

Available online at www.ujpronline.com Universal Journal of Pharmaceutical Research

An International Peer Reviewed Journal

ISSN: 2831-5235 (Print); 2456-8058 (Electronic) Copyright©2022; The Author(s): This is an open-access article distributed under the terms of

the CC BY-NC 4.0 which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited



RESEARCH ARTICLE

THE CYTOTOXICITY ACTIVITY OF ETHANOLIC EXTRACT OF ACANTHUS ILICIFOLIUS L LEAVES USING BRINE SHRIMP LETHALITY TEST (BSLT) METHOD

Dyah Ratna Ayu Puspita Sari^{*1}, Putu Yudha Ugrasena², Ni Wayan Ria Ekayani²

¹Associate's Degree of Pharmacy, Akademi Kesehatan Bintang Persada, Denpasar, Indonesia. ²Department of Pharmacy, Sekolah Tinggi Kesehatan Bali Wisnu Dharma, Indonesia.

Article Info:

Cite this article

7(1):56-59.

Article History:

Sari DRAP, Ugrasena PY, Ekayani NWR. The

cytotoxicity activity of ethanolic extract of

Acanthus ilicifolius L leaves using Brine Shrimp

Lethality Test (BSLT) method. Universal

Journal of Pharmaceutical Research 2022;

Dyah Ratna Ayu Puspita Sari, Associate's

Degree of Pharmacy, Akademi Kesehatan

https://doi.org/10.22270/ujpr.v7i1.724

Bintang Persada, Denpasar, Indonesia

*Address for Correspondence:

Received: 2 December 2021 Reviewed: 6 January 2022

Accepted: 9 February 2022

Published: 15 March 2022

Abstract

Aim and objective: *Acanthus ilicifolius* also known as jeruju is a plant that has a lot of bioactivity that can be used as a potential medicinal plant development. This study aimed to determine the cytotoxicity activity of ethanol extract of *A. ilicifolius* leaves.

Methods: This study used Brine Shrimp Lethality Test (BSLT) method using *Artemia salina* larvae with test solution series concentration of 10, 50, 100, 500, 1000 ppm and control without extract. *A. salina* was added to each test tube. After 24 hours, the larvae mortality was observed. LC_{50} assessment was analyzedwith probit analysisby Microsoft excel.

Result: The largest mortality percentage was shown at a concentration of 1000 ppm ethanol extract of *A. ilicifolius* leaves with an average mortality value of 80%, while the concentrations of 10 ppm, 50 ppm, 100 ppm, and 500 ppm had an average mortality value of 20%, 40%, 57% and 63%. The ethanol extract of *A. ilicifolius* leaves categorized as toxic with a LC₅₀ value of 103,6 ppm.

Conclusion: It was concluded that ethanol extract of *A. ilicifolius* leaves has cytotoxicity activity potential.

Keywords: Acanthus ilicifolius, Artemia salina, BSLT, cytotoxicity.

E-mail: dyrasari2@gmail.com

INTRODUCTION

Tel-+6285340632711;

Nowadays, the use of medicinal plants is very popular almost all over the world. They have been used traditionally as medicine for thousands of years in countries such as China, India, Thailand, Japan and Indonesia. Indonesia is a country that has abundant biodiversity. Indonesia has tropical forests with 3000 species of plants and 1,845 of them are medicinal plants¹. Medicinal plants are well-known as the basic materials of herbal medicine and traditional medicine. Medicinal plants are plants that contain active substances in part or all of plant parts that can be used to treat or prevent disease². Some medicinal plants that are widely used are Artocarpus altilis, Centella asiatica, Piper betle Hibiscus rosasinensis, Blumea balsamifera, Alium sativum, Curcuma longa, Mimosa pudica, Carica papaya, and Acanthus ilicifolius. The various kinds of pharmacological effects of these plants are as antioxidants, antibacterial, antihypertension, antidiabetic, anti-inflammatory, analgesic and many other effects^{3,4,5}. This pharmacological activity is due to the presence of chemical constituents contained in the plants such as secondary metabolites. Secondary metabolites are various chemical compounds produced by plant cells with various biological effects⁶. Various types of secondary metabolites in plants play a role in providing pharmacological activity.

Acanthus ilicifolius also known as Jerujuis one of the plants that is used as a medicinal plant. A. ilicifolius is a member of the Acanthaceae family and is amangrove vegetation plants grow in tropical and subtropical intertidal habitats7. A. ilicifolius has bioactive compounds such as alkaloids, flavonoids, tannins, saponins, terpenoid, and steroids⁸. Andriani et al., reported that methanolic extract of A. ilicifolius leaves contained alkaloids, flavonoids, polyphenols, tannins, steroids, and glycosides⁹. Phenylethanoid glycosides mostly found in A. *ilicifolius* leaf ethanolic extract¹⁰. The main type of phenylethanoid glycosides containedin A. ilicifolius are isoacteoside and acteoside¹¹. Previous studies showed that A. ilicifolius leaves extract has antifungal, antioxidant, analgesic, antimicrobial, and hepatoprotective activities^{11,12,13}. A.

ilicifolius leaves infusion at a concentration of 40% has potential effect as analgesic in mice¹³. Other studies have reported that methanolic extract of *A. ilicifolius* leaves have strong antioxidant (IC₅₀: 17.51 µg/ml) and antifungal activity against *Candida albicans*⁹. Zhang *et al.*, reported that phenylethanoid glycosides content of *A. ilicifolius* has potential as hepotoprotective in liver injury induced by carbon tetrachloride (CCl₄) *in vivo* and *in vitro*¹¹.

The pharmacological effects of plants are due to the presence of secondary metabolites of the plants. The pharmacology activity of these active components as herbal medicines can be determined through a preliminary analysis in the form of a cytotoxicity analysis. The method that is often used in the analysis of cytotoxicity is the Brine Shrimp Lethality Test (BSLT). This assay can describes the toxicity effect of the extract against *Artemia salina* larvae. This test can be used as an initial step to identify more plant bioactivity. Therefore, this present study was conducted to determine the cytotoxic activity of ethanol extract of *A. ilicifolius* leaves against *A. salina*.

MATERIALS AND METHODS

Solvents

96% ethanol (Brataco®), aquadest (Brataco®), Tween 80 (Sigma aldrich®), H₂SO₄(Merck®), FeCl₃ (Brataco ®), HCL (Merck®), chloroform (Brataco®), ammonia (Brataco®)

Sample Preparation

Fresh leaves of *A. ilicifolius* were collected and picked in November 2021 from Gitgit Village, Sukasada District, Buleleng Regency, and Bali and were botanically identified by Balai Konservasi Tumbuhan Kebun Raya "Eka Karya" Bali (LIPI).

Extraction

The fresh leaves of *A. ilicifolius* were washed with flowing water and dried by aeration without exposure to direct sunlight. The dried leaves were powdered by blender. Leaf powder was extracted using 96% ethanol for five days with stirring. The filtrate was collected and evaporated using rotary evaporator and obtained 9.3 grams of the crude extract.

Phytochemical Screening

Test for alkaloids

Total 0.1 gm of extract was added with 10 ml of chloroform and a few drops of ammonia. The chloroform fraction was separated and acidified with the addition of a few drops of concentrated sulfuric acid. The acid fraction was divided into 2 tubes, and each was added with Meyer and Dragendroff reagent. The formation of a white precipitate on Meyer's reagent and a red precipitate on Dragendroff's indicates the presence of alkaloids¹⁴.

Test for flavonoids

Total 2 ml of extract was heated and then added with ethanol. The solution is added with magnesium powder and HCl. The presence of flavonoids is indicated by the formation of a red or orange color¹⁴.

Test for phenolic compound

Total 0.1g of the extract was dissolved in methanol and then 2-3 drops of 5 % FeCl₃ were added. The presence

of phenolic compounds in the extract was indicated by the formation of a dark green $color^{15}$.

Test for saponins

Total 1g extracts added 5 mL of aquadest, shaken vertically for 10 seconds. Formation of foam 1-10 cm high which is stable for not less than 10 minutes indicates the presence of saponins¹⁶.

Test for steroids

Total 0.5g of extract was added 2-3 drops of anhydrous acetic acid then added 3 drops of sulfuric acid, left for a few minutes. The color changes that occur are observed. A blue or green discoloration indicates the presence of steroids¹⁷.

Test for tannins

Total 0.1g of *A. ilicifolius* leaves extract was dissolved in methanol, and then 2-3 drops of 1% FeCl₃ solution were added. The formation of a dark green or bluish black color indicates the presence of tannins¹⁵.

The cytotoxicity assay

The cytotoxicity assay was carried out with the Brine Shrimp Lethality Test (BSLT) by using *A. salina*. Preparation of *A. salina* larvae is carried out by incubating the eggs for 48 hours in an aquarium filled with seawater. The test solution of *A. ilicifolius* leaves ethanol extract was made in a series of concentrations of 10, 50, 100, 500, 1000 ppm. After the solvent evaporated, 50 L of tween, 1 ml of seawater, and added 10 ml A. *salina* larvae into each tube. Then the sea water is added again up to 5 ml. Normal control was also made without the addition of extract. After 24 hours, the larvae mortality was observed¹⁸⁻²⁰.

%Mortality =
$$\frac{\text{Total Larvae Mortality}}{\text{Total Larvae}} x 100\%$$

 LC_{50} assessment was analyzed with Probit analysis by Microsoft excel. The determination of the toxicity category is determined based on the LC_{50} value divided into three categories, that is non-toxic with an LC_{50} value of > 1000 ppm, toxic with an LC_{50} value of 30-1000 and ppm very toxic with an LC_{50} value of <30 ppm²⁰.

RESULTS AND DISCUSSION

Phytochemical screening

The phytochemical group test was performed and the result indicated that alkaloids, flavonoids, phenols, saponins, steroids, and tannins were detected in the ethanol extract of *A. ilicifolius* leaves.

Cytotoxicity activity

Cytotoxic activity assay was carried out by Brine Shrimp Lethality Test (BSLT). This method is a simple method that is often used for cytotoxic assay. Based on the Table 1, it showed that in the control group there was no larvae mortality. The largest mortality percentage was shown at a concentration of 1000 ppm ethanol extract of *A. ilicifolius* leaves with an average mortality value of 80%, while the concentrations of 10 ppm, 50 ppm, 100 ppm, and 500 ppm had an average mortality value of 20%, 40%, 57% and 63%. It shows that larvae mortality is not affected by seawater but by *A. ilicifolius* leaf extract. The higher the concentration of the extract, the higher the mortality of *Artemia salina* larvae. Based on the result, it showed that the ethanol extract of A. ilicifolius leaves categorized as toxic with a LC₅₀ value of 103.66 ppm. So that it has cytotoxicity potential, this is related to secondary metabolite compounds such as phenolics, flavonoids and tannins, which are contained in the extract which at certain levels has the potential for cytotoxicity and caused larvae mortality.

Concentration	Log 10	Replication	Total larvae	Mortality	% Mortality	Probit	LC 50
(ppm)	10	1		0	Į.	0	(ppm)
a . 1		1	10	0	0%	0	- 0
Control	-	2	10	0	0%	6 0	0
		3	10	0	0 %	0	
10		1	10	2			
	5.00	2	10	2	20%	4.16	
		3	10	2			
50		1	10	3			-
	5.70	2	10	5	40%	4.75	
		3	10	4			
100		1	10	6			-
	6.00	2	10	4	57%	5.18	103,6
		3	10	7			
500		1	10	7			-
	6.70	2	10	4	63%	5.33	

10

10

10

3

1

2

	3	10	10	
•	is related to flavonoids		CONFLIC	T OF INTERE

The mechanism of larvae mortality function of phenolic compounds tannins in A. ilicifolius leaves which can inhibit larval feeding power (antifedant). The way these compounds work is by acting as stomach poisoning. Therefore, when these compounds enter the larva's body, the digestive system will be disturbed. This causes the larvae to fail to get a taste stimulus so they are unable to recognize the food so that the larvae starve to death²¹.

7.00

CONCLUSION

Ethanol extract of A. ilicifolius leaves is toxic based on Brine Shrimp Lethality Test and has cytotoxicity activity potential with a LC_{50} value of 103.6 ppm.

ACKNOWLEDGEMENT

1000

The authors Acknowledge Akademi Kesehatan Bintang Persada for supporting this study.

AUTHOR'S CONTRIBUTION

Sari DRAP: writing original draft, methodology. Ugrasena PY: research design, data collection. Ekayani NWR: statistical analysis, conceptualization. Final manuscript was read and approved by all authors.

DATA AVAILABILITY

The data supporting the findings of this study are not currently available in a public repository but can be made available upon request to the corresponding author.

EST

80%

None to declare.

8

8

6

REFERENCES

1. Zuraida Z. Toxicity analysis of several forest plants using Brine Shrimp Lethality Test (BSLT) Method. J Penelit Has Hutan 2018; 36(3):239-46. https://doi.org/10.20886/jphh.2018.36.3.239-246

5.84

- Siregar RS, Tanjung AF, Siregar AF, Bangun IH, Mulya MO. Literature study of utilization of traditional medicinal plants. Semin Soc Sci Eng Hum 2020; 4049:385-91. https://doi.org/10.1186%2Fs13002-018-0212-0
- 3. Sari DRAP, Ahmad FF, Djabir YY, Yulianty R. Breadfruit leaves extract (Artocarpus altilis) effect on pancreatic damage in diabetic type II animal model induced by alloxannicotinamide. Med Clin Pract 2020; 3:100099. https://doi.org/10.1016/j.mcpsp.2020.100099
- 4. Yunita E, Sari DRAP. Antibacterial potential of Centella asiatica against gram positive and gram negative bacteria. J Edukasi Mat dan Sains 2020; IX(2):236-40. https://doi.org/10.5281/zenodo.4305192
- 5. Bamola N, Verma P, Negi C. A Review on some traditional medicinal plants. Int J Life-Sciences Sci Res 2018; 4(1):1550-6. https://doi.org/10.21276/ijlssr.2018.4.1.7
- 6. Hussein RA, El-Anssary A. Plants secondary metabolites: the key drivers of the pharmacological actions of medicinal plants. Herb Med 2019; 11-30. http://dx.doi.org/10.5772/intechopen.76139
- 7. Singh D, Aeri V. Phytochemical and pharmacological potential of acanthus ilicifolius. J Pharm Bioallied Sci 2013 Jan; 5(1):17-20. https://dx.doi.org/10.4103%2F0975-7406.106557
- 8. Ernianingsih WS, Mukarlina, Rizalinda. Ethnopharmacology of mangrove plants Achantus ilicifolius L, Acrostichum speciosum L. and Xylocarpus rumphii Mabb. In Sungai Tekong village, Sungai Kakap district, Kubu Raya Regency. J Protobiont 2014; 3 (2)(2):252-8. http://dx.doi.org/10.26418/protobiont.v3i2.6833

9. Andriani D, Revianti S, Prananingrum W. identification of compounds isolated from amethanolic extract of Acanthus

Ilicifolius leaves and evaluation of their antifungal and antioxidant activity. Biodiversitas 2020; 21(6):2521-5. https://doi.org/10.13057/biodiv/d210625

10. Zhang M, Ren X, Yue S, Zhao Q, Shao C, Wang C. Simultaneous quantification of four phenylethanoid glycosides in rat plasma by UPLC-MS/MS and Its application to a pharmacokinetic study of *Acanthus ilicifolius* herb. Molecules 2019 Aug; 24(17).

https://doi.org/10.3390/molecules24173117

- 11. Zhang M-Q, Ren X, Zhao Q, Yue S-J, Fu X-M, Li X, et al. Hepatoprotective effects of total phenylethanoid glycosides from Acanthus ilicifolius L. against carbon tetrachlorideinduced hepatotoxicity. J Ethnopharmacol 2020 Jun; 256: 112795. https://doi.org/10.1016/j.jep.2020.112795
- 12. Avijit D, Md RS, Md SIH, Md H, ASM. M–Al-H. Phytochemical screening and the evaluation of the antioxidant, cytotoxic and antimicrobial properties of *Acanthus ilicifolius* (Family: Acanthaceae). Int Res J Pharm. 2012; 3(8):153–6.https://doi.org/10.1186/s40816-019-0100-8
- Lara AD, K FS, Lara AD. Analgesic activity test of *Acanthus ilicifolius* L. leaf infusion in male white mice (*Mus musculus*). Indonesian J Pharm Sci 2021; 3(2):71-80.
- Harborne JB. Phytochemical methods: A guide to the modern way of analyzing plants. Bandung: ITB Publisher; 1987.

- 15. Robinson T. Kandungan Organik Tumbuhan Tingkat Tinggi. Bandung: Penerbit ITB; 1991
- Ministry of Health RI. Indonesian Pharmacopoeia. Edition V. Jakarta: Ministry of Health of the Republic of Indonesia; 1995,
- Fauziah A, Sudirga SK, Parwanayoni NMS. Antioxidant test leunca plant leaf extract *Solanum nigrum* L. J Metamorfosa 2021;8(1):23-24.

https://doi.org/10.24843/metamorfosa.2021.v08.i01.p03

- Colegate SM, Molyneux RJ. (Eds.). Bioactive natural products: detection, isolation, and structural determination. CRC Press; 2007
- 19. Hamidi MR, Jovanova B, Panovska TK. Toxicological evaluation of the plant products using Brine Shrimp (*Artemia* salina L.) model. Maced Pharm Bull 2014; 60(1): 9-18. https://doi.org/10.33320/MACED.PHARM.BULL.2014.60.01.00 2
- Meyer BN, Ferrigni NR, Putnam JE, Jacobsen LB, Nichols DE, McLaughlin JL. Brine Shrimp: A convenient general bioassay for active plant constituents. Planta Med 1982 May; 45(5):31-4. https://doi.org/10.1055/s-2007-971236
- Muaja AD, Koleangan HSJ, Runtuwene MRJ. Toxicity test with BSLT method and analysis of phytochemical content of *Saurauia bracteosa* leaf extract with soxhletation method. J MIPA 2013; 2(2):115.

http://dx.doi.org/10.35799/jm.2.2.2013.3000