

Available online at www.ujpronline.com Universal Journal of Pharmaceutical Research An International Peer Reviewed Journal ISSN: 2831-5235 (Print); 2456-8058 (Electronic)

Copyright©2023; The Author(s): This is an open-access article distributed under the terms of the CC BY-NC 4.0 which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited



RESEARCH ARTICLE

EFFICACY OF DRACAENA CINNABARI AS TOOTH WHITENING NATURAL PRODUCT: A SPECTROPHOTOMETRIC ANALYSIS

Wedad Omar Ali Elaiwa¹, Ibrahim Zaid Al-Shami¹, Mohsen AL-Hamzi¹, Abdul Wahab Al-kholani¹, Nagwa Othman¹, Mokhtar Abd Hafiz Abd Alhamid Al-Ghorafi²

¹Department of Conservative Dentistry, Faculty of Dentistry, Sana'a University, Republic of Yemen. ²Department of Pharmaceutical Organic Chemistry, Faculty of Pharmacy, Sana'a University, Republic of Yemen.

Article Info:

Abstract



Article History: Received: 4 April 2023 Reviewed: 11 May 2023 Accepted: 23 June 2023 Published: 15 July 2023

Cite this article:

Elaiwa WOA, Shami IZA, Hamzi MA, Kholani AWA, Ghorafi MAAA, Othman N. Efficacy of *Dracaena cinnabari* as tooth whitening natural product: A spectrophotometric Analysis. Universal Journal of Pharmaceutical Research 2023; 8(3):19-23.

https://doi.org/10.22270/ujpr.v8i3.945

*Address for Correspondence:

Dr. Mokhtar AbdHafiz Abd Alhamid Al-Ghorafi., Department of Pharmaceutical Organic Chemistry, Faculty of Pharmacy, Sana'a University, Republic of Yemen. Tel: +967770010749.

E-mail: mok.alghorafi@su.edu.ye

Objective: The aim of this study was to determine the efficacy of a homemade tooth-whitening recipe using *Dracaena cinnabari* (DC) resin, which is traditionally used for tooth decay treatment and cleaning in Soqatra. The study investigated the antioxidant activity of the resin extracts in methanol as tooth-whitening natural product.

Materials and Methods: A total of 40 bovine teeth with initial color B2 were selected and divided into four groups. One group was kept as a negative control, while the other three were stained with Yemeni coffee solution for one week. The first group was treated with 10% carbamide peroxide (CP) home bleaching, the second group was treated with 10% DC gel with rubbing movement (DC Ru), and the third group was treated with 10% DC gel without rubbing (DC) for 6 hours daily for 14 days. Color measurements were taken at different time points using a spectrophotometer device.

Results: All experimental gels resulted in greater color change compared to the negative control, with DC Ru showing the greatest ΔE^* value (p<0.001) compared to the DC group (p<0.006) and the CP group (p<0.001). The second reading of stabilization of all gels resulted with resembled ΔE^* values to the first reading of stabilization.

Conclusion: The study suggests that the experimental gels containing phenolic content with strong antioxidant effects may reserve significant clinical potential as active agents for tooth-whitening without using HP/CP. Further studies are needed to measure the effect on surface roughness and color stability.

Keywords: *Dracaena cinnabari*, natural product, spectrophotometric analysis, tooth whitening.

INTRODUCTION

Bleaching is One of the most aesthetic procedures in dentistry and a more conservative choice for vital teeth whitening than other procedures like ceramic laminates, it is a well-established technique, and it applied a high level of effectiveness^{1,2}. The homebleaching procedure with 10% CP gel has been considered the safest method for bleaching teeth accepted by the American Dental Association (ADA) with minimal adverse effects, however, since this tooth-bleaching modality is characterized by patientapplied, but there is a risk of gel application overexposed dentine in patients with gingival recession and abrasion lesions. Moreover, the incorrect use of the tray may result in gel overflow, with extended softtissue exposure which causes gingival irritation and material ingestion³ Herbal and natural products have

been used in dental practice for years and have become more common these days due to their biocompatibility, high antimicrobial activity, antioxidant, and antiinflammatory properties⁴. *Dracena cinnabari* (*D. cinnabari*) resin approved its effectiveness as antimicrobial, antiviral⁵, antioxidant⁶, and antiinflammatory⁷.

According to our knowledge, none of the studies has steadily investigated the effects of *D. cinnabari* resin extract as a natural bleaching product. Therefore, the present study, which is aimed to highlight the potential possibilities of new material of a bleaching agent, evaluates its antioxidant effect and compares it with other commercial bleaching materials.

MATERIALS AND METHODS

Plant Material

The *D. cinnabari* plant was collected in its natural habitat on Soqotra Island. The botanical name of this endemic wild tree is *D. cinnabari* Balf. f. (Dracaenaceae). The English common name for both the tree and its resin is dragon's blood. The Arabic name "Dam Alakhwin" means "Brother's blood" and is also used for both the tree and its resin. The Soqotri resin (dragon's blood = Dam Alakhwin) is a high-quality, pure red blood resin that is known on the island as "Emzoloh." It is collected from the incision of the young stem bark of the female tree. This standard pure resin can be described as an authentic superior Soqotri resin⁹.

Collection of plants:

The resin of \hat{D} . *cinnabari* was collected from a young fresh stem female tree on Soqatra Island, Yemen. Dragon's blood resin was purchased from a wholesale supplier of traditional Unani medicine in March 2022. The plant samples were identified and authenticated by the Environmental Protection Authority of Yemen and have been deposited at the herbarium of the Department of Pharmacognosy, Faculty of Pharmacy, Sana'a University, Yemen.

Preparation of *D. cinnabari* resin methanol extract:

The powdered resin of *D. cinnabari* (50g) was extracted with methanol in a 1:10 ratio. The mixture was shaken at room temperature for 3 days, then sonicated at 45° C for 30 minutes to enhance the extraction. The methanol was then separated from the extract using a rotary evaporator under reduced pressure at 40° C, resulting in a gummy red resin extract^{8,9}.

Preparation of *D. cinnabari* Gel as tooth bleaching agent

The gel composition was presented in Table 1. The sodium saccharin and the EDTA were previously dissolved in water and the methyl paraben and the menthol in ethanol. The carbomer 940 was dispersed in water with the glycerin and the pH was adjusted to 7, In this study, the stock solution was prepared by using ethanol, it was found to be less toxic than the other solvents, hence it was safer, and favored to be used for preparing the bleaching gel, it contained 10% of resin extract.

Preparation of specimens

Forty freshly bovine incisors devoid of intrinsic stains, cracks, and fractures were selected with their initial color equal to B2, and had their roots sectioning using a carborundum wheel disk (Maxman, China), The dental pulp tissue was removed from the crown portions using an #80 K-file (Mani, Tokyo, Japan) The teeth received prophylaxis with Robinson brush (Escova, USA), with water coolant until flat surfaces were obtained¹⁰. The prepared specimens from extraction until use and during the treatment process the teeth were kept in Distilled water at 37°C¹¹.

Artificial staining procedure

The teeth were artificially stained to mimic natural staining in the mouth. The specimens were numbered from 1 to 40 and divided into two groups: 10 teeth were soaked in distilled water and 30 teeth were soaked in Yemeni coffee solution for one week. The solution was replaced daily. After seven days, the teeth were

rinsed with water for 10 seconds and polished with a rubber cup and pumice stone/water solution to remove any undesirable external staining. The teeth were then fixed in a dense silicon paste (Zhermack, Italy) mold on a wooden plate to standardize the measurement angle¹².

Bleaching procedure

Before bleaching, the specimens were cleaned with an ultrasonic cleanser for 90 seconds. Since there was no standard application protocol for the experimental groups (D. cinnabari and D. cinnabari Rubbing), both materials were applied according to the manufacturer's instructions for home bleaching with carbamide peroxide (CP). After baseline measurements, all specimens were randomly assigned to four groups (n =10) based on the bleaching gel used. The first group was treated with 10% CP Home bleaching (Whiteness Perfect 10% FGM, Brazil) (positive group) for 6 hours daily was bleaching gel was applied 1-mm a thick layer with a brush to the middle third of facial surfaces of the teeth, the gel then removed and the surface cleaned using gauze soaked in distilled water and then the specimen's surfaces were washed and dried with absorbent paper. Samples were not air-dried with any system that could dehydrate the sample, after that stored in distilled water for the remaining time (18 h). This process was repeated each day for 14 days to simulate recommended home use, in addition, While the gel is in contact with the specimen, the latter is kept in a wet environment placing it over a cotton pellet embedded in distilled water. At the end of 14-day bleaching procedure. All the specimens removed from the bleaching gel, cleaned under running water and then stored in distilled water for 24 h, Then three measurements at the middle third of the facial surface of the specimen taken as previously described for the The group baseline measurement. second (experimental groups=20) was treated with 10% D. cinnabari gel and divided into two subgroups (n=10). One group was applied by rubbing movement (D. cinnabari RU) on the central portion of the buccal surfaces and the specimen was brushed with hand pressure on an electrical brush (power flex- Jordan) for one minute. Third group applied without rubbing (D. cinnabari) only applied 1 mm of a thick layer of gel with the brush for 6 h daily as the same application of the positive group (CP) for 14 days.

The fourth group (negative group) did not receive any bleach treatments and was treated in the same manner as experimental groups in the rinsing and drying by drops of distilled water are applied to the facial surface. All of the procedure's steps are carried out by the same operator (specimen preparation, staining, and bleaching). For color reading, a Spectrophotometer device was employed (Easy Shade, VITA Zahnfabrik, Bad Säckingen, Germany), and the specimen was placed over a black background, Three color readings were performed for each sample at identical positions using the CIE L* a* b* coordinates and the mean of three color readings was considered the color value on the L*, a*, and b* axis, at baseline (T0), one-day poststaining (T1), 7-days post-staining (T2), immediately after bleaching (T3), 7 days post-bleaching, (T4) and 14 days post-bleaching (T5).

Statistical Analysis

The data was collected and analyzed using IBM SPSS Version 25. The descriptive statistics were presented as means and standard deviations (SD) for L*, a*, and b*. A one-way ANOVA was used to statistically analyze the data. Repeated measures ANOVA was used to evaluate the effect of time on delta score (color change). Multiple comparisons of means were performed using the t-test with *p*-value adjustment, which was set at the significance level of $p \le 0.05\%$. An independent t-test was performed to identify significantly different group means when the ANOVA test was significant.

RESULTS AND DISCUSSION

The duration of immersion time in coffee statistically significantly (p < 0.05) increased the change in the color for experimental materials, DC Ru specimens were statistically significantly (p < 0.002) more stable in color than (CP and DC groups) with *p*-value (.054 and .038) respectively after 1 week of immersion in coffee. While in the DW group, there was an insignificant difference (0.263) after 1 week of immersion in DW as shown in (Table 2). According to the statistical analysis with comparison as shown in (Figure 2), after one day of immersion in the coffee solution, DC Ru specimens discolored markedly from (1.74) to (3.44) after one week of immersion as same as CP specimens changing from (2.6) to (5.1) after one week of immersion, however, DC specimens in coffee showed slightly more color changes (3.73) compared to (2.57) after one day of immersion in the coffee. Figure 1 and Figure 2 shows the mean values and SD of all experimental groups according to the staining solution. In this study,

 ΔE values for the experimental groups were determined for DC Ru, CP, and DC specimens (7.09, 6.7, 4.6) respectively.

Table 1	:	Formulation	of	the	D.	cinnab	ari	gel
---------	---	-------------	----	-----	----	--------	-----	-----

Component	Amount
Dragon's blood extract	10%
Carbomer 940	5%
Glycerin	3%
Sodium saccharin	1%
Methyl paraben	0.2%
EDTA	0.1%
Menthol	0.02%
Purified Water	81.5%

Based on the perceptibility thresholds defined in the literature, the overall color difference between the experimental groups is considered moderately perceivable. All gels resulted in greater color change when compared to the negative group, the highest value occurred at DC Ru gel with (p < 0.001), than the positive group (CP) demonstrated greater ΔE^* values (p < 0.001) and after that DC experimental gels with (p < 0.006) when compared to negative group DW in Table 2, Table 3 and Table 4). The main purpose of this study was to evaluate the effect of D. cinnabari used by the population to obtain tooth whitening, the resin is widely used in traditional folk medicine in Yemen for the treatment of dental diseases, and it is readily widely spread in the market with low cost. Furthermore, it has been found to have several pharmacological properties such as antimicrobial, antioxidant, anti-inflammatory, cytotoxicity and antitumor activities¹³. The D. cinnabari resin extract was prepared in this study by using absolute methanol (100%) as a solvent.

	GROU	Р	Ν	Mean	SD	T-test	<i>p</i> -value
СР	Coffee	One Day	10	2.656130	2.9361182	2.215	0.054
		One Week	10	5.112230	4.4636632		
DC	Coffee	One Day	10	2.572570	1.9435405	2.437	0.038
		One Week	10	3.735810	2.2208448		
DC Ru	Coffee	One Day	10	1.744360	1.2360171	4.344	0.002
		One Week	10	3.448320	1.5729662		
DW		One Day	10	5.369750	3.4682066	1.193	0.263
	DW	One Week	10	6.453140	3.1471311		

Table 2: Mean values and SD of all experimental groups according to the staining solution.

N = Number of specimens, SD = Standard Deviation, CP = Carbamide peroxide, DC = D. *cinnabari* without rubbing, DC Ru = D. *cinnabari* with rubbing, DW = Distilled water



Figure 1: Mean color changes (ΔE^*) of the different experimental groups over time.

Group		Ν	Mean	SD				
	T1_T0	10	4.565030	3.9272792				
	T2_T0	10	5.369750	3.4682066				
CP	T3_T0	10	6.751030	2.8149650				
	T4_T0	10	6.121770	2.7157788				
	T5_T0	10	6.190390	3.0087140				
	T1_T0	10	2.572570	1.9435405				
	T2_T0	10	3.206900	1.6220585				
DC	T3_T0	10	4.647570	1.2595474				
	T4_T0	10	4.266320	1.2653632				
	T5_T0	10	3.532910	1.2188543				
	T1_T0	10	4.744360	3.2360171				
	T2_T0	10	4.426730	3.1394743				
DC Ru	T3_T0	10	7.093210	1.4872108				
	T4_T0	10	6.684370	1.5232603				
	T5_T0	10	5.893280	1.3320478				
	T1_T0	10	2.654330	2.9367385				
	T2_T0	10	2.567670	1.6738546				
DW	T3_T0	10	3.395460	3.0504153				
	T4_T0	10	4.063950	3.6182684				
	T5_T0	10	4.991270	4.1195389				

Table 3: Multiple comparisons of mean color change (ΔE values) between all experimental groups and DW by using an independent t-test.

CP = Carbamide peroxide, DC = D. cinnabari without rubbing, DC Ru = D. cinnabari with rubbing, DW = Distilled water, N = Number of specimens, SD = Standard Deviation, T0 = baseline, T1 = after 24 hours post-staining, T2 = 7 days post-staining, T3 = immediately after bleaching, T4 = 7 days post-bleaching, T5 = 14 days post-bleaching.



Figure 2: Mean color changes (ΔE^*) of the different experimental groups over time.

Although there is no previous report of a natural bleaching agent containing *D. cinnabari*. The first null hypothesis was rejected because both experimental gels (DC and DC RU) groups were effective on tooth whitening. Considering *D. cinnabari* gel with the strong antioxidant activity of its high phenolic content⁹, they can break down macromolecules/stains into smaller parts, thus increasing light reflection from the tooth surface, thereby increasing the lightness and thereby resulting in a whitening effect. While the

second null hypothesis was true (No differences in color change between *D. cinnabari* and CP), regarding to the results, which demonstrated (DCRU) experimental gels produced color changes as same as the positive group (CP), this stain removal effect could be due to the mechanical rubbing of *D. cinnabari*.

Similar to this study Toothpastes containing activated charcoal showed better effectiveness in whitening teeth than toothpastes containing blue covarine and hydrogen peroxide¹⁶.

 Table 4: Multiple comparisons of mean color change (ΔE values) between all experimental groups and DW by using an independent t-test.

	Group	Ν	Mean	SD	T-test	<i>p</i> -value			
	СР	10	6.751030	2.8149650	4.039	0.001			
	DW	10	2.567670	1.6738546					
	DC	10	4.647570	1.2595474	3.140	0.006			
13-10	DW	10	2.567670	1.6738546					
	DC Ru	10	7.093210	1.4872108	6.391	0.000			
	DW	10	2.567670	1.6738546					

T0 = baseline, T3 = immediately after bleaching, CP = Carbamide peroxide, DC = D. cinnabari without rubbing, DC Ru = D. cinnabari with rubbing, DW = Distilled water, N = Number of specimens, SD = Standard Deviation.

Miswak, a natural product that is similar to the current study, has been shown to have antioxidant properties^{17,18}. *Salvadora persica* paste, which is made from miswak, has the potential to be an effective, alternative teeth-whitening product, especially for removing extrinsic staining¹⁹.

Limitation of the study

The present study tested only one concentration of the natural material used in this research is the same as commercial materials with 10% and the experimental gels were applied according to the directions for positive control. There was no artificial saliva to mimic the oral environment.

CONCLUSIONS

Within the limitations of this study, it could be concluded that successfully obtained *D. cinnabari* gel extract with 10 % concentration using methanol as a solvent. The results showed that coffee was a statistically significant effect on the pigmentation of the teeth. Specimens treated with *D. cinnabari* gel and its effectiveness in whitening the teeth, especially *D. cinnabari* RU specimen by rubbing, as is popularly known were significantly as same as affect those treated with Carbamid Peroxide due to the experimental gels (*D. cinnabari*) containing phenolic content with strong antioxidant effects.

CONFLICT OF INTEREST

No conflict of interest is associated with this work.

ACKNOWLEDGEMENTS

Authors are thankful to all who contributed to the completion of this work and especially Department of Conservative Dentistry, Faculty of Dentistry, Sana'a University.

DATA AVAILABILITY

Data will be made available on request.

AUTHOR'S CONTRIBUTION

Elaiwa WOA: preparation of specimens, bleaching procedure study design. Shami IZA: data collection, data analysis. Hamzi MA: editing, revision. kholani AWA: collection of plants. Ghorafi MAAA: field works, thesis writing. Othman N: writing, review, and editing, data curation.

REFERENCES

- 1. Meireles SS, Heckmann SS, Leida FL, *et al.* Efficacy and safety of 10% and 16% carbamide peroxide tooth-whitening gels: a randomized clinical trial. Operative Dent 2008; 33(6), 606–612. *https://doi.org/10.2341/07-150*
- Ribeiro JS, de Oliveira da Rosa WL, da Silva AF, Piva E, Lund RG. (2020). Efficacy of natural, peroxide-free toothbleaching agents: A systematic review, meta-analysis, and

technological prospecting. Phytoth Res: PTR 2020; 34(5), 1060–1070. https://doi.org/10.1002/ptr.6590

- Soares, DG, Basso FG, Pontes EC, et al. Effective toothbleaching protocols capable of reducing H (2) O (2) diffusion through enamel and dentine. J Dent 2014; 42(3), 351–358. https://doi.org/10.1016/j.jdent.2013.09.001
- Parham S, Kharazi AZ, Bakhsheshi-Rad HR, et al. 2020. Antioxidant, antimicrobial, and antiviral properties of herbal materials. Antioxidants 2020; 9(12), 1309. https://doi.org/10.3390/antiox9121309
- Mothana RAA, Mentel R, Reiss C, Lindequist U. Phytochemical screening and antiviral activity of some medicinal plants from the island Soqotra. Phytother Res 2006; 20(4):298–302. https://doi.org/10.1002/ptr.1858
- Juránek I, Suchý V, Stará D, Mašterova I, Grancaiová Z. Antioxidative activity of homoisoflavonoids from *Muscari* racemosum and D. cinnabari. Pharmazie 1993; 48(4):
- Alwashli A, Alaoui K, Al-Sobarry M. Anti-inflammatory and Analgesic effects of ethanolic extract of *D. cinnabri* Balf as endemic plant in Yemen. Int J Pharma Bio Sci 2012; 3(2): 96–106.
- Al-Afifi N, Alabsi A, Kaid F, Bakri M, Ramanathan A. (2019). Prevention of oral carcinogenesis in rats by *Dracaena cinnabari* resin extracts. Clinical oral investigations 2019; 23(5):2287–2301. https://doi.org/10.1007/s00784-018-2685-6
- Al-Fatimi M. (2018). Ethnobotanical survey of Dracaena cinnabari and investigation of the pharmacognostical properties, antifungal and antioxidant activity of its resin. Plants (Basel, Switzerland) 2018; 7(4):91. https://doi.org/10.3390/plants7040091
- Ablal MA, Adeyemi AA, Jarad FD. The whitening effect of chlorine dioxide- An *in vitro* study. J Dentistry 2013; 41, e76-e81.https://doi.org/10.1016/j.jdent.2013.05.006
- 11. Vieira GF, Arakaki Y, Caneppele TMF. 2008. Spectrophotometric assessment of the effects of 10% carbamide peroxide on enamel translucency. Brazilian Oral Res 2008; 22, 90-95. https://doi.org/10.1590/s1806-83242008000100016
- Haruyama A, Kojima M, Kameyama A, Muramatsu T. Combined use of baking soda and electric toothbrushing for removal of artificial extrinsic stain on enamel surface: An *in vitro* study. J Clin Exp Dent 2022; 14(1):e 9. https://doi.org/10.4317/jced.58708
- Al-Awthan YS, Bahattab OS. Phytochemistry and pharmacological activities of *D. cinnabari* resin. BioMed Res Int 2021; 1-7. https://doi.org/10.1155/2021/8561696
- 14. Gupta D, Gupta RK. Bio protective properties of Dragon's blood resin: *in vitro* evaluation of antioxidant activity and antimicrobial activity. BMC Comp Alt Med 2011; 11:13. *https://doi.org/10.1186/1472-6882-11-13*
- Annegowda HV, Bhat R, Min-Tze L, Karim AA, Mansor SM. Influence of sonication treatments and extraction solvents on the phenolics and antioxidants in star fruits. J Food Sci Tech 2012; 49(4): 510–514. https://doi.org/10.1007/s13197-011-0435-8
- 16. Aydin N, Karaoglanoglu S, Oktay EA, Ersöz B. Determination of the whitening effect of toothpaste on human teeth. Odovtos-Int J Dental Sci 2022; 24(1):67-75. https://doi.org/10.15517/ijds.2021.46376
- Nordin A, Saim AB, Ramli R, Hamid AA, Nasri NWM, Idrus RBH., 2020. Miswak and oral health: An evidencebased review. Saudi J Biol Sci 2020; 27(7):1801-1810. https://doi.org/10.1016/j.sjbs.2020.05.020
- Niazi F, Naseem M, Khurshid Z, Zafar MS, Almas K. Role of *Salvadora persica* chewing stick (miswak): A natural toothbrush for holistic oral health. European J Dentis 2016; 10(02): 301-308.https://doi.org/10.4103/1305-7456.178297
- Halib N, Nuairy NB, Ramli H, Ahmad I, Othman NK, Salleh SM, Bakarudin SB. 2017. Preliminary assessment of *Salvadora persica* whitening effects on extracted stained teeth. J Appl Pharm Sci 2017; 7(12):121-125. https://doi.org/10.7324/JAPS.2017.71217