, Raasch R, Rutala WA. Nosocomial infections in the ICU: the growing importance of antibiotic-resistant pathogens. Chest 1999; 115: 34S-41S. https://doi.org/10.1378/chest.115.suppl_1.34S
- Olayinka AT, Onile BA, Olayinka BO. Prevalence of multidrug resistant (MDR) *Pseudomonas aeruginosa* isolates in Surgical Units of Ahmadu Bello University Teaching Hospital, Zaria, Nigeria: an Indication for Effective Control Measures Annals of African Medicine. 2004; 3 (1):13-16.
- Sarwat T, Rashid M, Rastogi V, Chander Y. A comparative study of Antibiogram of *Pseudomonas aeruginosa* in Hospital and community acquired infections. Int J Curr Microbiol App Sci 2015; Special Issue-1: 286-291 <https://doi.org/10.1016/j.sjbs.2014.06.0>
- Shaikh S, Fatima J, Shakil S, Mohd S, Rizvi D, Kamal MA. Prevalence of multidrug resistant and extended spectrum beta-lactamase producing *Pseudomonas aeruginosa* in a tertiary care hospital. Saudi J Biol Sci 2015; 22, 62-64 <https://doi.org/10.1289/EHP292>
- Mohan BS, Lava R, Prashanth HV, Nambiar V, Metri B, Nayak V, Sri Krishna R. Prevalence and antibiotic sensitivity pattern of *Pseudomonas aeruginosa*; An emerging nosocomial pathogen. Int J Biol Med Res 2013; 4(1): 2729-2731. <https://doi.org/10.1155/2011/605195>
- Shenoy S, Baliga S, Saldanha DR, Prashanth HV. Antibiotic sensitivity patterns of *Pseudomonas aeruginosa* strains isolated from various clinical specimens. Indian J Med Sci 2002; 56(9):427-30. <https://doi.org/10.4103/0253-7613.44156>
- Liu Q, Li X, Li W. Influence of carbapenem resistance on mortality of patients with *Pseudomonas aeruginosa* infection: a meta-analysis. Sci Rep. 2015; 5, 11715. <https://doi.org/10.1038/srep11715>
- Gonlugur U, Bakici MZ, Akkurt I, Efeoglu T. Antibiotic susceptibility patterns among respiratory isolates of Gram-negative bacilli in a Turkish University hospital. BMC Microbiol 2004; 4: 32. <https://doi.org/10.1186/1471-2180-4-32>
- Khan M.A., Faiz A. Antimicrobial resistance patterns of *Pseudomonas aeruginosa* in tertiary care hospitals of Makkah and Jeddah. Ann Saudi Med 2016; 36: 23-28. <https://doi.org/10.5144/0256-4947.2016.23>

27. Bhandari S, Banjara MR, Lekhak B, Bhatta DR, Regmi SR. Multi-drug and pan-drug resistant *Pseudomonas aeruginosa*: a challenge in post-antibiotic era. Nepal J Sci Tech 2012; 13(2):197-202. <https://doi.org/10.3126/njst.v13i2.7736>
28. Bayani M, Siadati S, Rajabnia R, Taher AA. Drug Resistance of *Pseudomonas aeruginosa* and Enterobacter cloacae Isolated from ICU, Babol. Northern Iran Int J Mol Cell Med 2012; 2: 204-209. PMID: 24551814
29. Turner PJ. Meropenem and imipenem activity against *Pseudomonas aeruginosa* isolates from the MYSTIC Program, Diagnostic Microbiology and Infectious Disease 2006; 56 (3): 341-344. <https://doi.org/10.1016/j.diagmicrobio.2006.07.015>
30. Juayang AC, Maestral DG Jr, Gallega CT, et al. Review on the antimicrobial resistance of pathogens from tracheal and endotracheal aspirates of patients with clinical manifestations of pneumonia in Bacolod City in 2013. Int J Bacteriol 2015; 2015:942509. <https://doi.org/10.1155/2015/942509>
31. Cavallo JD, Hocquet D, Plesiat P, Fabre R, Roussel-Delvallez M. Susceptibility of *Pseudomonas aeruginosa* to antimicrobials: a 2004 French multicentre hospital study. J Antimicrob Chemother 2007; 59, (5): 1021-1024. <https://doi.org/10.1093/jac/dkm076>
32. Koirala P, Bhatta DR, Ghimire P, Pokhrel BM, Devkota U. Bacteriological profile of tracheal aspirates of the patients attending a neuro-hospital of Nepal. Int J Life Sci 2010;4:60-65. <https://doi.org/10.1155/2013/847569>
33. Poole K. Aminoglycosides resistance in *Pseudomonas aeruginosa*. Antimicrob Agents Chem 2005; 49:479-87. <https://doi.org/10.1128/AAC.49.2.479-487.2005>
34. Rashid A, Chowdhury A, Rahman SHZ, Begum SA, Muazzam N. Infections by *Pseudomonas aeruginosa* and antibiotic resistance pattern of the isolates from Dhaka Medical College Hospital. Bangladesh J Med Microbiol 2007; 1(2):48-51. <https://doi.org/10.1186/s13104-015-1497-x>
35. Fazlul MKK, Zaini MZ, Rashid MA, Nazmul MHM. Antibiotic susceptibility profile s of clinical isolates of *Pseudomonas aeruginosa* from Selayang Hospital, Malaysia. Biomed Res 2011; 22(3):263-66. PMID: 22589655
36. Al-KabsiAM, Yusof MYBM, Sekaran SD. Antimicrobial resistance pattern of clinical isolates of *Pseudomonas aeruginosa* in the University of Malaya Medical Center, Malaysia. Afr J Microbiol Res 2011; 5(29):5266-72. <https://doi.org/10.5897/AJMR11.284>
37. ZhanelG G, LaingN M, Nichol K. A. Antibiotic activity against Urinary Tract Infection (UTI) isolates of Vancomycin Resistant Enterococci (VRE): results from the 2002 North American vancomycin resistant enterococci susceptibility study (NAVRESS). J Antim Chem 2003; 52 (3): 382-388. <https://doi.org/10.1093/jac/dkg352>
38. Arora D, Jindal N, Kumar R, Romit. Emerging antibiotic resistance in *Pseudomonas aeruginosa*. Int J Pharm Pharm Sci 2011; 3(2):82-4. <https://doi.org/10.12688/f1000research.19509.1>
39. Bassetti D, Cruciani M, Solbiati M, Rubini F, Gandola L, Valenti G, et al. Comparative efficacy of ceftriaxone versus ceftazidime in the treatment of nosocomial lower respiratory tract infections. Chemotherap 1991; 37:371-5. 20. <https://doi.org/10.1128/JCM.00893-06>
40. Mody L, Bradley SF, Strausbaugh LJ, Muder RR. Prevalence of ceftriaxone and ceftazidime resistant gram-negative bacteria in long term-care facilities. Infect Control Hosp Epidemiol 2001; 22(4):193-4. <https://doi.org/10.1086/503397>
41. Dash M, Padhi S, Narasimham MV, Pattnaik S. Antimicrobial resistance pattern of *Pseudomonas aeruginosa* isolated from various clinical samples in a tertiary care hospital, South Odisha, India. Saudi J Health Sci 2014; 3, 15-19. <https://doi.org/10.4103/2278-0521.130200>
42. Tam VH, Gamez EA, Weston JS, Gerard LN, LaRocco MT, Caeiro JP, et al. Outcomes of Bacteremia due to *Pseudomonas aeruginosa* with Reduced Susceptibility to Piperacillin-Tazobactam: Implications on the appropriateness of the resistance breakpoint. Clin Infect Dis 2008; 46:862-867. <https://doi.org/10.1086/528712>
43. Khan JA, Iqbal Z, Ur Rahman S, Farzana K, Khan A. Prevalence and resistance pattern of *Pseudomonas aeruginosa* against various antibiotics. Pakistan J Pharm Sci.2008; 21(3):311-315.
44. Sasirekha B, Manasa R, Ramya P, Sneha R. Frequency and antimicrobial sensitivity pattern of extended spectrum β -lactamases producing *E. coli* and *Klebsiella pneumoniae* isolated in a tertiary care hospital. J Medical Sci 2010; 3(4): 265-271.
45. Ullah F, MalikS A, Ahmed J. Antimicrobial susceptibility and ESBL prevalence in *Pseudomonas aeruginosa* isolated from burn patients in the North West of Pakistan. Burns 2009; 35 (7): 1020-1025. <https://doi.org/10.1016/j.burns.2009.01.005>
46. Srikumar R, Li XZ, Poole K. Inner membrane efflux components are responsible for β -lactam specificity of multi drug efflux pumps in *Pseudomonas aeruginosa*. J Bacteriol 1997; 179(2): 7875-7881. <https://doi.org/10.1128/jb.179.24.7875-7881.1997>
47. Jombo GTA, Jonah P, Ayeni JA. Multidrug Resistant *Pseudomonas aeruginosa* in Contemporary Medical Practice: Findings from urinary isolates at a Nigerian University teaching hospital. Nigerian J Phys Sci 2008; 23(1-2):105-109. <https://doi.org/10.4314/njps.v23i1-2.54944>
48. Chaudhury A. *In vitro* activity of Cefpirome: a new fourth generation cephalosporin. Indian J Med Micro 2003; 21 (1): 52-55. PMID: 10992706
49. Strateva T, Ouzounova RV, Markova B, Todorova A, Marteva-ProevskaY, Mitov I. Widespread detection of VEB-1-type extended-spectrum beta-lactamases among nosocomial ceftazidime-resistant *Pseudomonas aeruginosa* isolates in Sofia, Bulgaria. J Chem 2007; 19 (2): 140-145. <https://doi.org/10.1179/joc.2007.19.2.140>
50. Mathur P, Kapil A, Das B, Dhawan B. Prevalence of extended spectrum β -lactamase producing gram negative bacteria in a tertiary care hospital. Indian J Med Res 2002; 115(2): 153-157. <https://doi.org/10.7860/JCDR/2013/6460.3462>
51. Nwankwo EOK, Shuaibo SA. Antibiotic susceptibility pattern of clinical isolates of *Pseudomonas aeruginosa* in a tertiary health institution in Kano, Nigeria J Med Biomed Sci 2010; 37-40. PMID: 22589655
52. Lim KT, Yasin RY, Yeo CC. Genetic fingerprinting and antimicrobial susceptibility profiles of *Pseudomonas aeruginosa* hospital isolates in Malaysia. J Microbiol Infectious Diseases 2009; 42:197-209. PMID: 19812853
53. Unan D, Gnsereen F. The resistance of *P. aeruginosa* strains isolated from nosocomial infections against various antibiotics. Mikrobiyol Bult. 2000; 34: 255-60 <https://doi.org/10.1128/CMR.00040-09>
54. Sabir R, Alvi SFD, Fawwad A. Antimicrobial susceptibility pattern of aerobic microbial isolates in a clinical laboratory in Karachi- Pakistan. Pak J Med Sci 2013; 29(3): 851-5 <https://doi.org/10.12669/pjms.293.3187>
55. Gad GF, El-Domany RA, Zaki S, Ashour HM. Characterization of *Pseudomonas aeruginosa* Isolated from Clinical and Environmental Samples in Minia, Egypt: Prevalence, Antibiogram and Resistance Mechanisms. J Antimicrob Chemother 2007; 60: 1010-7. <https://doi.org/10.1093/jac/dkm348>
56. Parmar H, Dholakia A, Vasavada D, Singhala H. The current status of antibiotic sensitivity of *Pseudomonas aeruginosa* isolated from various clinical samples. Int J Res Med 2013; 2(1): 1-6. <https://doi.org/10.4103/0253-7613.44156>
57. Berglund B. Environmental dissemination of antibiotic resistance genes and correlation to anthropogenic contamination with antibiotics. J Infect Ecol Epidemiol 2015; 5: 28564. <https://doi.org/10.3402/iee.v5.28564>