

RESEARCH ARTICLE

EVALUATION OF ANTICOCCIDIAL ACTIVITY OF METHANOLIC EXTRACT OF RICINUS COMMUNIS IN BROILER

Rahma Hamayun¹, Muhammad Fazeel²

¹Northern Border University, Kingdom of Saudi Arabia. ²University of Agriculture, Pakistan.

Article Info:

Cite this article:

+0547282585.

Article History:

Hamayun R, Fazeel M. Evaluation of

anticoccidial activity of methanolic extract of

Ricinus communis in broiler. Universal Journal

Dr. Rahma Hamayun, Northern Border

University, Kingdom of Saudi Arabia, Tel-

of Pharmaceutical Research 2021; 6(4):26-31.

https://doi.org/10.22270/ujpr.v6i4.637

*Address for Correspondence:

E-mail: r_hamayun@hotmail.com

Received: 3 June 2021 Reviewed: 6 July 2021

Accepted: 10 August 2021

Published: 15 September 2021



Aims and Objectives: Coccidiosis is recognized as the parasitic disease which has the greatest economic impact on poultry production. The emergence of resistant strains to available drugs has become a major problem in order to treat/control coccidiosis. Botanicals can act as alternative to anticoccidial drugs. This study has therefore, been planned to evaluate both *in vitro* and *in vivo* anticoccidial activity of the plant *R. communis*.

Methods: In *in vitro* trial 20, 10, 5, 2.5, 1.25and 0.625percent, DMSO dissolved crude aqueous methanolic extract of *R. communis* was used to investigate its inhibitory effect upon sporulation of oocysts. The research was done in University of Agriculture, Faisalabad, Pakistan, with the collaboration of Parasitology, Pathology and Poultry departments. In *in vivo* trial, the plant was used at three graded concentrations for evaluation of its anticoccidial activity in broiler birds. A total of 144 (one-day-old) broiler chicks were divided into six groups each having 24 chicks. At age of 15 days, groups I, II and III were given 4%, 5% and 6% of dried powder of *R. communis* respectively. Group IV was served as positive control (infected, toltrazuril treated), group V as negative control (infected, non-medicated) and group VI was serve as non-infected and non-medicated control. All groups except group VI were infected orally with 50,000 sporulated oocysts of mixed *Eimeria* species at 18th day of age.

Result: After 7 days of inoculation, six birds from each group were slaughtered to get results on oocysts score, lesion score, relative organ weight, hematology and immunomodulatory effect. Data was analyzed by using analysis of variance (ANOVA) and group means were compared by Duncan's multiple range tests.

Conclusions: It was concluded that *R. communis* can be used as a prophylactic and therapeutic agent at local and regional level for the control of coccidiosis in broilers.

Keywords: anticoccidial, immunomodulatory, *Ricinus communis*, sporulated oocysts.

INTRODUCTION

Commercial poultry farming is one of the most flourishing industries of the world as it provides the cheapest source of animal protein to humans¹. Poultry sector generates employment (direct/indirect) for about 1.5 million people and recorded a rapid growth of 7-8% in Pakistan during the year 2012-13. Its share is 26.8 percent in the total meat production of the country. However some viral, bacterial and parasitic diseases involving the Gastro intestinal tract of birds possesses a great threat to poultry industry². Coccidiosis is caused by apicomplexan parasites of genus *Eimeria*³, which inhabits and colonies the intestinal mucosa⁴. There are about 1800 *Eimeria* species which effect the intestinal mucosa of different animals and birds⁵, however in poultry, nine different *Eimeria* spp. are recognized⁶ in which *E. brunette*, *E. maxima*, *E. necatrix* and *E. tenella* are extremely pathogenic, while *E. acervulina*, *E. mitis*, *E. mivati*, *E. praecox* and *E. hagani* are comparatively less harmful^{7,8}. *Eimeria tenella* is most destructive among other *Eimeria* spp. and cause caecal coccidiosis⁹. *Eimeria tenella* sporozoites, through villi of epithelial cells invades caecal mucosa and cause severe damage to epithelium, blood in feaces, reduced weight gain and feed efficiency and ultimately death of birds⁹. The repeated use of synthetic chemicals and anticoccidial feed additives have not only provoked anticoccidial

ISSN: 2456-8058

26

drug resistance in *Eimeria* spp¹⁰, but also have detrimental effects on health of birds and humans¹¹. The use of live vaccine to control coccidiosis is a good alternate however it may lead to development of clinical disease in the broilers under poor management¹². The ineffectiveness of anticoccidial drugs and vaccines and emergence of resistant *Eimeria* spp. have forced the scientists to sort out the alternatives for control of disease^{13,14}. The effect of plants and herbal products has been reported by various authors, during the last decade against experimental coccidial infection in birds^{15,16}.

Reason for selection of this plant

R. communis also known as castor plant belongs to family Euphorbiaceae. It is a widely used and potent medicinal plant amongst all the thousands of medicinal plants. The leaves, roots and seed oils of this plant have been used for the treatment of inflammation, liver disorders and hypoglycemia^{17,32}. The stem of *R*. communis have anticancer, anti-diabetic and anti activity³⁸. The protozoal plant has various phytochemical constituents like flavonoids, saponins, glycosides, alkaloids and steroids^{42,44} which are responsible for its activity. The review of literature has proven its efficacy as an antibacterial, antiprotozoal and an antifungal agent³³⁻⁴⁷. However the anticoccidial activity of R. communis has not been evaluated up till now. Therefore, the present study has been designed to evaluate the anticoccidial activity of R. communis and its effect on immunomodulation and hematology in broiler chicken.

MATERIALS AND METHODS

Plant materials

The plant *R. communis* was procured from the local market of Faisalabad, Pakistan in the month of March. It was identified and authenticated by Herbarium of Department of Botany, University of Agriculture, Faisalabad, Pakistan. The plant specimen was kept in the Ethno veterinary Research and Development Centre, Department of Parasitology, UAF as voucher No. 0170. The leaves and the seeds were dried under shade and ground finely to powder in an electric mill. Powdered plant materials were extracted with methanol in a Soxhlet's apparatus at 80°C. The crude methanolic extract (CME) was evaporated in a rotary evaporator, under reduced pressure at $35^{\circ}C^{18}$. The CME will further be dried by using freeze dryer and then stored at $4^{\circ}C$ until used.

Collection of coccidial oocysts

Guts were opened and contents thus collected from intestines were examined microscopically. Coccidial oocysts were extracted following the method described by Ryley¹⁹.

Sporulation of oocysts

The contents of the positive samples were placed in 2.5% potassium dichromate solution. The petri dishes were partially covered to allow the passage of oxygen and incubated at $25-29^{\circ}$ C for 48 hours, providing 60-80% humidity¹⁹.

Isolation of the sporulated oocysts

The sporulated oocysts were separated by zinc sulphate floatation technique¹⁹. The counting of washed sporulated oocysts was done by McMaster technique²⁰. The required concentration of the sporulated oocysts (50,000/ml) was maintained with phosphate buffered saline.

Experimental Design

The experiment was conducted in two phases.

- 1. Sporulation inhibition Assay
 - 2. *In vivo* trials to evaluate the dose dependent anticoccidial effect of the *R. communis*

Sporulation Inhibition Assay

Oocysts were exposed to six concentrations of plant extract (w/v; 20, 10, 5, 2.5, 1.25 and 0.625 %)^{32-36,38-43}. Three replications were made for each concentration and the whole experiment was repeated to confirm the results. The experimental design used in the present study was approved by Department of Parasitology, University of Agriculture, Faisalabad review board, in accordance with approved published research ethics guidelines³⁹. An *in vitro* sporulation inhibitionassay²¹ was used to evaluate the effect of plant extracts on the sporulation of coccidial oocysts. In this assay, the unsporulated oocysts were incubated with plant extracts for 48 h at 25-29°C²¹. The number of sporulated and non-sporulated oocysts was counted and the percent sporulation was estimated by counting the number of sporulated oocysts in a total of 40 oocysts. The oocysts with 4 sporocysts was considered sporulated regardless the shape and size of the sporocysts. The oocysts were slightly flattened under the pressure of a cover slip to better illustrate morphology.

In vivo trial

In *in vivo* trial⁴⁵, 144 (day-old) broiler chicks were procured from local market. At 15th day of age, the chicks were randomly divided into six groups, each group having 24 chicks⁴⁶, and three graded doses of plant, were added in the feed²³. At 18th day of age, the chicks of all groups except group VI were inoculated sporulated oocysts (50,000/chick) of mixed *Eimeria* species. The detailed experimental lay out is as follows:

Group I, II and III: were administrated with 3 graded doses of plant

Group IV: were medicated with toltrazuril

Group V: were kept as infected and non-medicated control

Group VI: were kept as non-infected and non-medicated control

Parameters

Data on the following parameters was recorded i.e. weight gain of birds Feed consumption, FCR, effect on relative weight of organs, lesion scoring, oocyst scoring, fecal scoring, Hematological tests, serum chemistry and immunological evaluation.

Statistical Analysis

One way analysis of variance (ANOVA) and Duncan's multiple range tests was used for determination of statistical significance (p < 0.05). Data showing only percentages such as survival percentage were analyzed using the Chi square test.

RESULTS

Results of *in vitro* experiment:

At concentrations (20%, 10%, 5%, 2.5%, 1.25% and 0.63%), *R. communis* inhibited the sporulation 20%, 40%, 40%, 55%, 60% and 70% respectively, as

compared to C-I and C-II, which showed 80% and 86.25% sporulation respectively. All dilutions of *R*. *communis* significantly inhibited the sporulation (p<0.0001) in all *Eimeria* species as compared to both control as showed by the statistical analysis to both control.

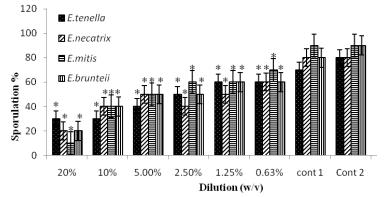


Figure 1: Effect of R. communis on % sporulation of oocysts.

Figure 1, shows effect of methanolic extract of *R*. *communis* extract on % sporulation at different dilution levels in 10 % DMSO solution. Results are the mean and standard error of means. p<0.0001, level of significance of the inhibitory effect and p<0.0005, level of significance of the damage oocysts, both compared with the untreated control groups.

Figure 2 shows the effect of methanolic extract *R. communis* extract on % damage of oocysts at different dilution levels in 10% DMSO solution. Results are the mean and standard error of means. *p<0.0001, level of significance of the inhibitory effect and *p<0.0005, level of significance of the damage Oocysts, both compared with the untreated control groups.

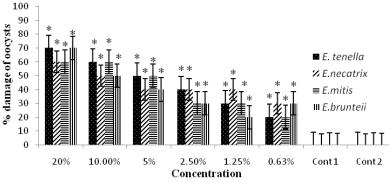


Figure 2: Effect of *R. communis* on % damage of oocysts.

The results (Table 1) Show that maximum survival (94.45%) and minimum mortality (5.55%) values was observed in 6% *R. communis* treated group among different concentrations. Highest 33.33 % mortality was observed in infected non-medicated control group. The results (Figure 3) show that the significantly better (p<0.05) weight gain shown by 6% *R. communis*

treated group during first, second and third week post infection as compared to infected unmedicated group. Although all doses showed better weight gain as compared to infected non-medicated group. Lymphoproliferative response to Phytohemagglutinin-P(PHAP) in experimental and control chickens at 24, 48 and 72hrs.

 Table 1: Effect of R. communis 4%, 5% and 6% treatment on mortality and survival % in broiler chicks artificially infected with mixed Eimeria species.

Groups	Mortality Days post inoculation					Total	% Mortality	% Survival
	3	4	5	6	7	Mortality	10101 tunity	Jui /Ivui
R. communis 4%	_	1	2	-	_	3	16.66	83.34
R. communis 5%	-	1	1	-	_	1	11.11	88.89
R. communis 6%	-	-	1	_	_	1	5.55	94.45
IM	-	-	2	_	_	2	16.66	83.34
INM	-	3	3	-	_	6	33.33	66.67
NN	_	0	0	_	_	0	0	100%

IM: Infected medicated group; INM: Infected non -medicated group; NN: Non-infected non- medicated group.

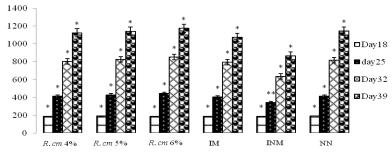


Figure 3: Effect of *R. communis* 4%, 5% and 6% treatment on means (n=24) weight gain at different weeks of post infection in broiler chicks artificially infected with mixed *Eimeria* species.

IM: Infected medicated Group; INM: Infected non-medicated Group; NN: Non-infected non-medicated Group

There was no significant difference in pre-PHAP infection cellular response in all groups at 24, 48 and 72hrs. *** and *** show that there is no significant

difference (p>0.05) in all groups. The results are the means and standard error of means (Figure 4).

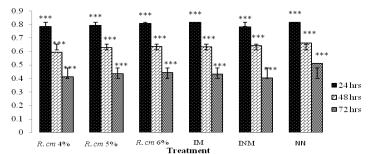


Figure 4: Lympho-Proliferative response to PHAP-Pre infection in *R. communis* 4%, 5% and 6% treated broiler hicks artificially infected with mixed *Eimeria* species. IM: Infected medicated Group; INM: Infected non-medicated Group; NN: Non-infected non-medicated Group

Total antibody titers at 7 days of PPI were nonsignificantly different (p>0.05) in all groups at 7 and 14 days of PSI these titers were non-significantly different (p>0.05) in *R. communis* 4, 5 and 6% but, were significantly different (p<0.05) to INM groups (Figure 5).

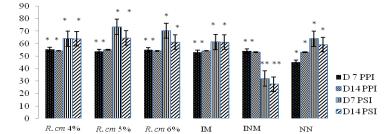


Figure 5: Total anti-SRBCs (sheep red blood cells) antibody titer in R. communis 4, 5 and 6% treated broiler

chicks artificially infected with Eimeria species.

PPI: Post primary injection; PSI: Post-secondary injection; IM: infected medicated; INM: infected non-medicated;

NN: non-infected non-medicated

DISCUSSION

Avian coccidiosis is a vital parasitic disease that has caused considerable economic losses throughout the world. An estimated annual economic loss is about \$800 million worldwide for the poultry industry due to commercial losses. The development of drug resistant parasites and failure of chemotherapeutic agents have forced the scientists to sort out alternative approaches for disease control. A large amount of human population relies on the use of plant based medicinal products to combat ailments and they are proved to be very effective as so far no resistance has been developed against them. Recently, a significant number

of scientific publications have demonstrated the potential benefit of different chemicals of plant origin against avian coccidiosis²³⁻²⁶. Castor oil (R. communis) is a very potent medicinal plant. It is found all over the world, both in tropical and temperate regions. The stem, roots, leaves and seeds of this plant have antiinflammatory, anti-oxidant, antimicrobial and anthelmintic properties. The oil has proven its efficacy against intestinal inflammation and is used as laxative also. Stem of R. communis have anticancer, antidiabetic and antiprotozoal activity28. The plant has various phytochemical constituents like flavonoids, saponins, glycosides, alkaloids, steroids and tannins. Immunostimulatory effect of R. communis was

evaluated by using Phytohemagglutinin-P (PHA-P) to detect cell-mediated immunity and antibody response. Sheep red blood cells (SRBCs) were used to detect the humoral immunity. Results of study showed that supplementation of *R. communis* at the rate of 6% per kilogram of feed improved the cellular and humoral immunity against infection of mixed *Eimeria* species in chickens. The similar immunostimulatory results on anticoccidial activity of different plants were also reported by previous studies^{28,28,30,46}. Keeping in view the above characteristics of *R. communis*, the current study was designed to evaluate the immunomodulatory and anticoccidial potential of this plant by both *in vitro* and *in vivo* methods.

In in vitro trial 20, 10, 5, 2.5, 1.2525 and 0.625 percent, DMSO dissolved crude aqueous methanolic extract of R. communis inhibited the sporulation of coccidian oocysts in dose dependent manner. In in vivo trial efficacy of R. communis was evaluated at three different concentrations 4, 5 and 6% respectively. Better anticoccidial results were observed in 6% R. communis treated group with improved weight gain, better FCR, reduced oocyst and lesion score (P > 0.05). R. communis also enhanced cellular and humoral immune response in broiler chickens. Total antibody titers and immunoglobulin's IgG and IgM were elevated. These antibody and immunoglobulin's titers were high in 6% R. communis treated group in broiler chickens artificially infected with mixed Eimeria species. R. communis increased the weight gain and feed consumption efficiency, however the feed conversation ratio decreased as compared to the infected non-medicated control. In coccidiosis, weight of liver, kidney and intestine increased, however, weight of heart, spleen, bursa, gizzard and proventriculus were similar in all groups. In plant treated groups, there is no increased weight gain of liver, kidney and intestine. R. communis decreased the blood in feaces at 4th, 5th and 6th day post inoculation of oocysts. Additionally, lesion score and oocysts score decreased at 7th day post inoculation of sporulated oocysts as compared to the infected non-medicated control group. Plant decreased the effect of coccidiosis in broilers. Whenever, damage in liver and kidney occurred, it affects their functions. The values of ALT, ASAT, LDH, urea and creatinine were lower in plant treated groups as compared to other three groups, which show that this plant has anticoccidial potential.

CONCLUSIONS

It was concluded from current study that *R. communis* when used up to 6% dose rate in feed produces anticoccidial effects in terms of improved body weight gain, better FCR, reduced intestinal lesion and fecal score. It has positive impact on hematological parameters such as Hb, PCV, RBCs and WBCs count and also enhances cellular and humoral immune response against coccidiosis in broiler chickens. *R. communis* can be used as prophylactic and therapeutic agent at local and regional level for the control of coccidiosis in broilers.

ACKNOWLEDGEMENTS

The authors extend their thanks and appreciation to the Northern Border University, Kingdom of Saudi Arabia to provide necessary facilities for this work.

AUTHOR'S CONTRIBUTION

Hamayun R: study design, writing original draft. **Fazeel M:** literature survey, critical review. The final manuscript was read and approved by all authors.

DATA AVAILABILITY

Data will be made available on reasonable request.

CONFLICT OF INTEREST

None to declare.

REFERENCES

- 1. Ahmad F, Haq AU, Ashraf G, Abbas, Siddiqui MZ. Effect of different light intensities on the production performance of Broiler chickens. Pak Vet J 2011; 31: 203-206.
- Hafez HM. Enteric Diseases of Poultry with Special Attention to Clostridium perfringens. Pak Vet J 2011; 31: 175-184.
- Williams RB, Carlyle WW, Bond DR, Brown IA. The efficacy and economic benefits of Paracox, a live attenuated anticoccidial vaccine, in commercial trials with standard broiler chickens in the United Kingdom. J Parasitol 1999; 29: 341-355. https://doi.org/10.1016/s0020-7519(98)00212-4
- Hadipour MM, Olyaie A, Naderi M, Azad F, Nekouie O. Prevalence of *Eimeria* species in scavenging native chickens of Shiraz, Iran. African J Micro Res 2011; 5:3296-3299. https://doi.org/10.5897/AJMR11.477
- Haug A, AG Gjevre, P Thebo, JG Mattsson, M Kaldhusdal. Coccidial infections in commercial broilers: epidemiological aspects and comparison of *Eimeria* species identification by morphometric and polymerase chain reaction techniques. Avian Pathol 2008; 37: 161-70. https://doi.org/10.1080/03079450801915130
- Morgan JA, Morri GM, Wlodek BM, et al. Real-time polymerase chain reaction (PCR) assays for the specific detection and quantification of seven *Eimeria* species that cause coccidiosis in chickens. Mol Cell Probes 2009; 23: 83-89. https://doi.org/10.1016/j.mcp.2008.12.005
- Moghaddam GH, Pourabad RF. Prevalence of *Eimeria* species among broiler chicks in Iran. Mun Ent Zool 2009; 4: 53-58.
- Jadhav, Nikam SV, Bhamre SN, Jaid EL. Study of *Eimeria* necatrix in broiler chicken from Aurangabad District of Maharashtra state India. Int Multidis Res J 2011; 1:11-12. Shirley MW. The genome of *Eimeria* species, with special reference to *Eimeria tenella* coccidium from the chicken. Int J Parasitol 2000; 30: 485-493. https://doi.org/10.1016/S0020-7519(99)00183-6
- Zaman MA, Iqbal Z, Abbas RZ, Khan MN. Anticoccidial activity of herbal complex in broiler chickens challenged with *Eimeria tenella*. J Parasitol 2012; 139: 237-243. https://doi.org/10.1017/S003118201100182X
- 10. Abbas RZ, Iqbal Z, Khan MN, Zafar MA, Zia MA. Anticoccidial activity of *Curcuma longa* L in Broilers. Braz Arch Biol Technol 2010; 1: 63-67. https://doi.org/10.1590/S1516-89132010000100008
- 11. Nogueira VA, Franca TN, Peixoto PV. Ionophore poisoning in animals. Pesq Vet Brasil 2009; 29: 191-197.

- Chapman HD. Practical use of vaccines for the control of Coccidiosis in the chicken. World's Poult Sci J 2000; 56: 7-20. https://doi.org/10.1079/WPS20000002
- Tacconelli E. Antimicrobial use: Risk driver of multidrug resistant microorganisms in healthcare settings. Curr Opin Infect Dis 2009; 22: 352-358.
 - https://doi.org/10.1097/QCO.0b013e32832d52e0
- 14. Abbas RZ, Colwell DD, Gilleard J. Botanicals: an alternative approach for the control of avian coccidiosis. Int J Agric Biol 2012; 68:203-215.
 - https://doi.org/10.1017/S0043933912000268
- Nweze NE, Obiwulu IS. Anticoccidial effects of Ageratum conyzoides. J Ethno pharm 2009; 122: 6-9. https://doi.org/10.1016/j.jep.2008.11.014
- Lorrain B, Dangles O, Genot C, Dufour C. Chemical modeling of heme-induced lipid oxidation in gastric conditions and inhibition by dietary polyphenols. J Agric Food Chem 2010; 58: 676-683. https://doi.org/10.1021/if903054e
- 17. Kensa M, Syhed Y. Phytochemical screening and antibacterial activity on *Ricinus communis*. Plant Sci F 2011; 1:167-173.
- Verma SK, Yousaf S, Singh SK, Prasad GBKS, Dua VK. Antimicrobial potential of roots of *Ricinus communis* against pathogenic microorganisms. Int J Pharm Bio Sci 2011; 2: 545-548.
- Ryley JF, Meade R, Burst JH, Robinson TE. Methods in coccidiosis research: separation of oocysts from faeces. J Parasitol 1976; 73: 311-326. https://doi.org/10.1017/S0031182000046990
- Gorden HM, Whitlock HV. A new technique for counting nematode eggs in sheep faeces. J Count Sci Indust Res 1939; 12: 50-59.
- Molan AL, Liu Z, De S. Effect of pine bark (*Pinus radiata*) extracts on sporulation of coccidian oocysts. J Parasitol 2009; 56: 1-5. https://doi.org/10.14411/fp.2009.001
- 22. Akhtar M, Tariq AF, Awais MM, Iqbal Z, Muhammad F, Shahid M, Sawicka EH. Studies on wheat bran araybinoxylan for its immunostimulatory and protective effect avian coccidiosis. Sci Direct 2012; 90: 333-339. https://doi.org/10.1016/j.carbpol.2012.05.048
- Youn HJ, Noh JW. Screening of the anticoccidial effects of herb extracts against *Emieria tenella*. Vet Parasitol 2001; 96: 257-263. https://doi.org/10.1016/s0304-4017(01)00385-5
- Jang SI, Jun MH, Lillehoj HS, Dalloul RA, Kong IK, Kim S, Min W. Anticoccidial effect of green tea-based diets against *Eimeria maxima*. Vet Parasitol 2007; 144: 172-175. https://doi.org/10.1016/j.vetpar.2006.09.005
- 25. Naidoo V, McGaw LJ, Bisschop SP, Duncan N, Eloff JN. The value of plant extracts with antioxidant activity in attenuating coccidiosis in broiler chickens. Vet Parasitol 2008; 153: 214-219.

https://doi.org/10.1016/j.vetpar.2008.02.013

- 26. Nematollahi A, GH. Moghaddam, Niyazpour F. Prevalence of *Eimeria* species among Broiler chicks in Tabriz (Northwest of Iran). Res J Poult Sci 2008; 2: 72-74.
- 27. Gupta MK, Singh AK, Kumar S. Pharmacognostical investigation of *Ricinus communis* stem. Int J Pharm Sci Res 2011; 16: 91-97.
- Awais MM, Akhtar M, Muhammad F, Haq AU, Anwar MI. Immunotherapeutic effects of some sugar cane (*Saccharum officinarum* L.) extracts against coccidiosis in industrial broiler chickens. Exp Parasitol 2011; 128:104-110. https://doi.org/10.1016/j.exppara.2011.02.024
- 29. Khan JA, Yadav KP. Assessment of antibacterial properties of *Ricinus communis*. J Pharm Bio Sci 2011; 11:1-3.
- Lee HA, Hong S, Chung YH, Song KD, Kim O. Anticoccidial effects of *Galla rhois* extract on *Eimeria tenella*-infected chicken. Lab Anim Res 2012; 28:193-197. https://doi.org/10.5625/lar.2012.28.3.193

- 31. Awais MM, Akhtar M, Iqbal Z, Muhammad F, Anwar MI. Seasonal prevalence of coccidiosis in industrial broiler chickens in Faisalabad, Punjab, Pakistan. Trop Anim Health Prod 2012; 44:323-8. https://doi.org/10.1007/s11250-011-0024-x
- 32. Donkor AM, Mosobil R, Suurbaar J. In vitro bacteriostatic and bactericidal activities of Senna alata, *Ricinus communis* and *Lannea barteri* extracts against wound and skin disease causing bacteria. J Anal Pharm Res 2016; 3(1): 40-46. https://doi.org/10.15406/japlr.2016.03.00046
- 33. Kumar P, Joshi S, Sati SC, Rai D. A comparative evaluation of phytochemical and antibacterial properties of *Ricinus communis* L and *Thevetia peruviana* Schum of Kumaun Himalaya. Mintage J Pharm Med Sci 2016; 5:13-19.
- Sogan N, Kapoor N, Singh H, Kala S, Nayak A, Nagpal BN. Larvicidal activity of *Ricinus communis* extract against mosquitoes. J Vector Borne 2018; 55:282–290 https://doi.org/10.4103/0972-9062.256563
- 35. Suurbaar J, Mosobil R, Donkor AM. Antibacterial and antifungal activities and phytochemical profile of leaf extract from different extracts of *Ricinus communis* against selected pathogens 2017; 10:660-666. https://doi.org/10.1186/s13104-017-3001-2
- Ahmed F, Iqbal M. Antioxidant activity of *Ricinus communis*. Org Med Chem Int J 2018; 5:555-567.
- Rashmi, Pathak DV, Kumar. Effect of *Ricinus communis* L on microorganisms: advantages and disadvantages. Int J Curr Microbio App Sci 2019; 8(04): 878-884. https://doi.org/10.20546/ijcmas.2019.804.099
- Singh RK, Gupta MK, Singh AK, Kumar S. Pharmacognostical investigation of *Ricinus communis* stem. Int J Pharm Sci Res 2010; 16: 89-94. https://doi.org/10.13040/IJPSR.0975-8232.1 (6).89-94
- Abbas A, Iqbal Z, Abbas RZ, Khan MK. *In-vitro* anticoccidial potential of *Saccharum officinarum* extract against *Eimeria oocysts*. Bol Latinoam Caribe Plant Med Aromat 2015; 14 (6): 456 – 461.
- Momoh AO, Oladunmoye MK, Adebolu TT. Evaluation of the antimicrobial and phytochemical properties of oil from castor seeds (*Ricinus communis* Linn). Environment Pharm L Sci 2012; 10: 21-27.
- 41. Jombo TA, Enenebeaku NO. Antibacterial profile of fermented seed extracts of *Ricinus communis*: findings from a preliminary analysis. J Physiol Sci 2008; 23: 55-59. https://doi.org/10.4314/njps.v23i1-2.54926
- 42. Cornelia PV, Nastold P, Jetter R. Homologous very longchain 1, 3-alkanediols in leaf cuticular waxes of *Ricinus communis*. Phytochem 2010; 62(3): 433-438. https://doi.org/10.1016/S0031-9422(02)00560-5
- 43. Iqbal J, Zaib S, Farooq U, Khan A, Bibi I, Suleman S. Antioxidant, antimicrobial, and free radical scavenging potential of aerial parts of *Periploca aphylla* and *Ricinus communis*. Int Schol Res Net Pharm 2012; 12:563-567. https://doi.org/10.5402/2012/563267
- Rathod G, Pandhure N, More P. Phytochemical analysis and antibacterial activity in *Ricinus communis* L. Per Res 2014, 3:35-45.
- 45. Abbas A, Iqbal Z, Abbas RZ, Khan MK. *In vivo* anticoccidial effects of *Beta vulgaris* (sugar beet) in broiler chickens. Microb Patho 2017; 111:139-144. https://doi.org/10.1016/j.micpath.2017.07.052
- 46. Akhtar M, Haia A, Awais MM, Iqbal Z, Muhammad F, Haq AU, Anwar MI. Immunostimulatory and protective effects of Aloe Vera against coccidiosis in industrial broiler chickens. Vet Parasitol 2012; 186: 170–177. https://doi.org/10.1016/j.vetpar.2011.11.059
- Hussain F, Ahmed F. Antioxidant, antidiabetic and antibacterial activities of *Ricinus communis* root extracts. J. Fundam App Sci 2019; 11(2), 551-662. https://doi.org/10.4314/jfas.v11i2.7