

 **Available online at** *[www.ujpronline.com](http://www.ujpronline.com/)*  **Universal Journal of Pharmaceutical Research**  *An International Peer Reviewed Journal*

 **ISSN: 2831-5235 (Print); 2456-8058 (Electronic)**

 **Copyright©2018; The Author(s): This is an open-access article distributed under the terms of the CC BY-NC 4.0 which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited**



#### **RESEARCH ARTICLE**

# **MYOCARDIAL POTENCY OF AN AQUEOUS EXTRACT OF** *HARUNGANA MADAGASCARIENSIS* **STEM BARK AGAINST ISOPROTERENOL-INDUCED MYOCARDIAL DAMAGE IN RATS**

**Esther Ngo Lemba Tom1\* [,](https://orcid.org/0000-0003-2838-1848) Florette Diane Nankia<sup>1</sup> [,](https://orcid.org/0000-0003-3064-6235) Nyemb Nyunaï<sup>2</sup> [,](https://orcid.org/0000-0002-1436-8178) Corine Girard Thernier<sup>3</sup> [,](https://orcid.org/0000-0002-5116-1395) Céline Demougeot<sup>3</sup> [,](https://orcid.org/0000-0003-0639-9756) Théophile Dimo[4](https://orcid.org/0000-0002-8565-1181)**

*Department of Biological Sciences, Higher Teachers' Training College, University of Yaoundé I, Yaoundé, Cameroon. Medical Research Centre, Institute of Medical Research and Medicinal Plants Studies (IMPM), Yaoundé, Cameroon. EA 4267 PEPITE, UFR Sciences Médicales et Pharmaceutiques, Université de Bourgogne Franche-Comté, 19 rue Ambroise Paré, bâtiment S, 25030, Besancon cedex, France.*

*<sup>4</sup>Department of Animal Biology and Physiology, Faculty of Sciences, University of Yaoundé I, P.O Box 812, Yaoundé, Cameroon.*

# **Article Info:**



#### **Article History:** Received: 9 December 2017 Reviewed: 13 January 2018

\_

Accepted: 17 February 2018 Published: 15 March 2018

#### **Cite this article:**

Tom ENL, Nankia FD, Nyunaï N, Girard-Thernier C, Demougeot C, Dimo T. Myocardial potency of aqueous extract of *Harungana madagascariensis* stem bark against isoproterenol-induced myocardial damage in rats. Universal Journal of Pharmaceutical Research 2018; 3(1): 17-24. *<http://doi.org/10.22270/ujpr.v3i1.R4>*

\_

\_

#### **\*Address for Correspondence:**

**Dr. Esther Ngo Lemba Tom,** Department of Biological Sciences, Higher Teachers' Training College, University of Yaoundé I, P.O Box 47, Yaoundé, Cameroon. Mobile: +237-699828838, E-mail: *[esther\\_ngotom@yahoo.com](mailto:esther_ngotom@yahoo.com)*

#### **Abstract**

**Objectives:** The present study was undertaken to evaluate the effects of *Harungana madagascariensis* on electrocardiographical, biochemical and histopathological changes in isoproterenol (ISO)-induced myocardial infarction in rats.

\_

**Methods:** Male *Wistar* albino rats were randomly divided and treated with the aqueous extract of *Harungana madagascariensis* stem bark (AEHM, 200 and 400 mg/kg per os), or normal saline or vitamin E for 7 days with concomitant administration of ISO (85 mg/kg, subcutaneously) on  $8<sup>th</sup>$  and  $9<sup>th</sup>$  days, at 24 h interval.

**Results:** The ISO injections to the rats caused cardiac dysfunction evidenced by a marked  $(p<0.01)$  elevation in ST-segment, a reduction in R wave amplitude (*p<*0.01), decrease in endogenous antioxidant reduced glutathione (GSH), increase in malondialdehyde (MDA), a lipid peroxidation marker, increase of cardiac marker enzymes lactate dehydrogenase (LDH), aspartate amino transferase (AST) and alanine amino transferase (ALT). All these changes in cardiac function as well as GSH, MDA and the enzymes (LDH, AST and ALT) were ameliorated when the rats were pretreated with AEHM. Additionally, the protective effects were strengthened by improved histopathological changes, which specify the protection of cardiomyocytes from the deleterious effects of ISO.

**Conclusion:** This study demonstrates the cardioprotective effect of *Harungana madagascariensis* on isoproterenol-induced myocardial infarction in rats. The mechanism might be associated with the enhancement of antioxidant defense, reduction of lipid peroxydation and it is confirmed by amending electrocardiographic pattern, improvement of cardiac markers and less histopathological damages following ISO-induced myocardial infarction. It could provide experimental evidence to support the use of *Harungana madagascariensis*  used in traditional medicine to treat cardiovascular disorders.

**Keywords:** Antioxidants, electrocardiography, *Harungana madagascariensis*, isoproterenol, myocardial infarction.

## **INTRODUCTION**

According to the recent world health organization survey, an estimated 17.7 million people died from to cardiovascular disease (CVDs) in 2015, representing 31% of all global deaths. Of these deaths, an estimated 80% were due to myocardial infarctions (MI) and strokes<sup>1</sup>[.](#page-6-0) According to the same survey, over three quarters of CVDs deaths occurred in low- and middleincome countries. MI is followed by several

biochemical alterations, such as lipid peroxidation, free radical damage, hyperglycemia, hyperlipidemia, elevation in cardiac markers and pro-inflammatory cytokines leading to qualitative and quantitative alterations of myocardiu[m](#page-6-1)**<sup>2</sup>** . Catecholamines at low concentrations are beneficial in regulating heart functions by exerting a positive inotropic action on the myocardiu[m](#page-6-2)**<sup>3</sup>** , whereas high concentrations of catecholamines or chronic exposure to catecholamines over a prolonged period produces deleterious effects on

the cardiovascular system**<sup>4</sup>** [.](#page-6-3) Isoproterenol (ISO) is a synthetic catecholamine, a non-selective βadrenoreceptor agonist, which causes severe stress in the myocardium and produces infarct like lesions, when injected in rat[s](#page-6-4)<sup>5</sup>. The ISO model is a well standardized and most reliable model for assessing the cardioprotective activity of several drugs. Since its pathophysiological and morphological changes following ISO administration are comparable to those taking place in human M[I](#page-6-5)**<sup>6</sup>** . Nowadays, a number of pharmacological interventions such as beta-blockers, angiotensine-converting enzyme inhibitors, antiplatelet agents, thrombolytics, calcium antagonists, nitrates, antioxidants have been shown to counteract the ill effect of myocardial ischemic injury and to reduce morbidity and mortality in patients with ischemic heart disease**[7,](#page-6-6)[8](#page-6-7)** . However, their chronic usage is often associated with adverse effects**<sup>9</sup>** [.](#page-6-8) Therefore, the development of new and safer drugs for the treatment and prevention of ischemic heart disease is still a major concern. There is increasing trend towards the application of herbal medicines to treat the cardiovascular diseases**[10,](#page-6-9)[11](#page-6-10)** . *Harungana madagascariensis* is one of the most popular trees in the African traditional medicine system. It is used as an abortifacient and antiseptic, in the treatment of cardiovascular disorders, anemia, tuberculosis, fever, angina, diarrhea, dysentery, syphilis, gonorrhea, malaria, parasitic skin diseases and wounds, as a natural source of dermatological agents and cosmetics**[12-](#page-6-11)[16](#page-7-0)**. Its benefits have also been reported in liver diseases, diabetes, pancreatic and biliary problems**[17,](#page-7-1)[18](#page-7-2)**. Biological studies on the barks or leaves of this plant revealed antihelminthiase properties**[19](#page-7-3)** , anti-plasmodial<sup>[20](#page-7-4)</sup>, antidiabetic<sup>[21](#page-7-5)</sup>, antimicrobial activities<sup>22</sup>, analgesic and anti-inflammatory  $\arctivities^{22}$  $\arctivities^{22}$  $\arctivities^{22}$ , analgesic and anti-inflammatory activities**[23](#page-7-7)**. Some of the constituents and isolated compounds from *H. madagascariensis* includes flavonoids, alkaloids, saponins, terpenes, cardiac glycosides, and tannins**[24](#page-7-8)**. A prenylated 1, 4 anthraquinone isolated from the hexane extract of the stem-bark of *H. madagascariensis* possess alphaglucosidase inhibition and antioxidant activities**[25](#page-7-9)**. In this context, an attempt has been made to investigate the effect of an aqueous extract of *H. madagascariensis* on maintaining the myocardial integrity in animals employing electrocardiographical, biochemical and histopathological parameters in ISO-induced myocardial infarction.

## **MATERIALS AND METHODS**

## **Plant material collection and extraction**

Fresh *H. madagascariensis* stem barks were collected at Essezok, Mbalmayo (Center Region, Cameroon) in June 2016. The identification of the plant was done at the Cameroon National Herbarium where voucher sample were deposited under the registration number NO. 4224 HNC. Bark pieces were dried under room temperature and powdered with the help of electrical grinder. Total 500 g of powder was introduced into 3.5 L of distilled water and boiled for 20 minutes. The resulting decoction was filtered through Whatman paper No. 3 and further lyophilized. A crude brown extract powder (HM extract, 31.73 g) was obtained, giving a yield of 6.35%.

## **Experimental animals**

Male albino *Wistar* rats (150-200 g) were obtained from the Animal House of the Faculty of Science at the University of Yaoundé I (Cameroon). They were kept at standard laboratory conditions under natural light and dark cycles, at constant room temperature  $(20 \pm 5^{\circ}C)$  and were allowed to have standard food and tap water freely. This study was approved by the Cameroonian National Ethical Committee (Ref NO. FW-IRB00001954).

# **Drugs and chemicals**

Isoproterenol hydrochloride was purchased from Sigma Aldrich, USA. Lactate deshydrogenase (LDH) kit for enzyme estimation was purchased from Hospitex Diagnostics. Aspartate amino transferase (AST) and alanine amino transferase (ALT) kits were from Fortress Diagnostics Biosystems. All chemicals used in the present study were of analytical grade.

# **Induction of experimental myocardial infarction**

Isoproterenol was freshly dissolved in 0.9% saline and injected (85 mg/kg) subcutaneously to the rats for two successive days (on days  $8<sup>th</sup>$  and  $9<sup>th</sup>$  respectively) at an interval of 24 h. Animals were sacrificed 48h after the first injection of isoproterenol.

# **Experimental design**

The animals were randomly divided into 7 groups consisting of 7 rats each. HM extract was dissolved in distilled water. Vitamin E was used as standard drug. Rats in group 1 (normal control) received distilled water (10 ml/kg) orally, for 9 days. Rats in group 2 (ISO control) received distilled water for 9 days and were injected isoproterenol (85 mg/kg, SC) on the 8th and 9th days at an interval of 24 hour. Animals of groups 3 to 5 were pretreated with the aqueous extract of HM (200 and 400 mg/kg/day) or vitamin E (100 mg/kg/day) orally for 9 days and on the 8th and 9th days they received isoproterenol SC at an interval of 24 hour. Rats in groups 6 and 7 were treated with the aqueous extract of HM (400 mg/kg/day) and vitamin E (100 mg/kg/day) orally for 9 days and on the 8th and 9 th days they were injected saline (0.1 ml/100g SC) at an interval of 24 h. Changes in body weight in all groups were noted every 2 days during the experimental period.

## **Electrocardiogram measurement**

Twenty four hours (24 h) after the last administration of the drugs, the animals were anesthetized by intraperitoneal injection of urethane (15 mg/kg). Needle electrodes were inserted under the skin of the animals in lead II position. Electrocardiograh recordings were made using Biopac Student Lab Experiment system (BSL 3.7, USA).

## **Blood collection and assessment of cardiac hypertrophy**

After recording the ECG, blood was collected from the abdominal aorta and allowed to clot for 1 h at room temperature. It serum was subsequently separated by centrifugation at 3000 rpm for 15 min at 4°C and stored at -20°C for biochemical assays. After the blood collection, the animals were euthanized. Their hearts were removed, rinsed in ice-cold normal saline and weighed. The wet heart weight to body weight ratio was calculated to assess the degree of myocardial weight gain.

#### **Assay of cardiac marker enzymes**

Activities of lactate dehydrogenase (LDH), aspartate aminotransferase (AST) and alanine aminotransferase (ALT) in the serum were measured using commercial kits (from Hospitex Diagnostics for LDH and Fortress Diagnostics for AST and ALT).

## **Estimation of lipid peroxidation product and reduced gluthathione in myocardium**

After weighing, the heart tissue was divided into two longitudinal parts. One part was homogenized in Mc Even physiological ice-cold solution (pH 7.4, 1:5 w/v). The homogenate was centrifuged at 3000 rpm for 30 min at 4°C and the supernatant was stored at -20°C for biochemical assays. Malondialdehyde (MDA), a thiobarbiturate reactive substance, was measured as a marker for oxidative stress in myocardial homogenates using trichloroacetic acid (TCA, 20%) and thiobarbituric acid (TBA, 0.67%)**[26](#page-7-10)**. The level of reduced glutathione (GSH) was estimated as previously described**[27](#page-7-11)** .

#### **Histopathological examination**

After weighing, the second part of the heart was fixed in 10% buffered formalin. The fixed tissues were embedded in paraffin, sectioned at 5 µm and stained with hematoxylin and eosin (Hand E). The sections were examined under a light microscope (Scientico STM-50) and photomicrographs were taken by a photomicroscope (Minisee 1.0) at x200 magnification. **Statistical analysis**

Results are shown as mean  $\pm$  SEM. The statistical comparisons among the groups were performed with tstudents test using Sigma Stat 3.5 statistical package. Mann Whitney post-test was employed to compare the mean values between the treated groups and the control. *p*-values less than 0.05 were considered as statistically significant.

## **RESULTS**

#### **Effect of** *H. madagascariensis* **on electrocardiogram**

The Lead II electrocardiograms obtained from the animals are shown in Figure 1. The rats receiving distilled water (control group) showed normal patterns of ECG, while those treated with isoproterenol alone (ISO group) demonstrated significant changes in ECG pattern. The changes included a marked elevation of ST segment from  $0.11 \pm 0.01$  mv in control group to  $0.18 \pm 0.01$  my in ISO-treated group ( $p < 0.01$ ) and a reduction in R wave amplitude from  $0.63 \pm 0.03$  mv in control group to  $0.3 \pm 0.02$  mv in ISO-treated group (*p<*0.01) which both are indicative of myocardial infarction (Figure 2).

**Table 1: Effect of** *H. madagascariensis* **aqueous extract on heart weight, body weight and heart weight/body weight ratio in rats.**

weight ratio in rats.			
Groups	Body weight (g) at the end of	Heart weight $(g)$	Heart weight/body
	the experimental period		weight ratio $(\% )$
Control	$193.57 \pm 8.69$	$0.61 \pm 0.03$	$0.31 \pm 0.01$
<b>ISO</b>	$185.01 + 7.88$	$0.74 \pm 0.03^*$	$0.40 \pm 0.01***$
<b>HM 200+ISO</b>	$178.71 + 4.20$	$0.71 \pm 0.02^*$	$0.39 \pm 0.01***$
$HM 400+ISO$	$179.57 + 2.42$	$0.73 \pm 0.02^*$	$0.40 \pm 0.01***$
$V$ it E+ISO	$185.00 \pm 8.82$	$0.75 \pm 0.03^*$	$0.40 \pm 0.01***$
<b>HM 400</b>	$190.29 \pm 6.75$	$0.56 \pm 0.02$ ###	$0.30 \pm 0.01$ ###
Vit E	192.86+4.47	$0.60+0.04$ <sup>#</sup>	$0.31 + 0.01$ ###

Values are given as mean $\pm$ SEM (n = 7).  $\dot{p}$  <0.05 and \*\* $p$  <0.001 as compared to the control group;  $\dot{p}$  <0.05 and  $\dot{p}$  =  $\dot{q}$  >  $\dot{q}$  =  $\dot{q}$ isoproterenol (ISO)-treated group. HM 200 or HM 400: *H. madagascariensis* aqueous extract (200 or 400 mg/kg); Vit E: vitamin E (100 mg/kg).





Values are given as mean±SEM (n = 7).  $\frac{4}{7}$  p<0.05 and  $\frac{4}{7}$  p<0.01 as compared to the control group;  $\frac{4}{7}$  p<0.05 and  $\frac{4}{7}$  p<0.01 as compared to the isoproterenol (ISO)-treated group. HM 200 or HM 400: *H. madagascariensis* aqueous extract (200 or 400 mg/kg) ; Vit E: vitamin E (100 mg/kg)

Oral treatment with *H. madagascariensis* at a dose of 200 mg/kg or vitamin E (100 mg/kg) resulted in a significant increase in the R wave amplitude from  $0.37\pm0.02$  mV in ISO-treated group to  $0.52\pm0.01$  and  $0.51\pm0.02$  mV in extract-treated group and vitamin E, respectively (*p<*0.01), Figure 2A). However, treatment with all doses of *H. madagascariensis* resulted in a non-significant decrease in the ST-elevation as compared to the rats treated with isoproterenol alone (Figure 2B).

## **Effects of** *H. madagascariensis* **on the heart weight to body weight ratio and body weight**

The mean body weight of the rats at the end of the experiment in all experimental groups had no significant change (Table 1). The heart weight and the ratio of heart weight to the body weight were increased significantly (*p<*0.05 and *p<*0.001 respectively) in ISO-administered groups when compared with control group. The extract of *H. madagascariensis* when given alone, significantly reduce the heart weight and the

ratio of heart weight to body weight (*p<*0.001) as compared to the ISO-treated group.

### **Effect of** *H. madagascariensis* **on serum marker enzymes**

As shown in Table 2, there was a significant rise observed in the levels of diagnostic marker enzymes (LDH (*p<*0.01), AST (*p<*0.05) and ALT (*p<*0.05)) in the serum of the ISO-treated rats. Pre-treatment with *H. madagascariensis* (200 and 400 mg/kg) as well as vitamin E (100 mg/kg) showed a significant reduction in the levels of all serum diagnostic marker enzymes compared to ISO group.

# **Effects of** *H. madagascariensis* **on lipid peroxidation and reduced glutathione level**

To determine the lipid peroxidation, MDA levels were measured in myocardial homogenates. Heart MDA levels increased insignificantly in isoproterenol alone treated rats as compared to the control group (Table 3). Pre-treatment with the *H. madagascariensis* (200 and 400 mg/kg) extract induced a dose-dependent but nonsignificant decrease of MDA levels of myocardium. There was a significant  $(p<0.001)$  decrease in GSH level in the heart of ISO-treated rats as compared to the control group. Pre-treatment with *H. madagascariensis* (200 and 400 mg/kg) significantly increased (*p<*0.001) the myocardial GSH level.



**Figure 1: Representative electrocardiogram tracings of control and experimental animals receiving isoproterenol (ISO),** *H. madagascariensis* **aqueous extract (200 or 400 mg/kg) + isoproterenol (HM 200 + ISO or HM 400 + ISO), vitamin E + isoproterenol (Vit E + ISO), the extract alone at 400 mg/kg (HM 400) and vitamin E alone (Vit E).**

The arrow indicates the decrease of the R wave amplitude. The ECG was recorded from II limb leads with recorder speed 0.5 s/div.



**Figure 2: Effects of oral administration of** *H. madagascariensis* **on R-amplitude (A) and ST segment (B) (recorded from limb lead II).**

Data are reported as mean±SEM (n = 7). \*  $p$  <0.05 and \*\*  $p$  <0.01 as compared to the control group;  $^{#}p$  <0.01 as compared to the isoproterenol (ISO)-treated group. HM 200 or HM 400: *H. madagascariensis* aqueous extract (200 or 400 mg/kg); Vit E: vitamin E (100 mg/kg).



**HM 400 + ISO**

**Figure 3: Effect of** *H. madagascariensis* **stem bark aqueous extract (HM) on histopathological changes in heart tissue.** 

**A:** Normal group received saline showing normal structure of myocardium;

**B:** Diseased group received two subcutaneous injections of isoproterenol (ISO, 85 mg/kg) showing necrosis of myofibrils and

edema through penetration of inflammatory cells;

**C, D**: HM 100 mg/kg and 200 mg/kg treated group showing lesser myocardial necrosis and edema following ISO administration. Heart tissues were stained with hematoxylin and eosin and visualized under light microscope at x200 magnification.

### **Histopathological examination of the cardiac tissue**

In the control group, myocardial fibers were arranged regularly with clear striation, without any damage (Figure 3A). Histopathological sections of the isoproterenol alone treated hearts displayed hypertrophy, degeneration of myocytes, infiltration of neutrophilic granulocytes and increased edematous

inter-muscular space and myofibroblasts (Figure 3B), whereas, the rats treated with *H. madagascariensis*  extract (200 and 400 mg/kg) showed protection from myocardial injury evidenced by decreased myocytes degeneration as well as edema and minimal inflammation (Figure 3C, D).

## **DISCUSSION**

The purpose of this work was to evaluate the potential cardioprotective role of *H. madagascariensis* aqueous extract stem bark aqueous extract (AEHM) in isoproterenol-induced myocardial damage model in rats. ISO in high doses induces morphological and functional alterations in the heart which closely resembles local myocardial infarction-like pathological changes seen in human myocardial infarction**[28](#page-7-12)**. It has been reported that auto-oxidation of excess catecholamines such as ISO results in free radical mediated peroxidation of membrane phospholipids and consequently leading to permeability changes in the myocardial membrane, intracellular calcium overload and irreversible damages**[29](#page-7-13)** .





Values are given as mean $\pm$ SEM (n = 7). \*\**p*<0.01 and \*\*\**p*<0.001 as compared to the control group; \*\**p*<0.01 and \*\*\**p*<0.01 as compared to the isoproterenol (ISO)-treated group. HM 200 or HM 400: *H. madagascariensis* aqueous extract (200 or 400 mg/kg) ; Vit E: vitamin E (100 mg/kg).

Electrocardiogram (ECG) is considered the most important clinical tool for the diagnosis of myocardial infarction**[30](#page-7-14)**. In the present study, subcutaneous injection of isoproterenol (85 mg/kg) for two consecutive days caused ST- segment elevation and Ramplitude depression. The elevated ST-segment reflects the potential difference in the boundary between ischemic and non-ischemic zones and a consequent loss of cell membrane function and the depressed R-amplitude might be due to the isoproterenol-induced myocardial edema**[31](#page-7-15)** . *H. madagascariensis* (200 mg/kg) pre-treatment as well as vitamin E markedly inhibited isoproterenol-induced Ramplitude depression and amended the ST-segment elevation, indicating its protective effects on cell membrane function and electrical discharges.

In the present study, we have observed a significant increase in the heart weight and the ratio of heart weight to body weight in ISO-treated rats. The observed increase in the heart weight in ISO-induced rats might be due to the increased water content, edematous intramuscular space and extensive necrosis of cardiac muscle fibers followed by the invasion of damaged tissues by the inflammatory cells **[31](#page-7-15)[,32](#page-7-16)**. Pretreatment with the plant extract or vitamin E did not modify this increase. These results suggest that AEHM does not affect the gain or loss of weight of this organ. However, the short duration of the preventive treatment (seven days) could be responsible for the observed result. It would be wise to consider a longer duration in future experiments to better elucidate the effects of the plant extract on this parameter.

Myocardium contains many diagnostic marker enzymes like lactate dehydrogenase (LDH), aspartate aminotransferase (AST) and alanine aminotransferase (ALT). Upon administration of isoproterenol, the oxygen demand of the heart increases with increase in ionotropic effect in the heart, resulting in prolonged ischemia and glucose deprivation. The cells are damaged with increased muscle contractility, which results in increasing the cell membranes permeability

allowing cardiac enzymes to leak out into the bloodstream**[33](#page-7-17)**. Increased activities of these marker enzymes in the serum are indicative of cellular damage and loss of functional integrity**[34](#page-7-18)**. In the present study, the significant increase observed in the activities of LDH, AST, ALT in the serum of ISO-induced rats may be due to the leakage of them from the heart as a result of necrosis induced by ISO. The aqueous extract of *H. madagascariensis* seems to preserve the structural and functional integrity and/or permeability of the cardiac membrane and thus restricting the leakage of these indicative enzymes from the myocardium, as evident from the markedly blunted levels of these enzymes in the extract pre-treated groups when compared to the ISO-treatment group, thereby establishing the cardioprotective effect of the aqueous extract of *H. madagascariensis.* Malondialdehyde (MDA), a product of the reaction of polyunsaturated fatty acids with reactive oxygen species, is a biomarker of oxidative stress. Since the major constituents of biological membranes are lipids, their peroxidation can lead to cell damage and death<sup>35</sup>. The concentration of MDA increases in response to the free radical production in myocardial infarction, and decreases by antioxidant systems**[35](#page-7-19)** . Increased formation of MDA of 49% as compared to the control group is an indication of the severity of the cellular injury to the heart induced by ISO and this can be linked with altered membrane structure and enzyme inactivation<sup>[36](#page-7-20)</sup>. . *H. madagascariensis* administration reduced MDA levels in a dose-dependent manner by 35 and 42% respectively at 200 and 400 mg/kg.

Glutathione (GSH) is a tripeptide which has a direct antioxidant function by reacting with superoxide radicals, peroxy radicals and singlet oxygen followed by the formation of oxidized GSH and other disulfides. It plays an important role in the regulation of variety of cell functions and in cell protection against free radical mediated injury**[37,](#page-7-21)[38](#page-7-22)**. Depressed GSH levels may be associated with an enhanced protective mechanism to oxidative stress in myocardial infarction. In this study,

ISO administration was found to reduce the levels of GSH. This observation concurs with several earlier findings**[29,](#page-7-13)[32,](#page-7-16)[35](#page-7-19)**. Pre-treatment with *H. madagascariensis*  (400 mg/kg) significantly improved the level of GSH. This points to the potential antioxidant and free radical scavenging activity of *H. madagascariensis*. In previous studies, *H. madagascariensis* has been described as an antioxidant and free radical scavenger**[24,](#page-7-8)[39](#page-7-23)**. The current study shows the antioxidant activity of *H. madagascariensis* and endorses its cardioprotective effect mediated by its antioxidant effect in myocardium. Histopathological examination of myocardial tissue in the control rats illustrated clear integrity of the myocardial cell when compared to the hearts of ISO treated rats. ISO-induced rats showed separations of cardiac muscle fibers, edema and extensive infiltration of neutrophil granulocytes. Pretreatment with the aqueous extract of *H. madagascariensis* (200 and 400 mg/kg) considerably attenuated the edema, reduced inflammatory cell infiltration and preserved normal cardiac muscle fibers structure, further confirming the cardioprotective effect of *H. madagascariensis*.

#### **CONCLUSIONS**

In conclusion, our study reveals that pre-treatment of rats with the aqueous extract of *H. madagascariensis* exerts a remarkable protective potential against damages caused by isoproterenol-induced myocardial infarction. This cardioprotective effect could be associated with the enhancement of antioxidant defense, reduction of lipid peroxydation and is confirmed by amending electrocardiographic pattern, improvement of cardiac markers and less histopathological damage following isoproterenol-induced myocardial infarction. Although this study has provided a possible new therapeutic tool for myocardial infarction, more studies are required to elucidate the precise mechanism of *H. madagascariensis* in reversing the pathogenesis of myocardial infarction.

#### **ACKNOWLEDGMENTS**

The authors would like to thank the International Foundation for Science (IFS) for the research grant No. F/5882-1 awarded to Dr. Ngo Lemba Tom Esther.

#### **AUTHOR'S CONTRIBUTION**

**Tom ENL:** writing original draft, conceptualization, methodology, investigation. **Nankia FD:** Writing, review, and editing, supervision. **Nyunaï N:** writing, review, and editing. **Girard-Thernier C:** writing, review, and editing. **Girard-Thernier C:**  methodology, investigation, formal analysis. **Demougeot C:** conceptualization, methodology, investigation. **Dimo T:** data curation, writing. The final manuscript was read and approved by all authors.

#### **DATA AVAILABILITY**

The datasets generated during this study are available from the corresponding author upon reasonable request.

## **CONFLICT OF INTEREST**

No conflict of interest associated with this work.

### **REFERENCES**

- <span id="page-6-0"></span>1. WHO media Center. Cardiovascular diseases (CVDs) Fact sheet. Updated May 2017. *<http://www.who.int/mediacentre/factsheets/fs317/en/>*
- <span id="page-6-1"></span>2. Kumar JS, Menon VP. Changes in levels of lipid peroxides and activity of superoxide dismutase and catalase in diabetes associated with myocardial infarction. Indian J Exp Biol 1992; 30(2):122-127. PMID: 1521861
- <span id="page-6-2"></span>3. Opie LH. The heart: Physiology, from cell to circulation. Philadelphia: Lippincott-Raven 1998; 3rd ed, 637.
- <span id="page-6-3"></span>4. Rahmathulla MSB, Lakshmi KD. Origination and development of isoproterenol-induced myocardial infarction in male Wistar rats. Int Res J Pharm 2013; 4(5):26-35. *<https://doi.org/10.7897/2230-8407.04508>*
- <span id="page-6-4"></span>5. Song L, Jiang W, Qing Y, Hu X, Li Y, Tong QY, Wu XH. The antagonistic effect of PI3K-gamma inhibitor AS605240 on cardiac hypertrophy and cardiac fibrosis induced by isoproterenol in rats. Sichuan Da Xue Xue Bao Yi Xueban 2011; 42(4): 471–474. PMID: 21866628
- <span id="page-6-5"></span>6. Nichtova Z, Novotova M, Kralova E, Stankovicova T. Morphological and functional characteristics of models of experimental myocardial injury induced by isoproterenol. Gen Physiol Biophys 2012; 31(2): 141–151. *[https://doi.org/10.4149/gpb\\_2012\\_015](https://doi.org/10.4149/gpb_2012_015)*
- <span id="page-6-6"></span>7. Verma S, Maitland A, Weisel RD, Li SH, Fedak PW, Pomroy NC, Mickle DA, Li RK, Ko L, Rao V. Hyperglycemia exaggerates ischemia-reperfusion-induced cardiomyocyte injury: reversal with endothelin antagonism. J Thorac Cardiovasc Surg 2002; 123(6):1120-1124. *<https://doi.org/10.1067/mtc.2002.121973>*
- <span id="page-6-7"></span>8. Moens AL, Claeys MJ, Wuyts FL, Goovaerts I, Van Hertbruggen E, Wendelen LC, Van Hoof VO, Vrints CJ. Effect of folic acid on endothelial function following acute myocardial infarction. Am J Cardiol 2007; 99(4):476- 481. *<https://doi.org/10.1016/j.amjcard.2006.08.057>*
- <span id="page-6-8"></span>9. Rajadurai M, Prince PSM. Comparative effects of Aegle marmelos extract and alpha-tocopherol on serum lipids, lipid peroxides and cardiac enzyme levels in rats with isoproterenol-induced myocardial infarction. Singapore Med J 2005; 46(2):78-81. PMID: 15678289
- <span id="page-6-9"></span>10. Xiong X, Borrelli F, de Sá Ferreira A, Ashfaq T, Feng B. Herbal medicines for cardiovascular diseases. Evid Based Complement Alternat Med. 2014, DOI: [10.1155/2014/809741](https://dx.doi.org/10.1155%2F2014%2F809741) *<https://doi.org/10.1155/2013/149363>*
- <span id="page-6-10"></span>11. Ojha S, Golechha M, Kumari S, Arya DS. Protective effect of Emblica officinalis (amla) on isoproterenol-induced cardiotoxicity in rats. Toxicol Ind Health 2011; 28(5) 399– 411. *<https://doi.org/10.1177/0748233711413798>*
- <span id="page-6-11"></span>12. Burkill, HM. The useful plants of west tropical Africa. 1985; Vol 2.
- 13. Tona L, Kambu K, Ngimbi N, Cimanga K, Vlietinck AJ. Antiamoebic and phytochemical screening of some Congolese medicinal plants. J Ethnopharmacol 1998; 61: 57– 65. *[https://doi.org/10.1016/S0378-8741\(98\)00015-4](https://doi.org/10.1016/S0378-8741(98)00015-4)*
- *14.* EME. Committee for Veterinary Medical Product *H. madagascariensis*. The European Agency for the Evaluation of Medicinal Products, 1999. *<https://doi.org/10.14302/issn.2328-0182.japst-18-2341>*
- 15. Erah PO, Asonye CC, Okhamafe AO. Response of Trypanosoma brucei-induced anaemia to a commercial herbal preparation. Afr J Biotech 2003; 2(9): 307–311.
- <span id="page-7-0"></span>16. Kamanzi AK, Schmid C, Brun R, Kone MW, Traore D. Antitrypanosomal and antiplasmodial activity of medicinal plants from Cote d'Ivoire. J Ethnopharmacol 2004; 90(2-3): 221–227. *<https://doi.org/10.1016/j.jep.2003.09.032>*
- <span id="page-7-1"></span>17. Adeneye AA, Olagunju JA, Elias SO, Olatunbosun DO, Mustafa AO, Adeshile OI, Ashaolu AO, Laoye TA, Bamigboye AO, Adeoye AO. *Harungana madagascariensis* in acute and repeated acetaminophen hepatotoxic rats. Int J Appl Res Na Prod 2008; 1(3): 29-42.
- <span id="page-7-2"></span>18. Nicolas JP. Plantes médicinales du nord de Madagascar ethnobotanique antakarana et informations scientifiques. Jardins du Monde 2012; 295: 134-135.
- <span id="page-7-3"></span>19. Koné MW, Kamanzi AK. Inventaire ethnomédical et évaluation de l'activité anthelminthique des plantes médicinales utilisées en Côte d'Ivoire contre les helminthiases intestinales. Pharm Méd Trad Afr 2006; 14:55- 72.
- <span id="page-7-4"></span>20. Lenta NB, Ngouela S, Boyom FF, Tantangmo F, Tchouya FGR, Tsamo E, Gut J, Rosenthal PJ, Connolly JD. Antiplasmodial activity of some constituents of the root bark of *Harungana madagascariensis* Lam. (Hypericaceae). Chem Pharmacol Bull 2007; 55(3):464-447. *<https://doi.org/10.1248/cpb.55.464>*
- <span id="page-7-5"></span>21. Mangambu M. Contribution à l'étude phytochimique de quelques plantes médicinales antidiabétiques de la ville de Bukavu et ses environs (Sud-Kivu, R. D. Congo). J Appl Biosci 2014; 75: 6211-6220. *<https://doi.org/10.4314/jab.v75i1.7>*
- <span id="page-7-6"></span>22. Etchiké CA, Aristide Sassa AM, Abba A, Nyonbourg E. Evaluation in vitro de l'activité antibactérienne de cinq plantes de la pharmacopée traditionnelle de l'Adamaoua (Cameroun) Cameroon J Exp Biol 2011, 7(1): 22-27. *<https://doi.org/10.4314/cajeb.v7i1.69788>*
- <span id="page-7-7"></span>23. Nwodo OFC. Antibiotic and anti-inflammatory analgesic activities of *Harungana madagascariensis* stem bark. Int J Crude Drug Res 1989; 27(3): 137–140. *<https://doi.org/10.3109/13880208909053953>*
- <span id="page-7-8"></span>24. Antia BS, Ita BN, Udo UE. Nutrient composition and in vitro antioxidant properties of *Harungana madagascariensis* stem bark extracts. J Med Food 2015; 18(5):609-614. *<https://doi.org/10.1089/jmf.2014.0084>*
- <span id="page-7-9"></span>25. Kouam SF, Yapna DB, Krohn K, Ngadjui BT, Ngoupayo J, Choudhary MI, Schulz B. Antimicrobial prenylated anthracene derivatives from the leaves of *Harungana madagascariensis*. J Nat Prod 2007; 70(4): 600–603. *<https://doi.org/10.1021/np060556l>*
- <span id="page-7-10"></span>26. Wilbur KM, Bernheim F, Shapiro OW. Determination of lipid peroxidation. Arch Biochem Biophysics 1949; 24(2): 305–310.
- <span id="page-7-11"></span>27. Ellman GL. Tissue sulfhydryl group. Arch Biochem Biophysics 1959; 82(1): 70-77.
- <span id="page-7-12"></span>*[https://doi.org/10.1016/0003-9861\(59\)90090-6](https://doi.org/10.1016/0003-9861(59)90090-6)* 28. Rona G. Catecholamine cardiotoxicity. J Mol Cell Cardiol 17(4):291–300.

*[https://doi.org/10.1016/s0022-2828\(85\)80130-9](https://doi.org/10.1016/s0022-2828(85)80130-9)*

- <span id="page-7-13"></span>29. Li H, Xie YH, Yang Q, Wang SW, Zhang BL, Wang JB, Cao W, Bi LL, Sun JY, Miao S. Cardioprotective effect of paeonol and danshensu combination on isoproterenolinduced myocardial injury in rats. PLoS One 2012; 7(11): e48872. *<https://doi.org/10.1371/journal.pone.0048872>*
- <span id="page-7-14"></span>30. Bahit MC, Criger DA, Ohman EM, Granger CB, Wagner GS. Thresholds for the electrocardiographic change range of biochemical markers of acute myocardial infarction (GUSTO-IIa data). Am J Cardiol 2002; 90(3):233–237. *[https://doi.org/10.1016/S0002-9149\(02\)02460-8](https://doi.org/10.1016/S0002-9149(02)02460-8)*
- <span id="page-7-15"></span>31. Khorrami A, Mojtaba H, Mehraveh G, Nasrin MD, Alireza G. Tacrolimus ameliorates functional disturbances and oxidative stress in isoproterenol-induced myocardial infarction. DARU J Pharm Sci 2014; 22(1):68. *<https://doi.org/10.1186/s40199-014-0068-3>*
- <span id="page-7-16"></span>32. Patel V, Upaganlawar A, Zalawadia R, Balaraman R. Cardioprotective effect of melatonin against isoproterenol induced myocardial infarction in rats: A biochemical, electrocardiographic and histoarchitectural evaluation. Eur J Pharmacol 2010; 644(1-3):160–168. *<https://doi.org/10.1016/j.ejphar.2010.06.065>*
- <span id="page-7-17"></span>33. Wang SB, Tian S, Yang F, Yang HG, Yang XY, Du GH. Cardioprotective effect of salvianolic acid on isoproterenolinduced myocardial infarction in rats. Eur J Pharmacol 2009; 615(1-3): 125–132.

*<https://doi.org/10.1016/j.ejphar.2009.04.061>*

- <span id="page-7-18"></span>34. Sabeena FKH, Anandan R, Kumar SH, Shiny KS, Sankar TV, Thankappan TK. Effect of squalene on tissue defense system in isoproterenol-induced myocardial infarction in rats. Pharmacol Res 2004; 50(3):231–236. *<https://doi.org/10.1016/j.phrs.2004.03.004>*
- <span id="page-7-19"></span>35. Tappel AL, Dillard CJ. *In vivo* lipid peroxidation: measurement via exhaled pentane and protection by vitamin E. Fed Proc 1981; 40(2):174–178. PMID: 7461141
- <span id="page-7-20"></span>36. Rathore N, John S, Kale M, Bhatnagar D. Lipid peroxidation and antioxidant enzymes in isoproterenol induced oxidative stress in rat tissues. Pharmacol Res 1998; 38(4): 297–303. *<https://doi.org/10.1006/phrs.1998.0365>*
- <span id="page-7-21"></span>37. Wattanapitayakul SK, Bauer JA. Oxidative pathways in cardiovascular disease roles, mechanisms, and therapeutic implications. Pharmacol Therapeut 2001; 89:187–206. *[https://doi.org/10.1016/S0163-7258\(00\)00114-5](https://doi.org/10.1016/S0163-7258(00)00114-5)*
- <span id="page-7-22"></span>38. Wu J, Hecker JG, Chiamvimonvat N. Antioxidant enzyme gene transfer for ischemic diseases. Adv Drug Deliver Rev 2009; 61(4):351–363. *<https://doi.org/10.1016/j.addr.2009.01.005>*
- <span id="page-7-23"></span>39. Iwalewa EO, Adewale IO, Taiwo BJ, Arogundade T, Osinowo A, Daniyan OM, Adetogun GE. Effects of *Harungana madagascariensis* stem bark extract on the antioxidant markers in alloxan induced diabetic and carrageenan induced inflammatory disorders in rats. J Compl Integr Med 2008; 5:1.

*<https://doi.org/10.2202/1553-3840.1088>*